

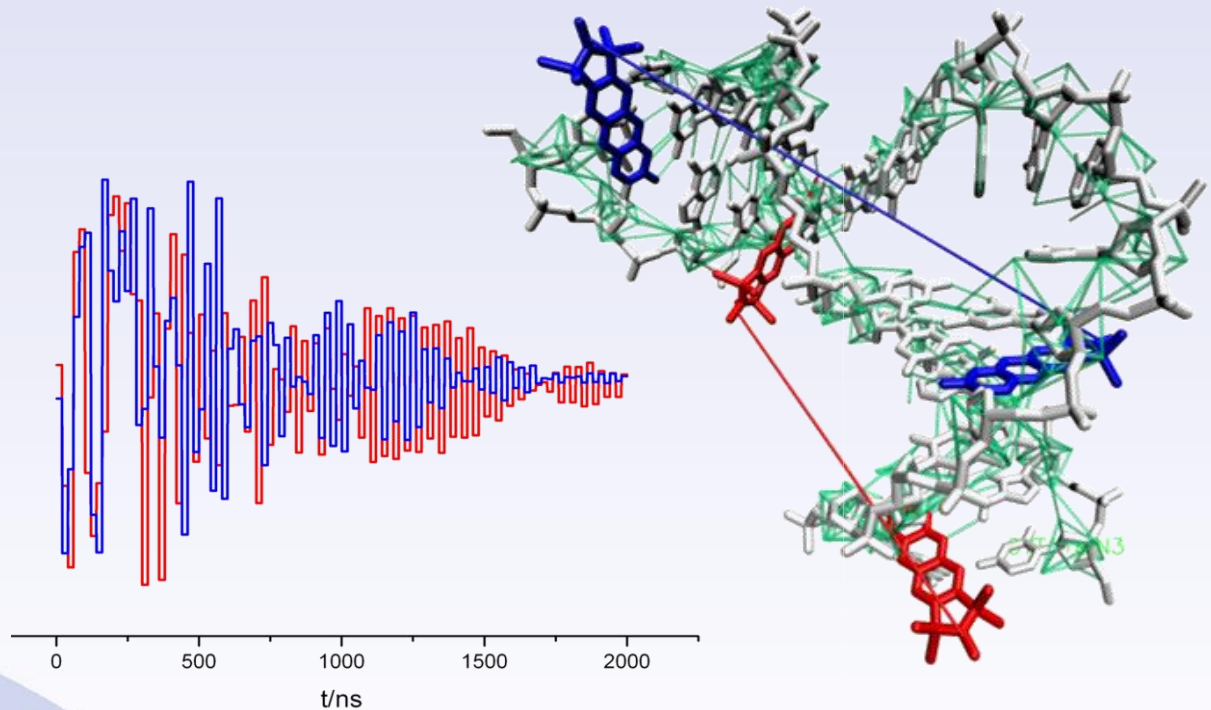
Structure and Dynamics of

Nucleic Acids

T. F. Prisner

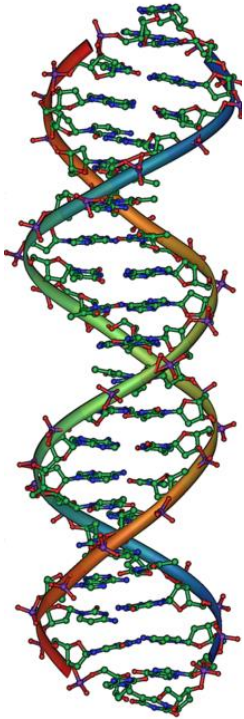
Institute of Physical & Theoretical Chemistry
Center of Biological Magnetic Resonance
Goethe University Frankfurt

www.uni-frankfurt.de



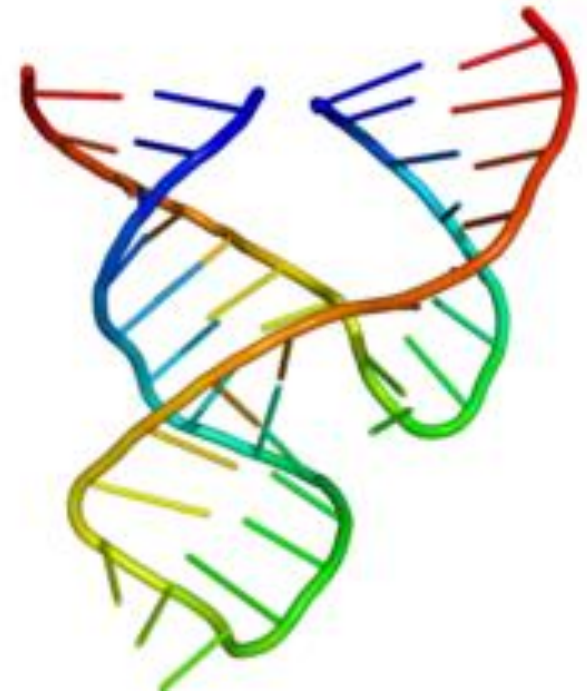
Structure and Flexibility of Nucleic Acids

Highly conserved structures for storage and translation of the genetic code



X-ray

Highly flexible structures for regulatory and catalytic functions of nucleic acids

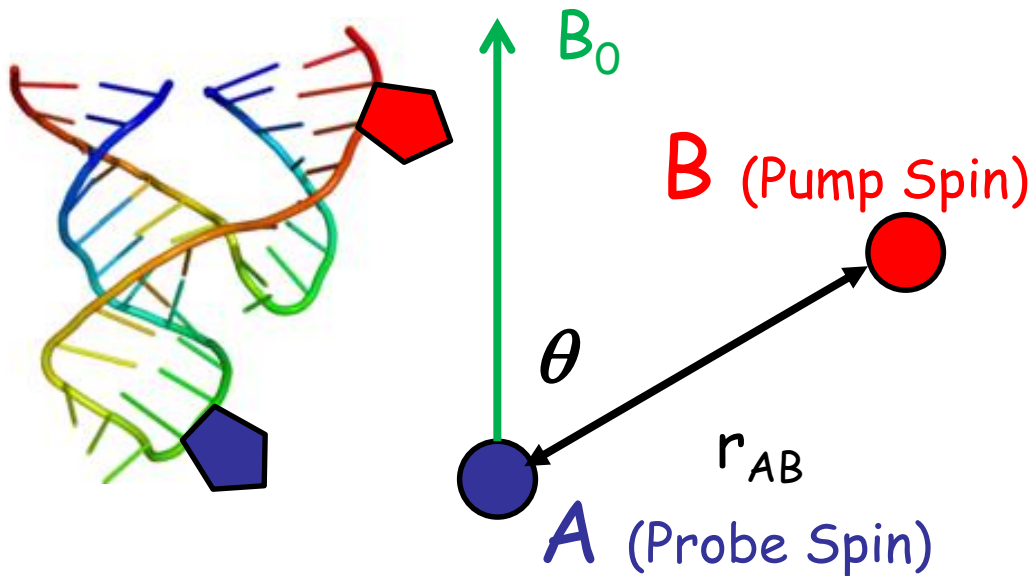


NMR, CD, IR
FRET, MD

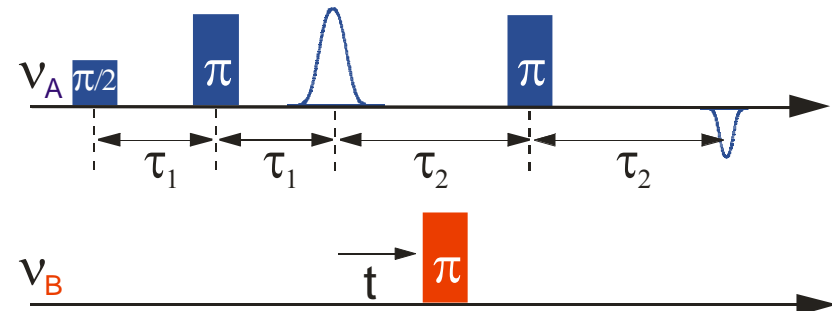
EPR



Pulsed Electron Electron Double Resonance



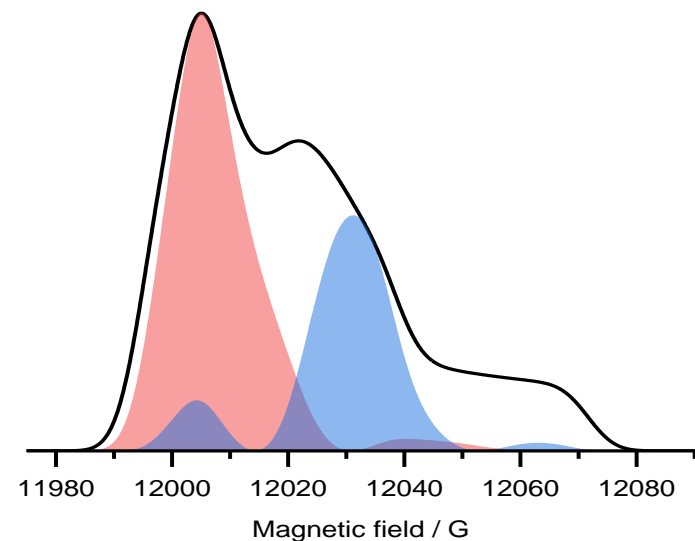
A. D. Milov, K. M. Salikov, M. D. Shirov.
Fiz. Tverd. Tela, **23**, 975 (1981)



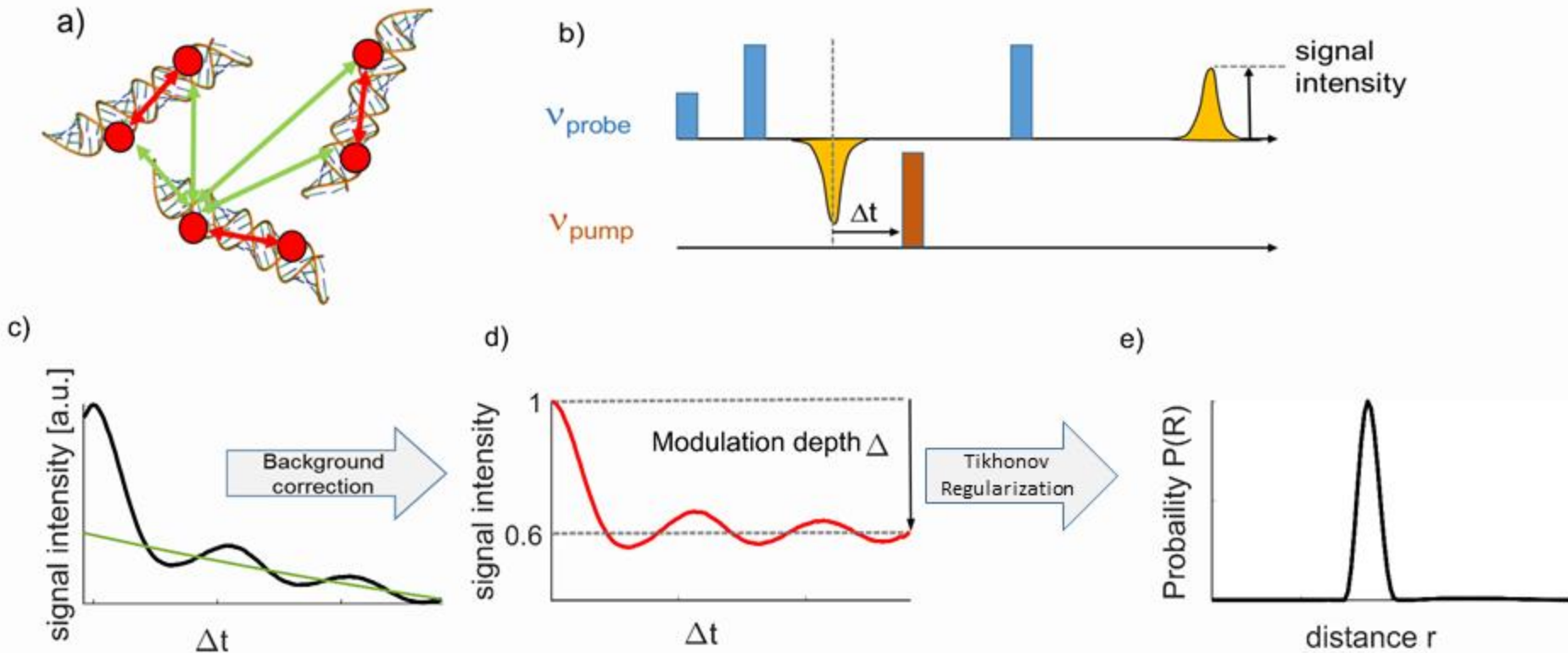
G. Jeschke, H.W. Spiess et al.,
JMR, **142**, 331 (2000)

$$\omega_{ee} = \frac{2\pi \cdot C}{r^3} (1 - 3 \cos^2 \theta)$$

$$C = 52.04 \text{ MHz} \cdot \text{nm}^3$$

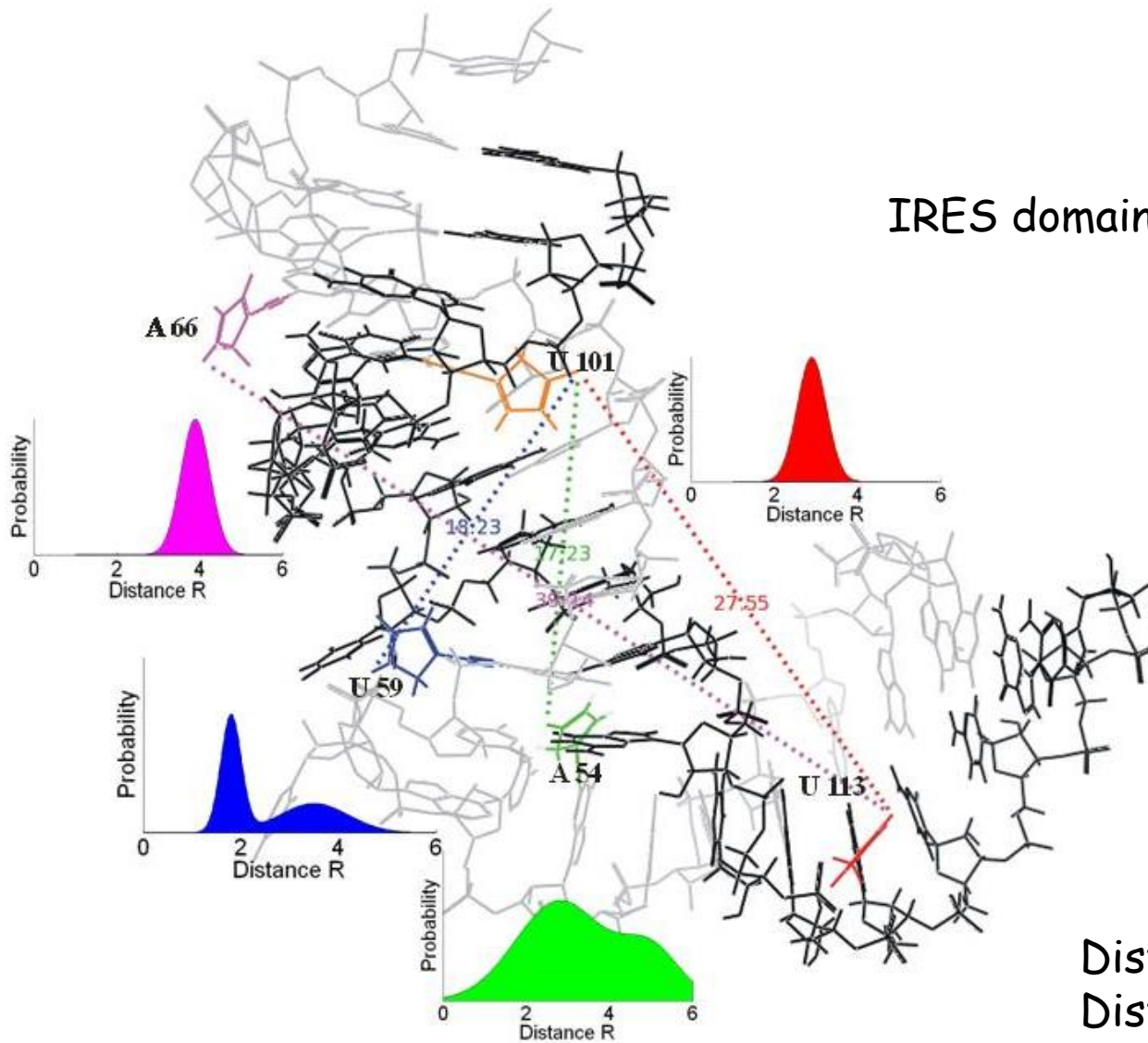


Analysis of PELDOR/DEER time traces



Distances and Distance Distributions

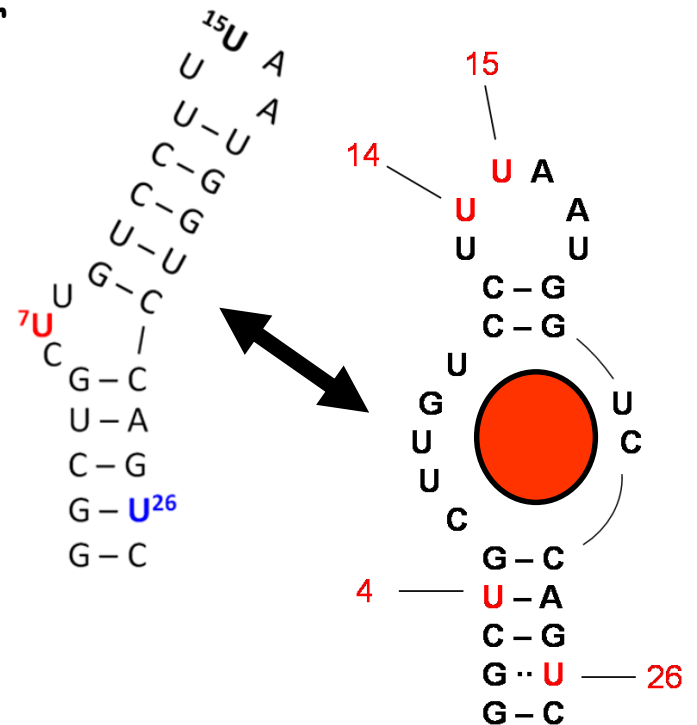
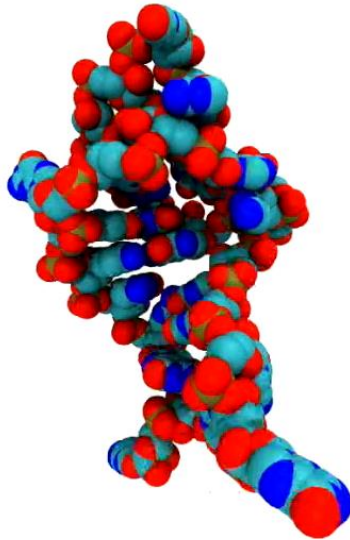
IRES domain of Hep Virus



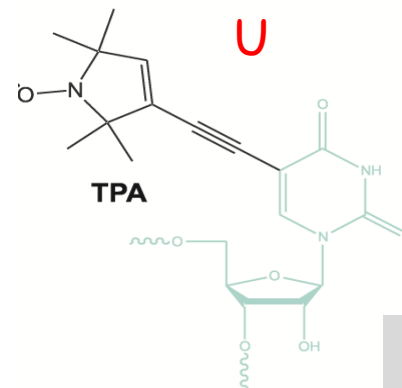
Distances &
Distance Distributions

Conformational dynamics of RNA

Neomycin Binding Aptamer



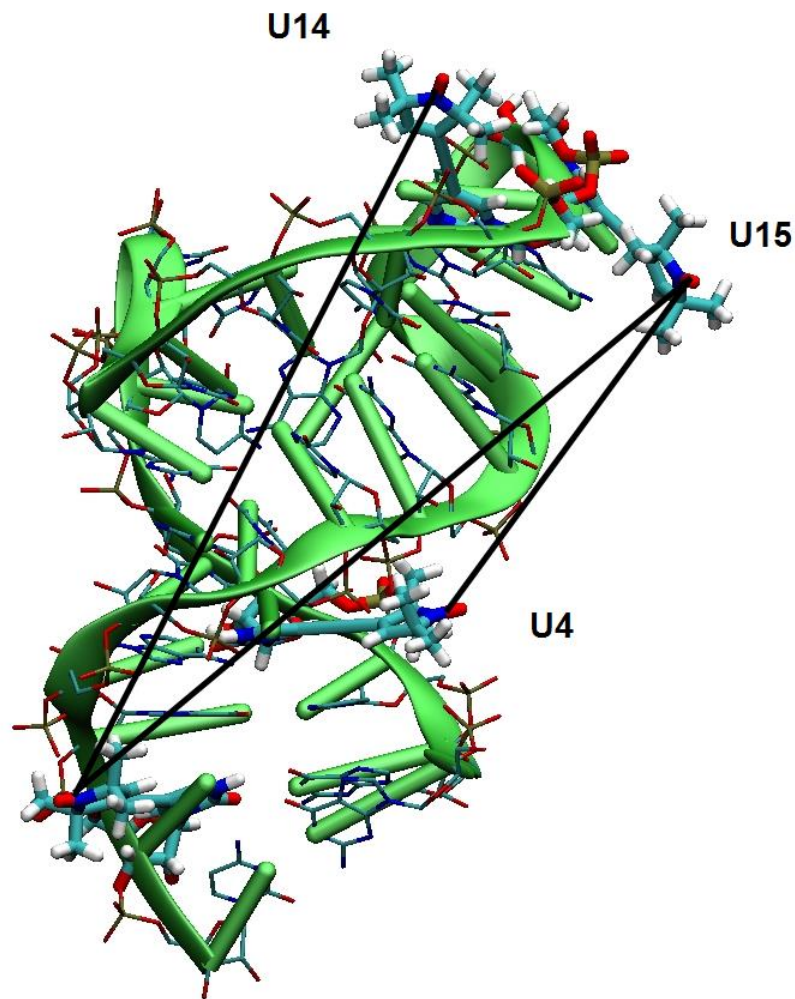
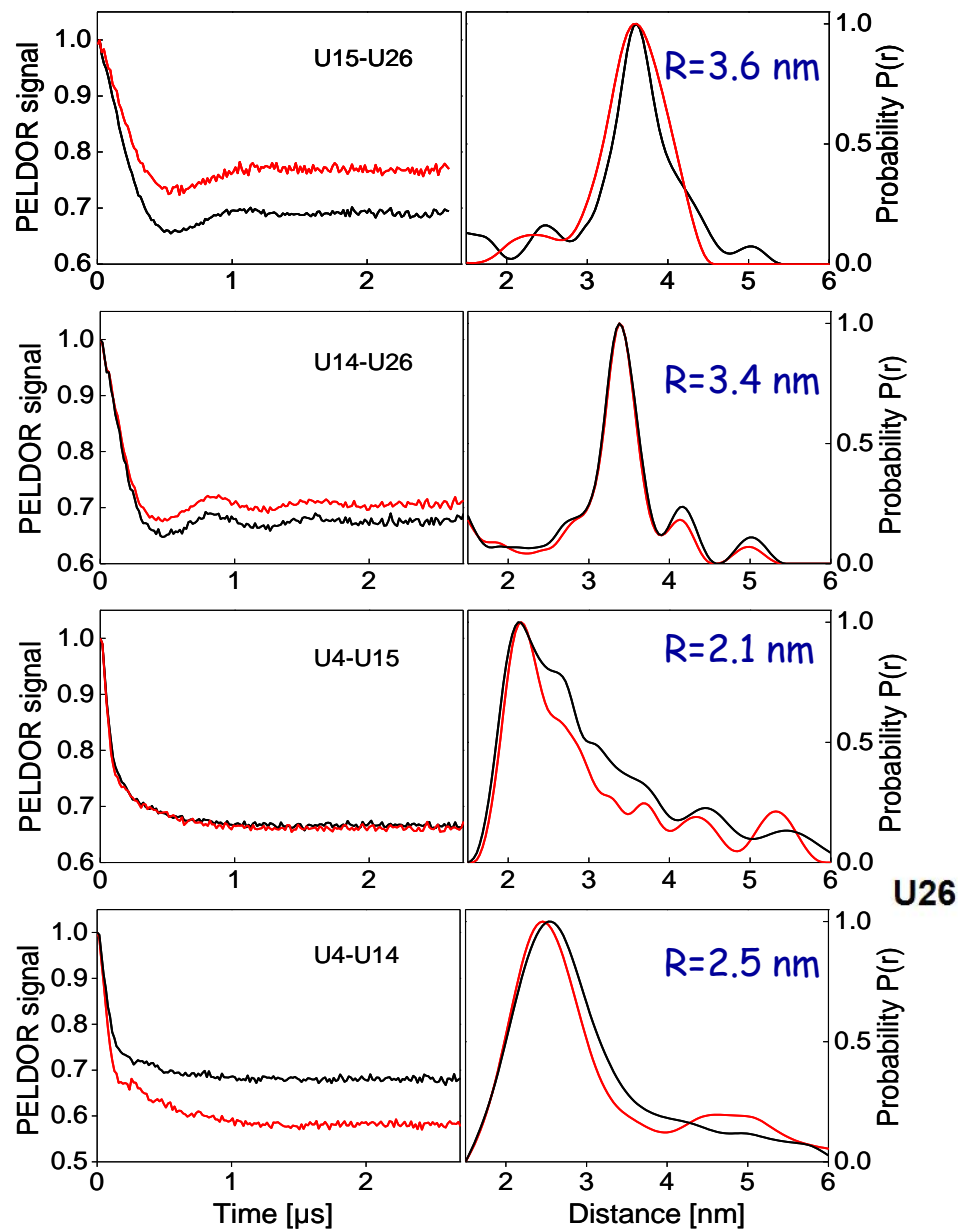
L. Stelzl, G. Hummer et al.
J. Chem. Theory Comput. **2017**, *13*, 3927, 6328



Synthesized by
J. Engels
Org. Chem.
Uni. Frankfurt

Nature Protocoll (2007)
NAR (2007)

Conformational dynamics of RNA

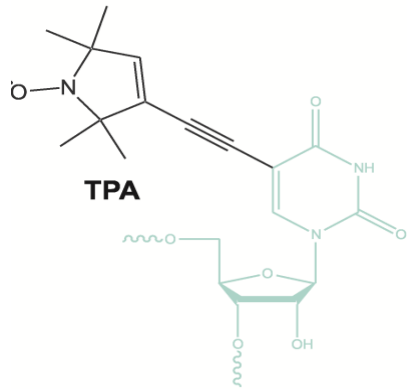


Wöhnert, J. et al *Angewandte* 2011

JACS (2011)

Spin labels for Nucleic Acids

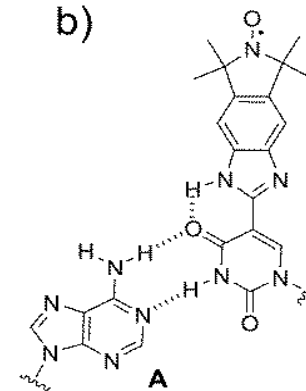
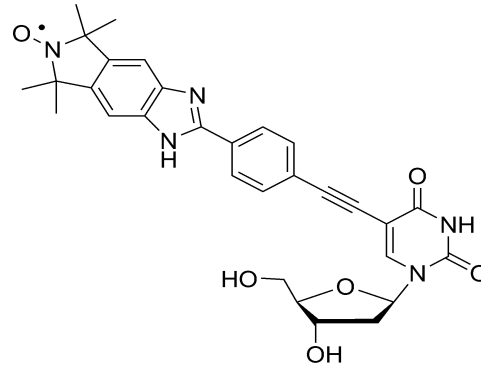
Joachim Engels
(Uni Frankfurt)



Rotate around triple bond

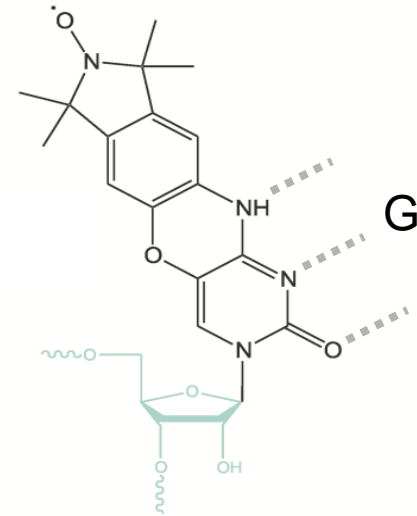
Nature Protocoll (2007)
NAR (2007)

Snorri Sigurdsson (University of Iceland)



Rigid in double stranded structures

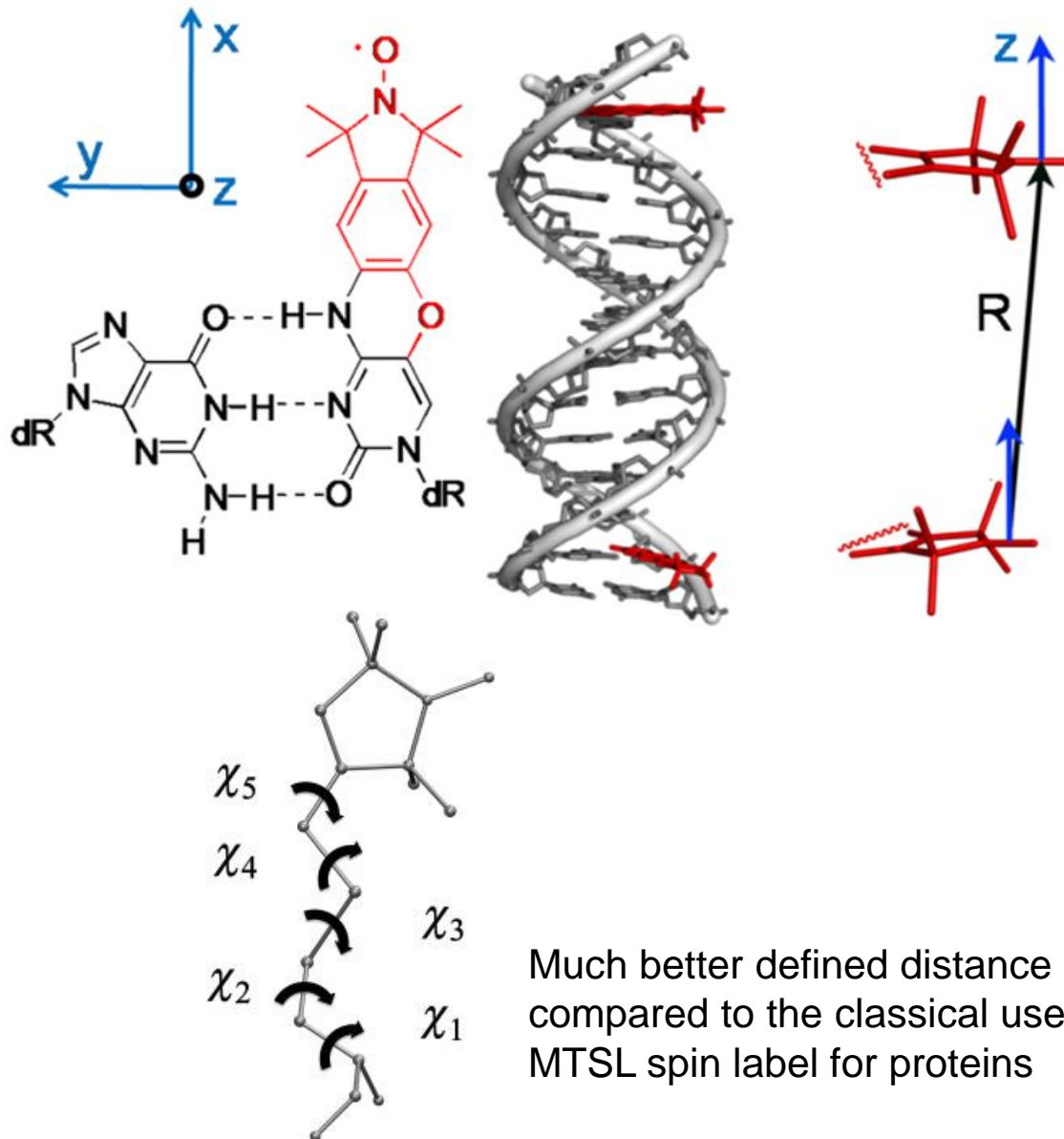
Chem. Eur. J. (2014)



Angewandte (2009)
JACS (2011)

- All nucleotides in DNA and RNA can be spin-labeled
- Different degrees of rotational flexibility of spin label
- Different orientation with respect to nucleotide
- Synthesis with modified nucleobase

A rigid spin-label analog of cytidine



Distance between two unpaired electron spins can be directly converted in structural information of nucleic acid molecule

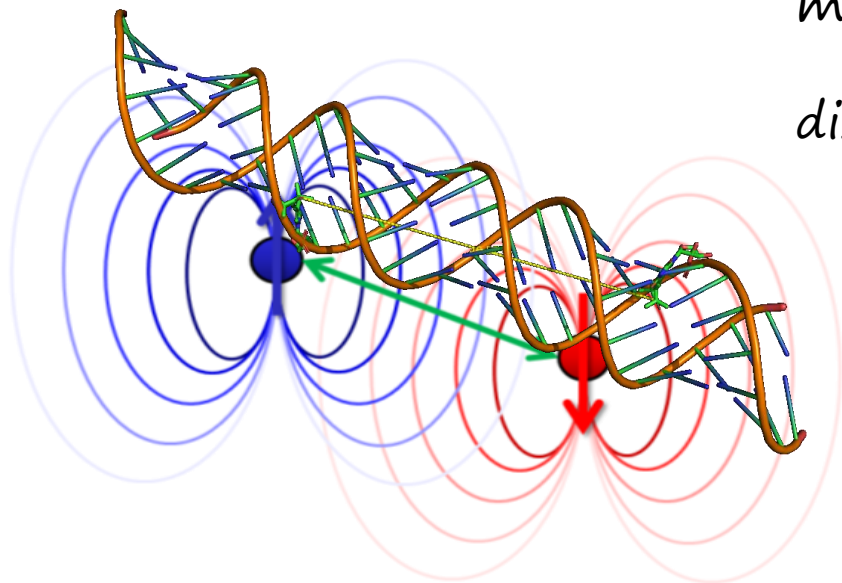
Snorri Sigurdsson
(University of Iceland)

Much better defined distance compared to the classical used MTSL spin label for proteins

Distance measurements of spin-labeled NA

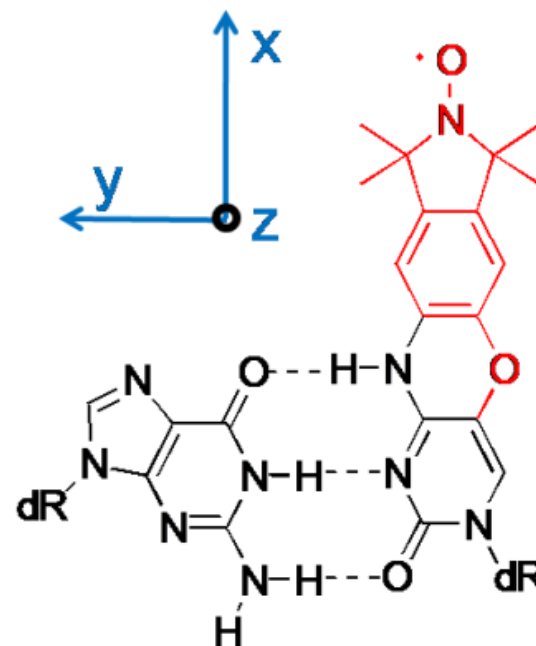
Magnetic dipole-dipole interaction
measured by pulsed EPR

distances of $\sim 1-10$ nm

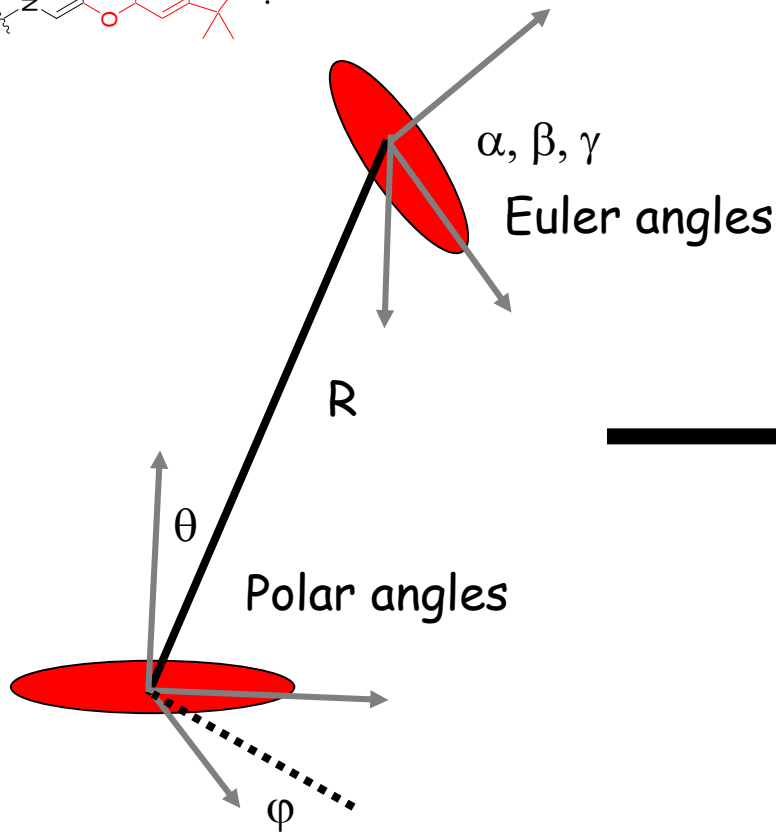
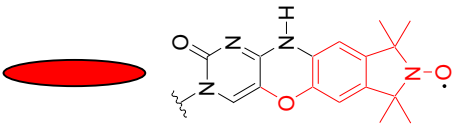


Cytidine analog spin label
rigidly incorporated into
ds DNA or ds RNA

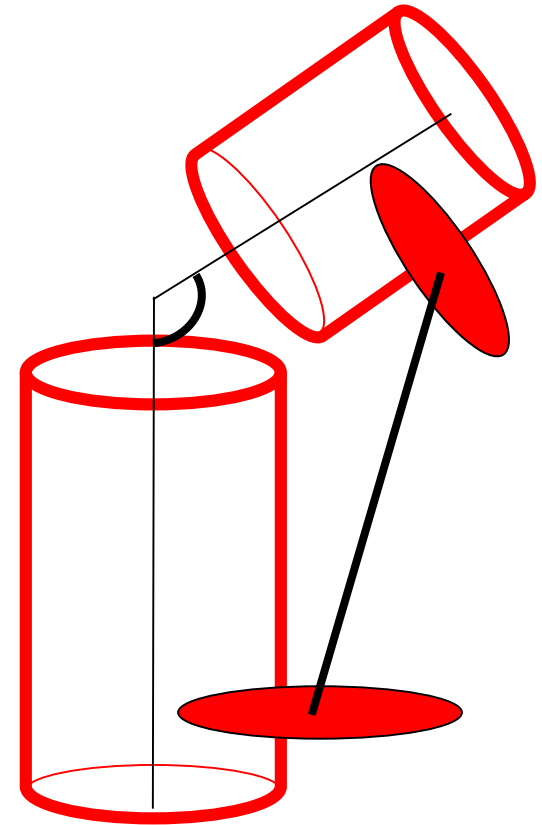
Collaboration with Snorri Sigurdsson
(Uni. of Iceland)



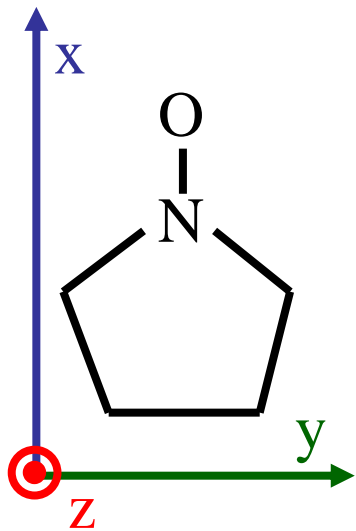
Investigation of conformational flexibility of dsDNA



Determination of distance R and geometry of spin labels



Relates directly to structure of helical parts in RNA / DNA



^{14}N ($I=1$)
 3 hyperfine lines
 (red, blue,
 green)

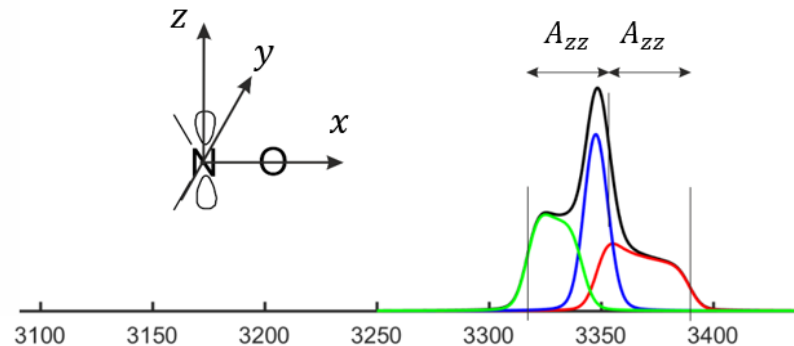
Anisotropic g -
 tensor from
 spin-orbit
 coupling
 (depending
 strongly on
 field)

X-band (9 GHz)
 0.3 T

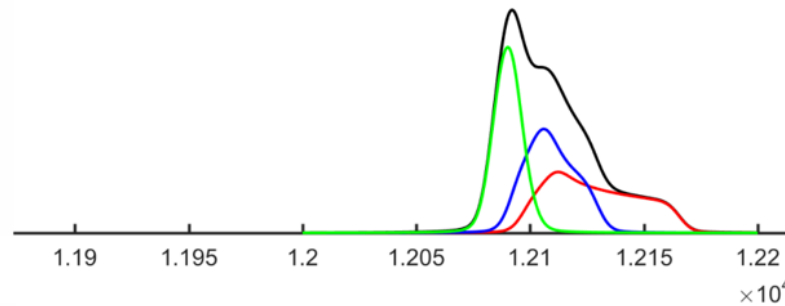
Q-band (35 GHz)
 1.2 T

G-band (180) GHz)
 6.4 T

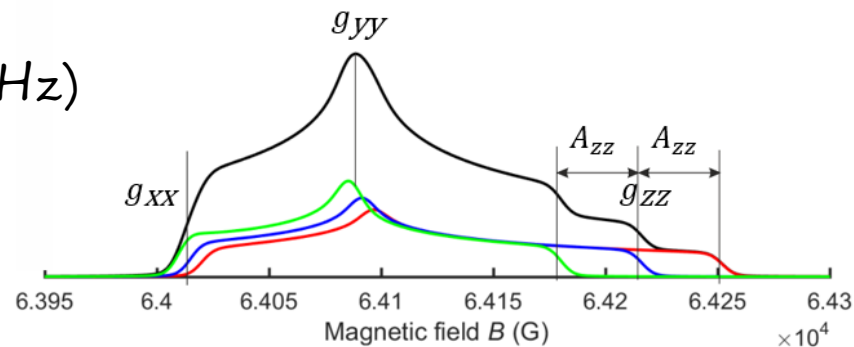
A)

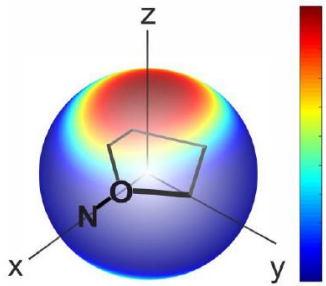


B)



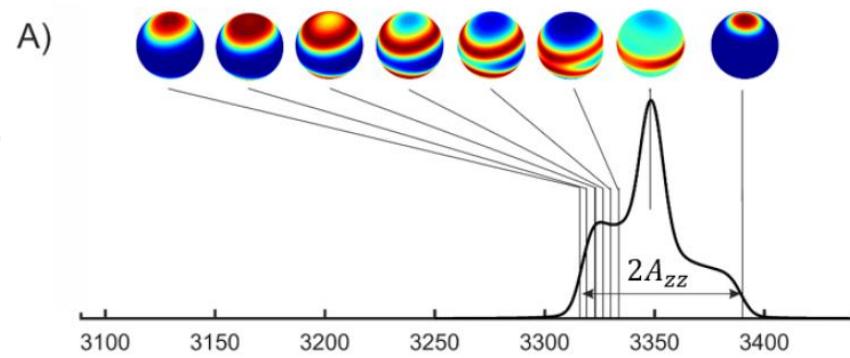
C)



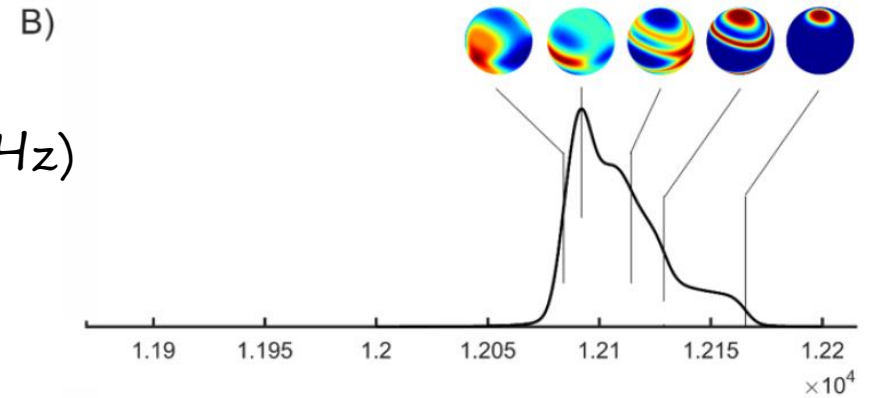


Selection of specific molecular orientations with respect to the magnetic field by frequency of MW pulses

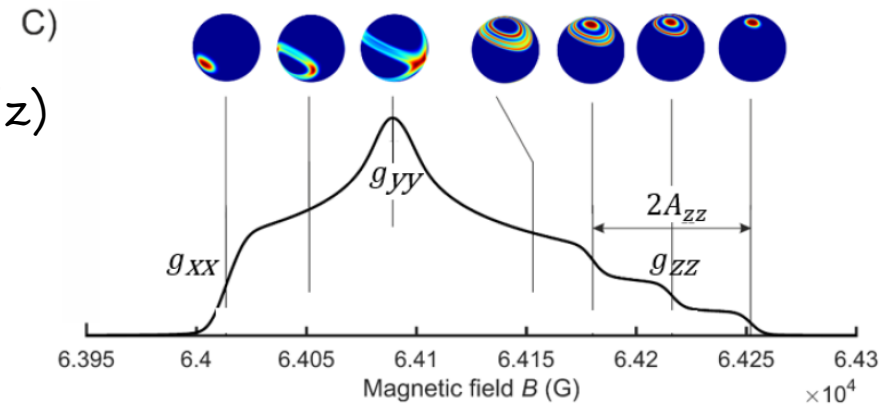
X-band (9 GHz)



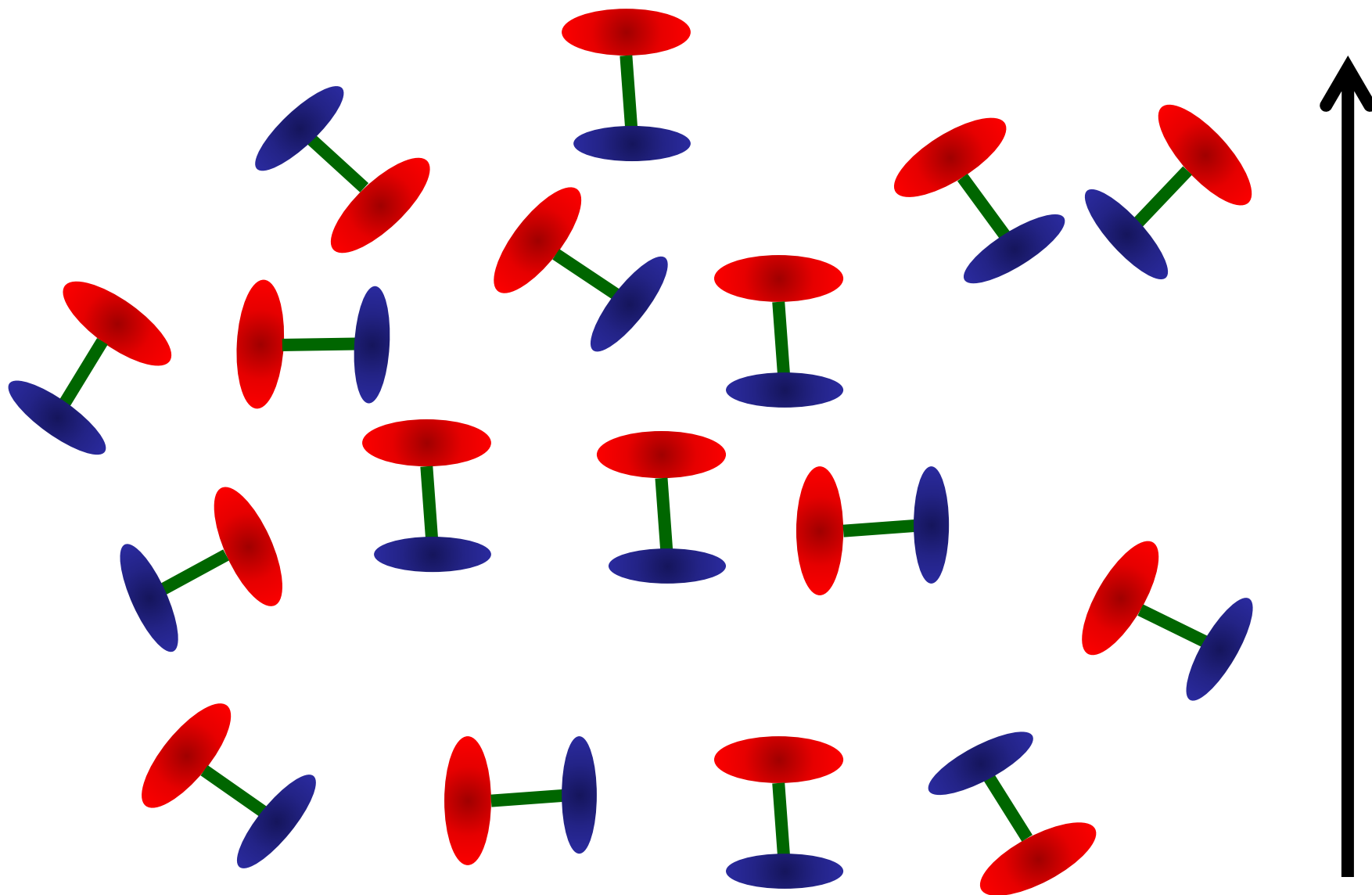
Q-band (35 GHz)



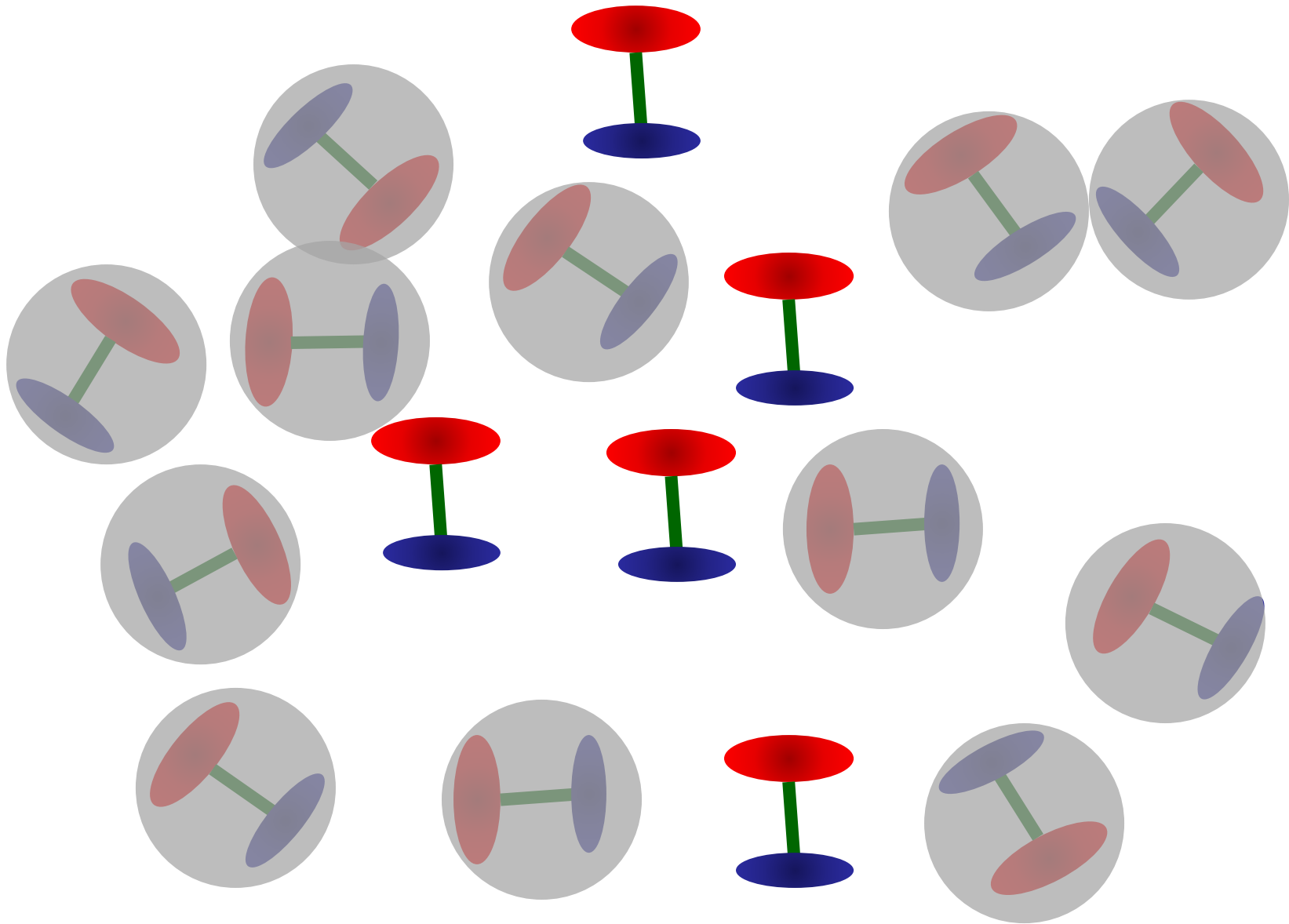
G-band (180) GHz)



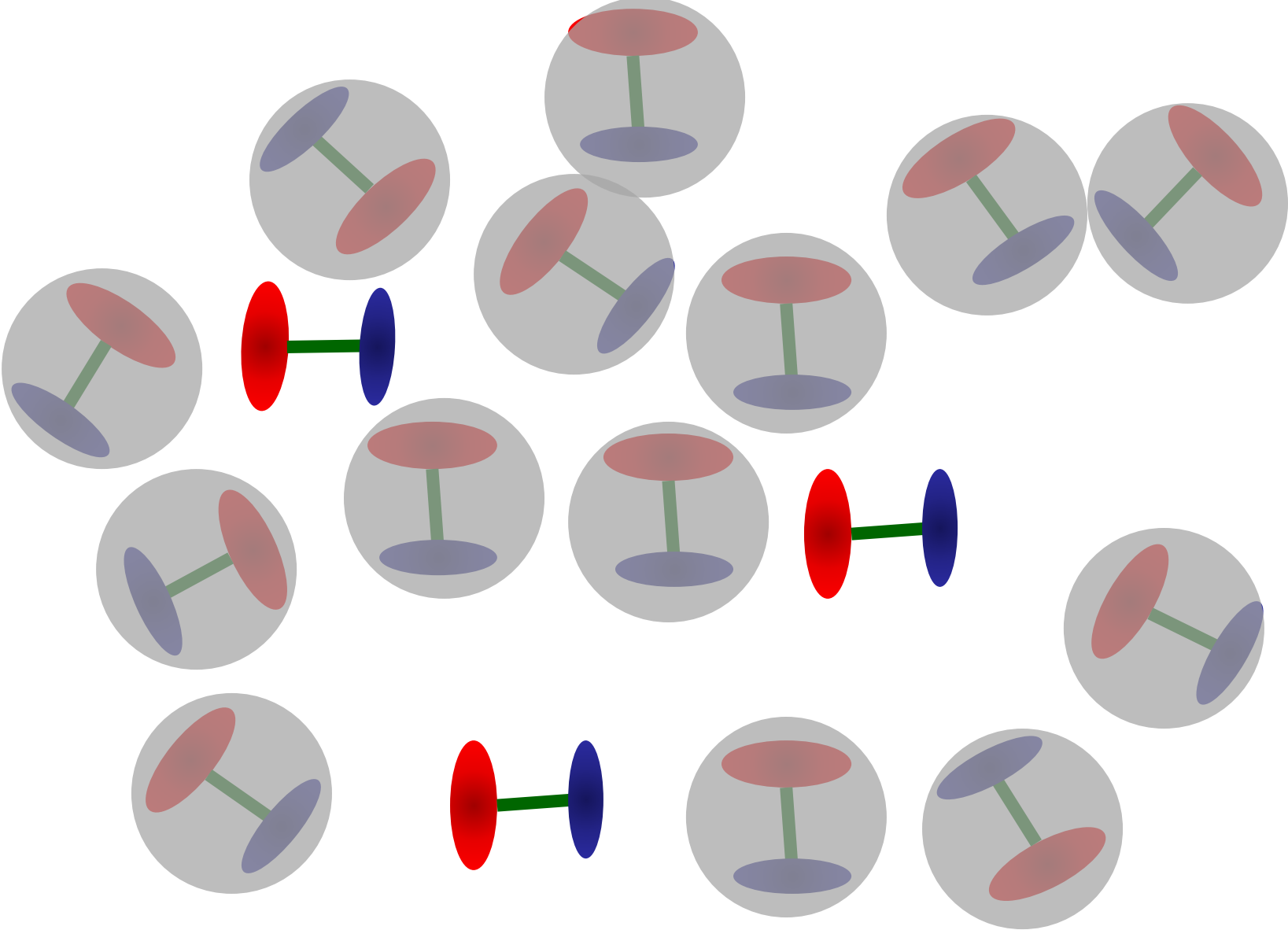
Randomly oriented powder sample



Select only small g value

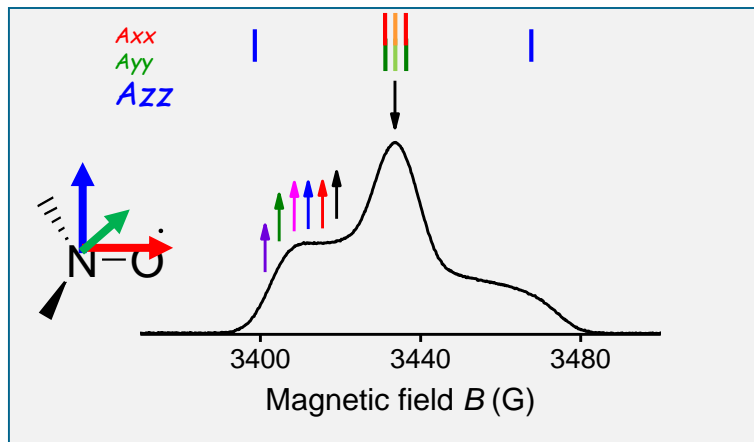


Select only large g-value

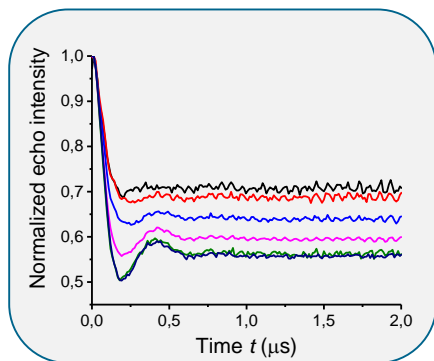


Multifrequency & Multifield PELDOR

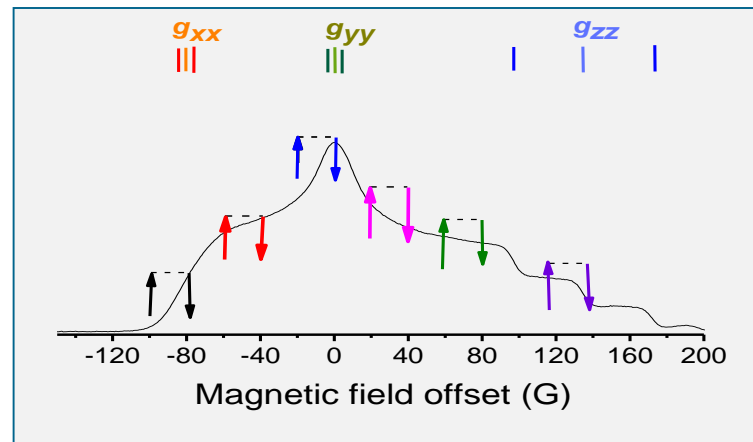
X-band (0.3 T / 9 GHz)



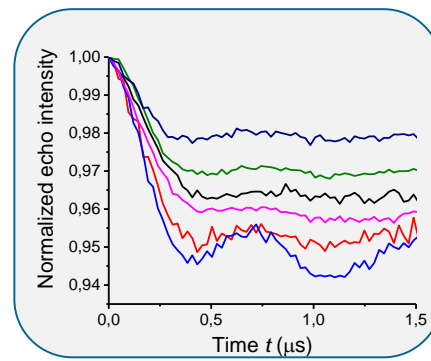
Anisotropic ^{14}N resolution



G-band (6.4 T / 180 GHz)

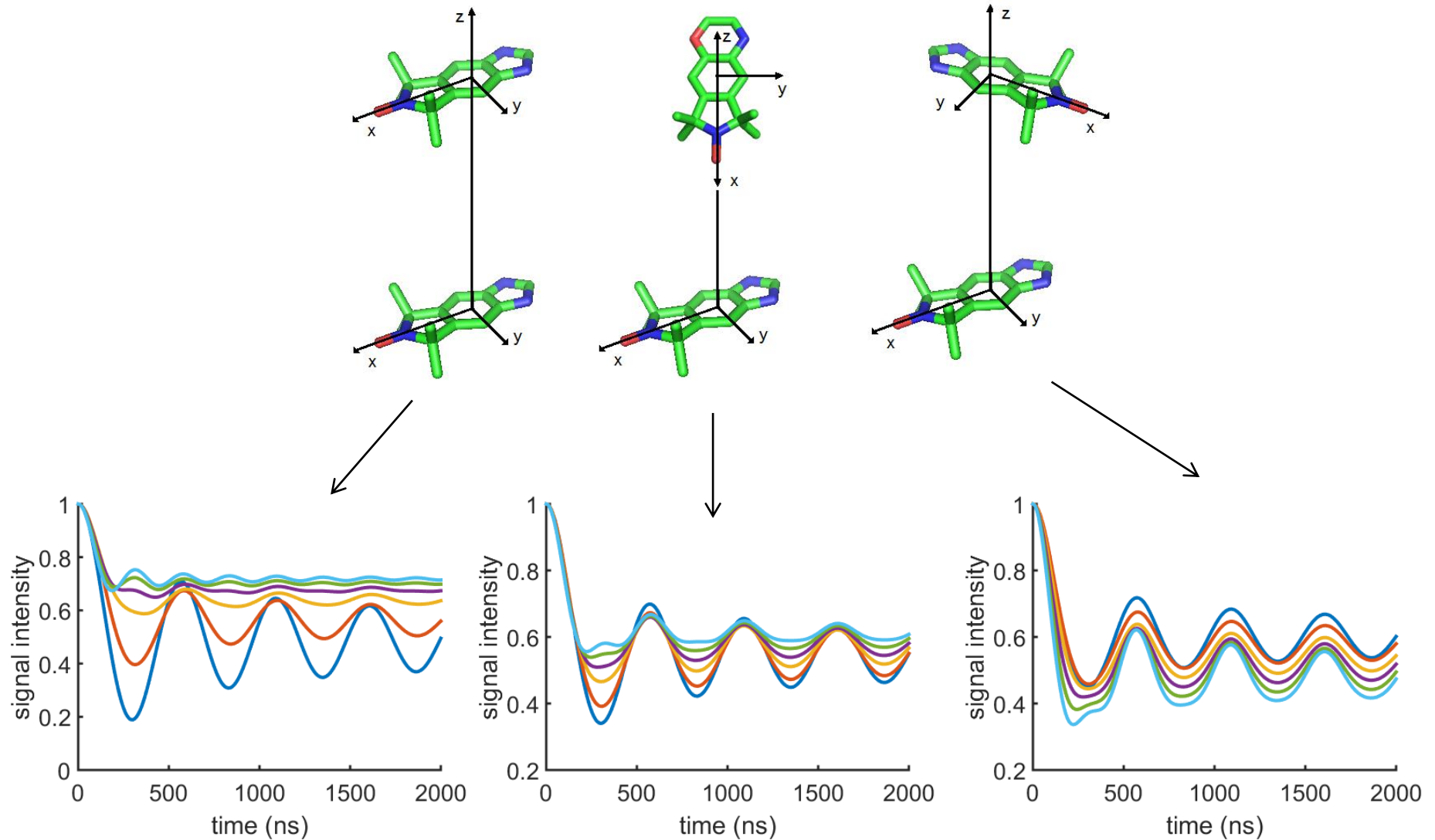


Anisotropic g tensor resolution

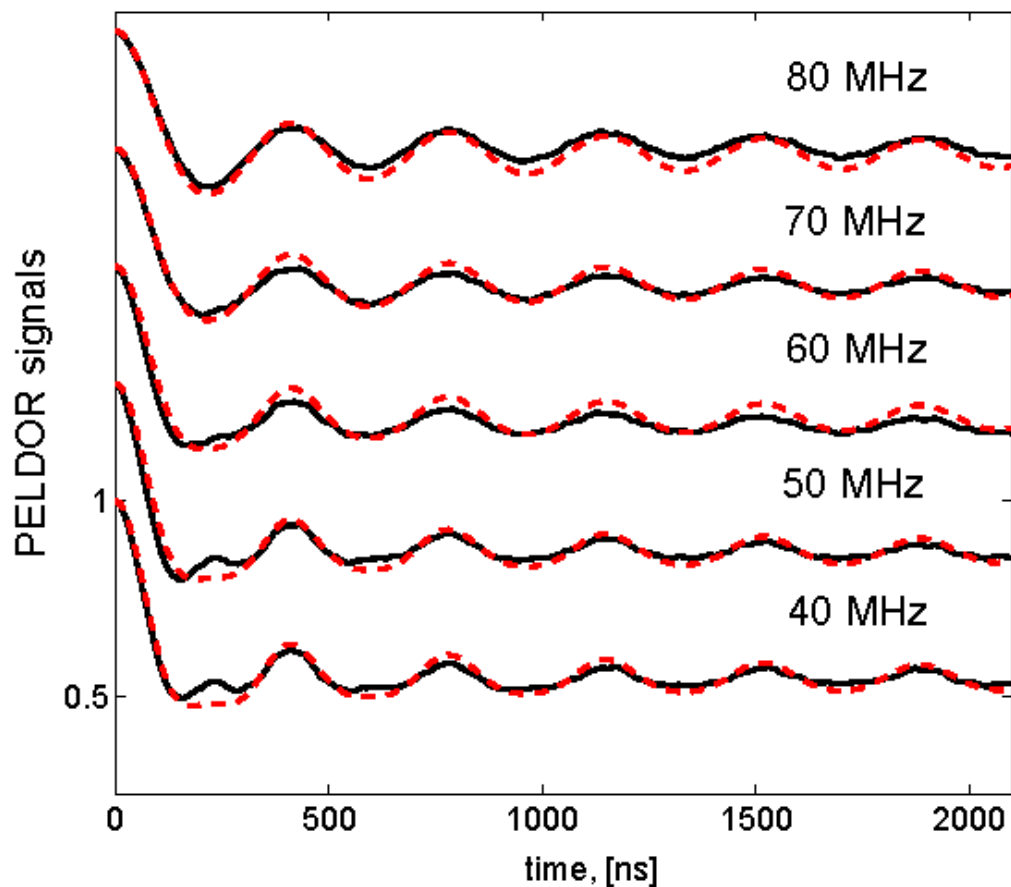
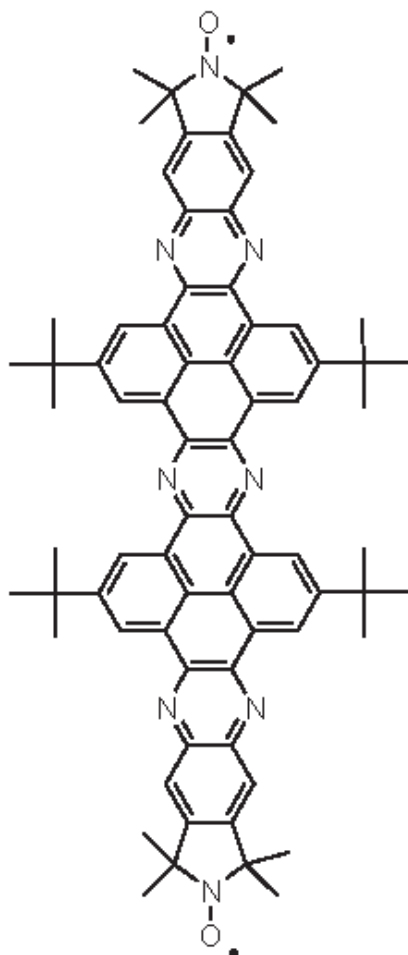


Multi-frequency / Multi-Field PELDOR Dataset

Unique pattern for a given distance / geometry

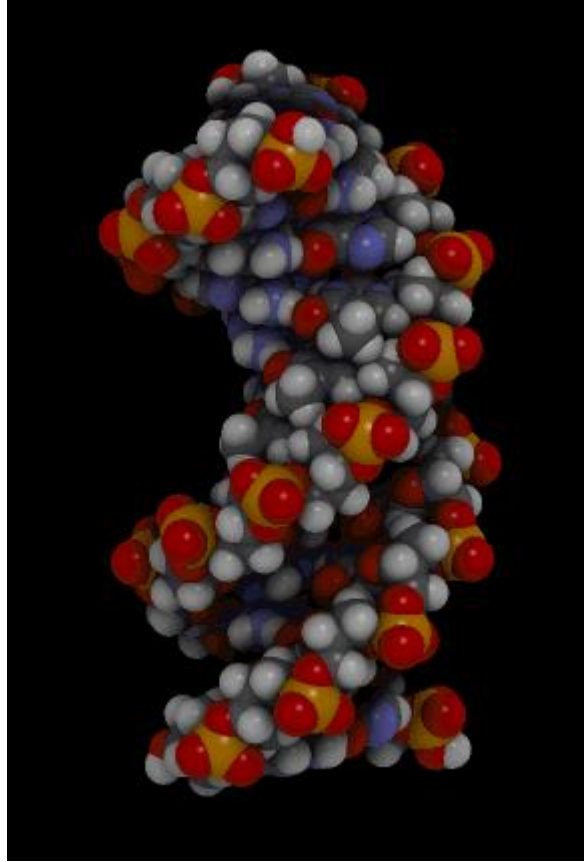


Test with rigid planar biradical

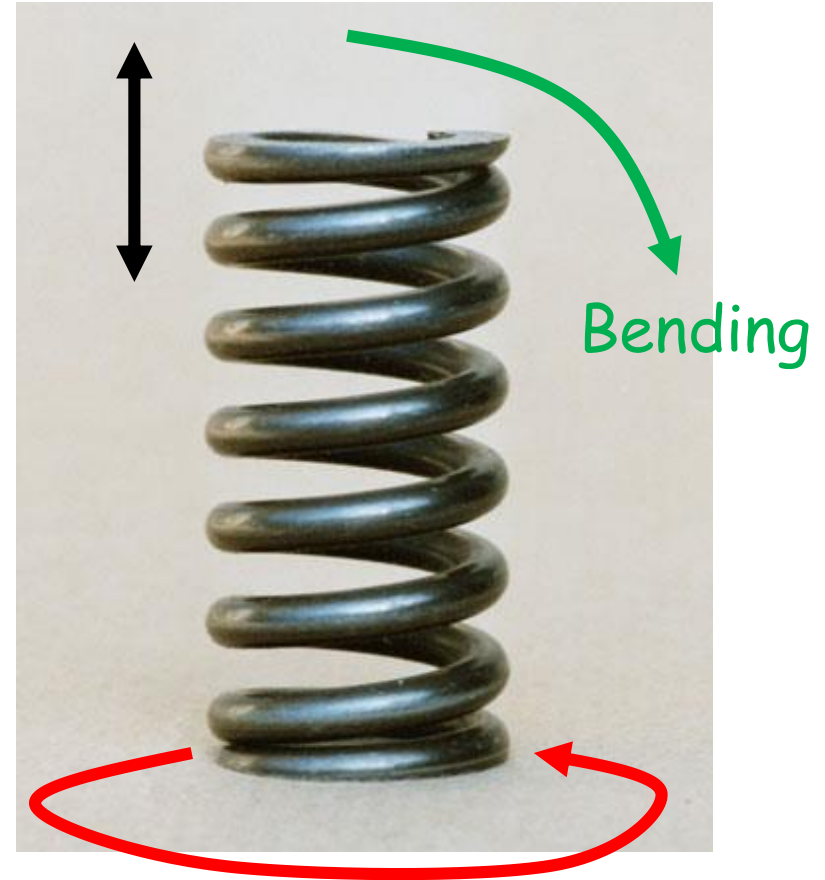


Unique solutions can be obtained for rigid molecules with a single conformation, if experiments performed at several magnetic field strengths.

Determination of internal dsDNA dynamics

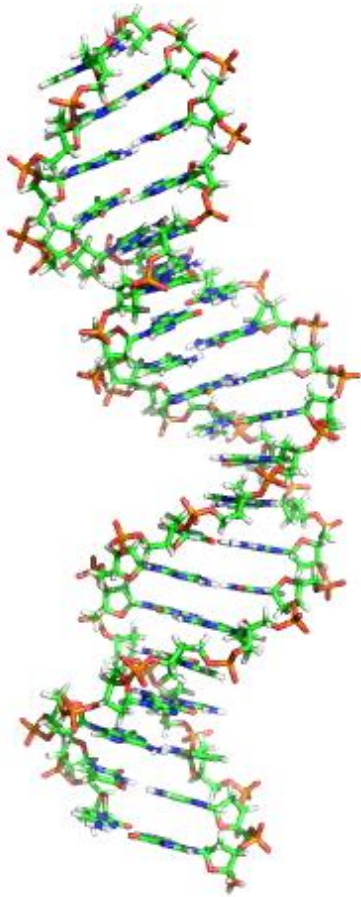


Stretching

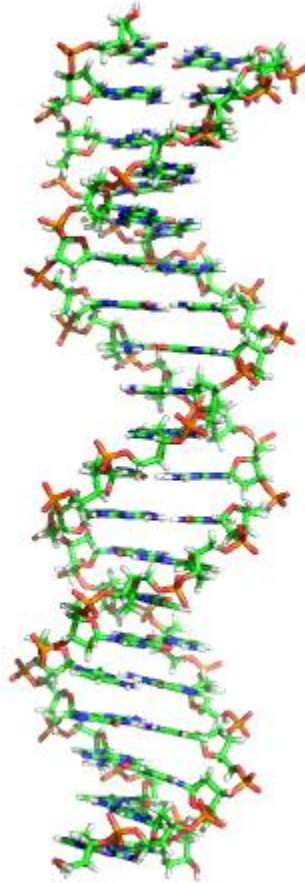


Twisting

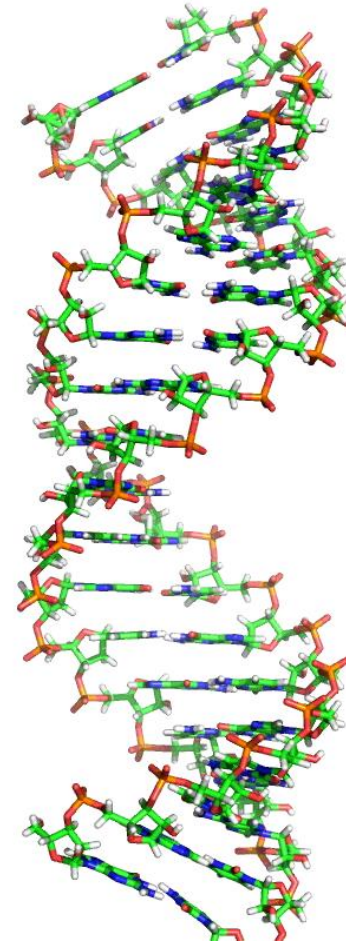
Investigation of dynamics of dsDNA



Bending
 $(\Delta\alpha)$

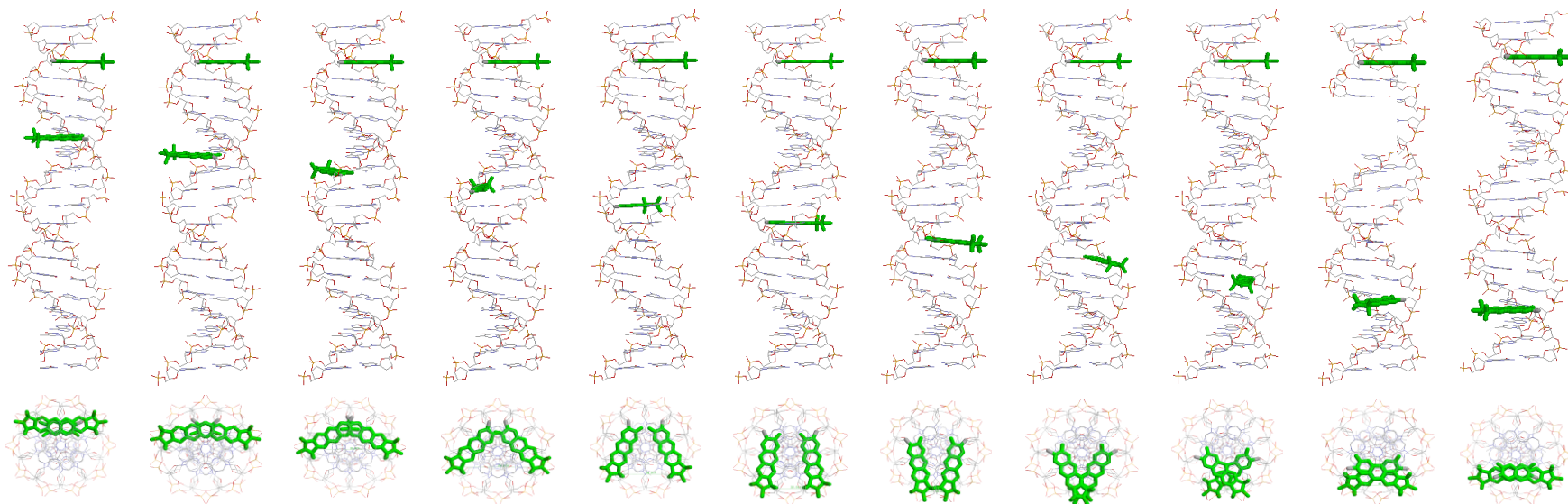


Twisting
 Δr

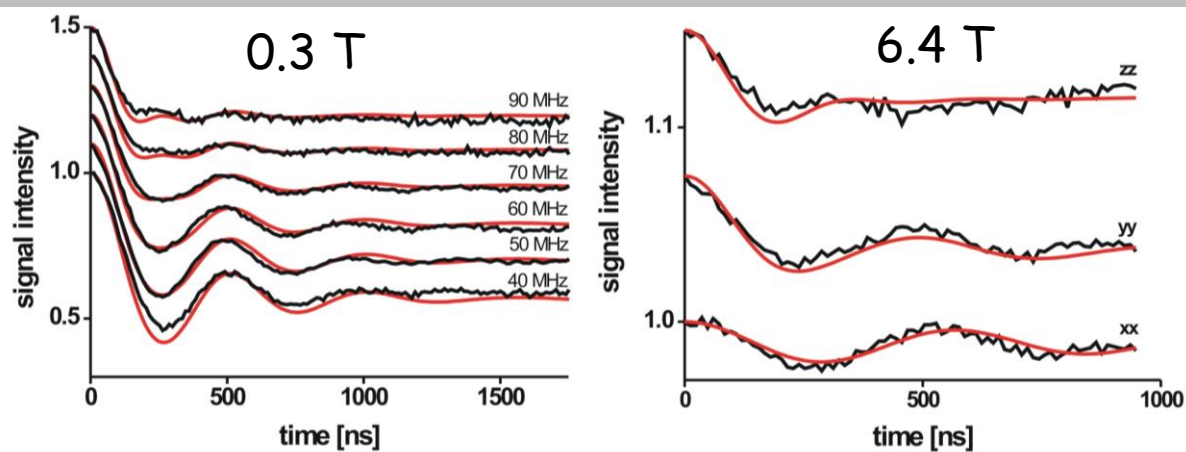


Stretching
 Δh

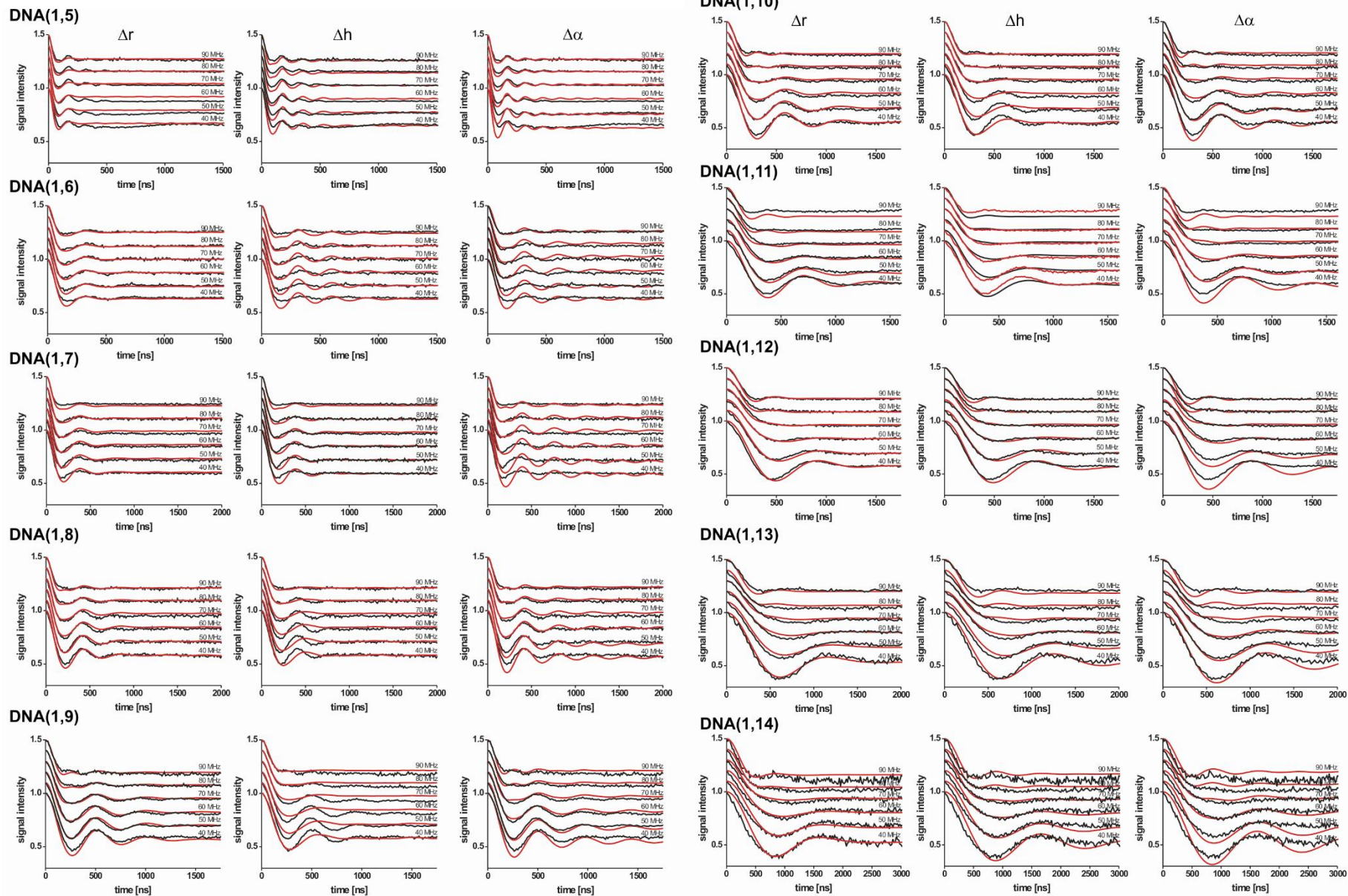
Set of samples with variable distance between spinlabels



Marko JACS (2011)



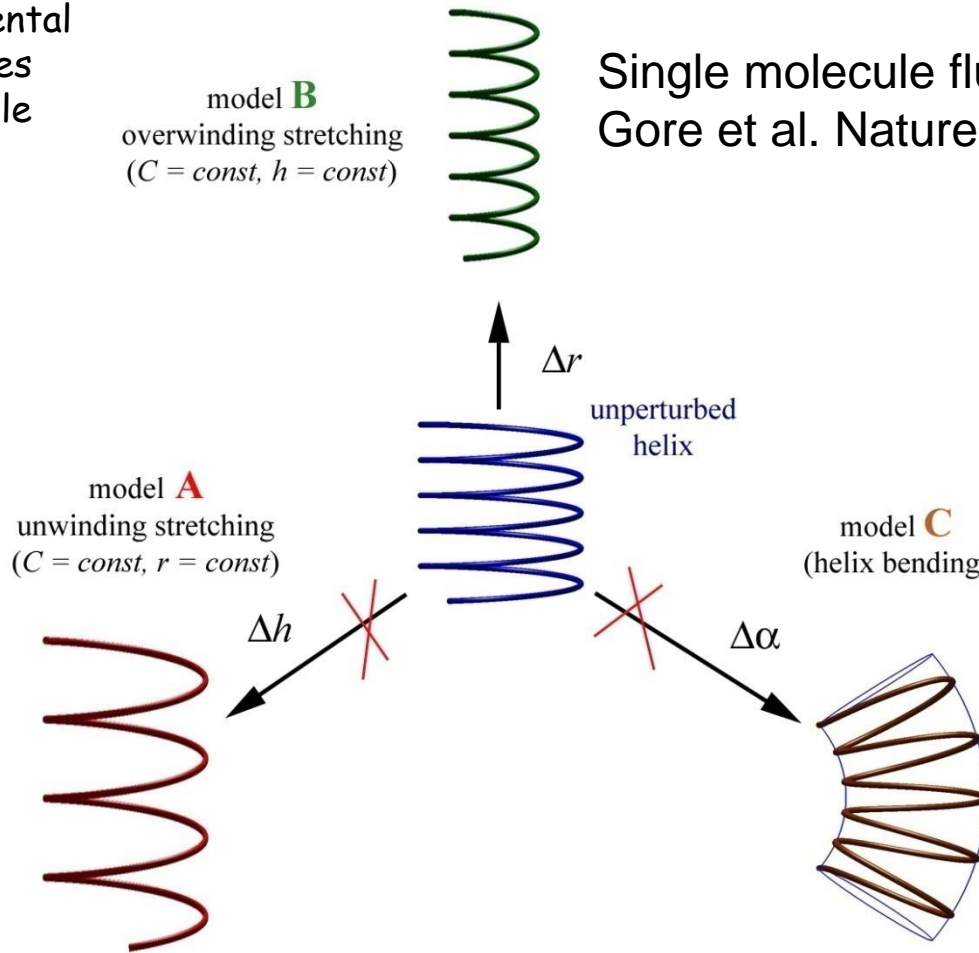
2D-PELDOR datasets and best model fits at 0.3 T



Models for conformational dynamics of dsDNA

The radius changing mode fits best our experimental data but with a 10 times softer stretching module

JACS 2011

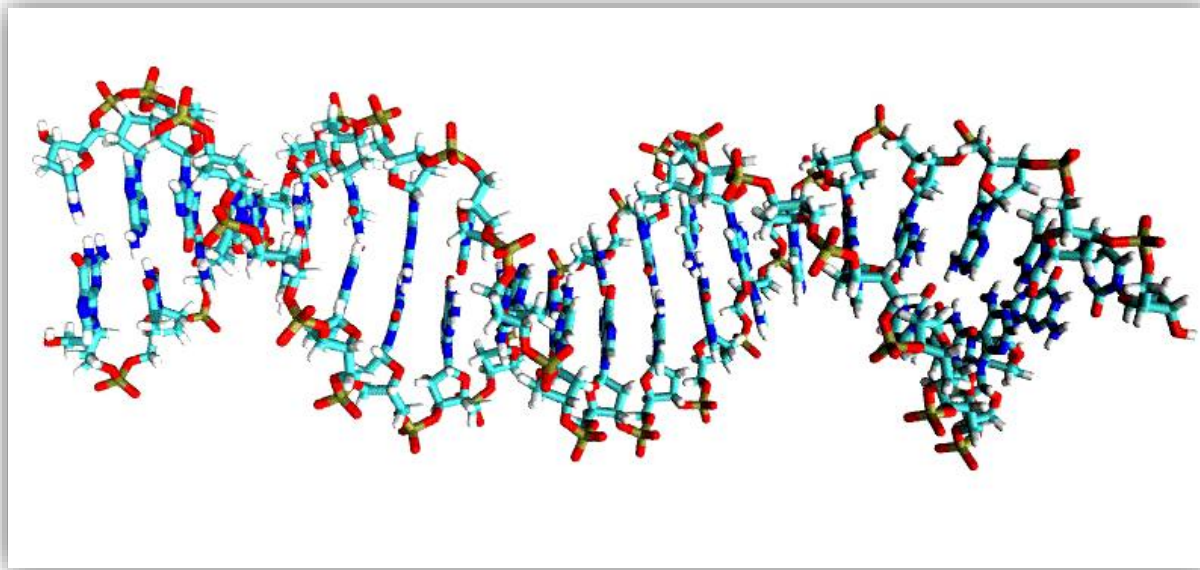


Single molecule fluorescence
Gore et al. Nature (2006)

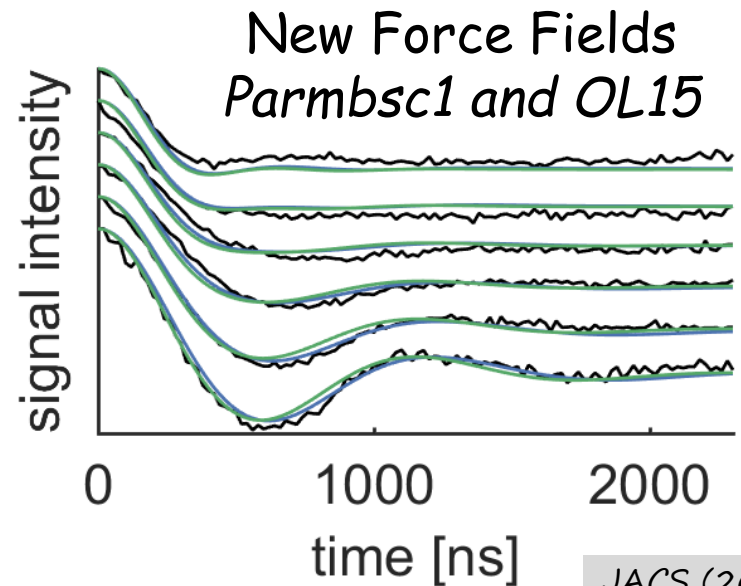
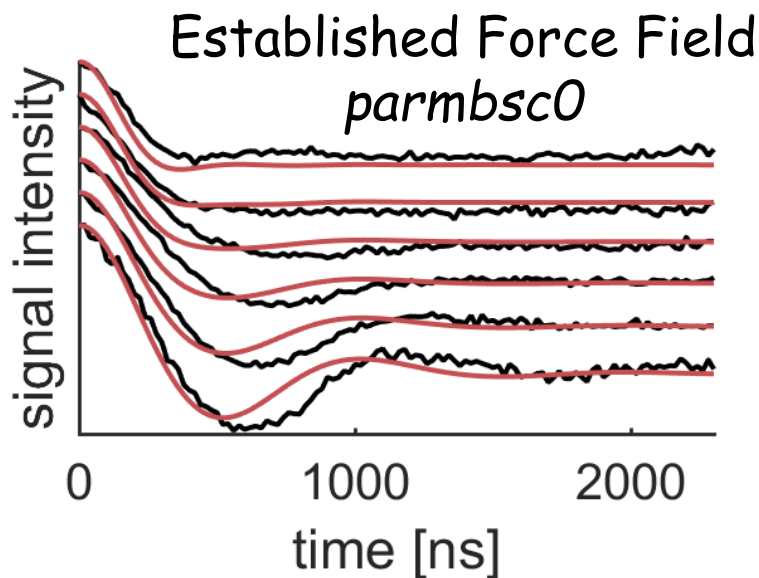
SAXSI
Mathew-Fenn et al. Science (2008)

Modeling
Becker et al. Science (2009)

Comparison with MD simulations of dsDNA



Collaboration with
G. Hummer
(MPI Biophysics,
Frankfurt)



Echo sequences with broadband shaped pulses

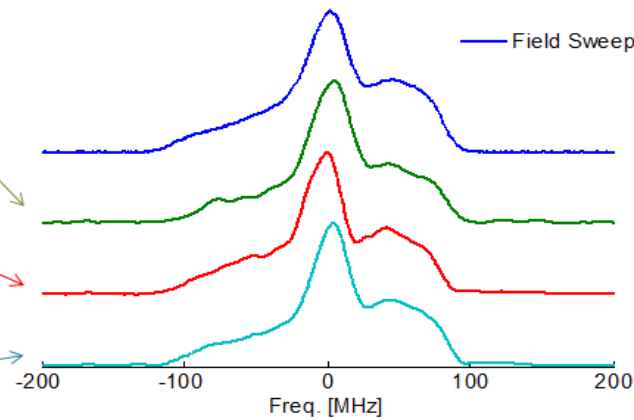
Broadband refocused echo



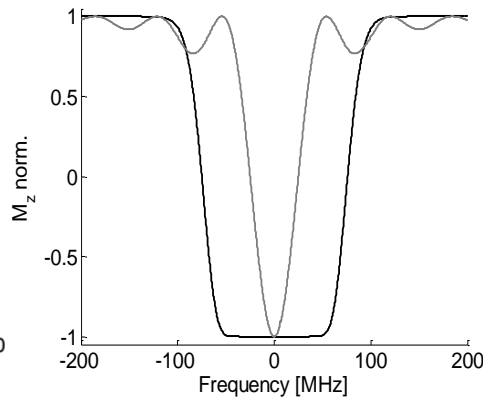
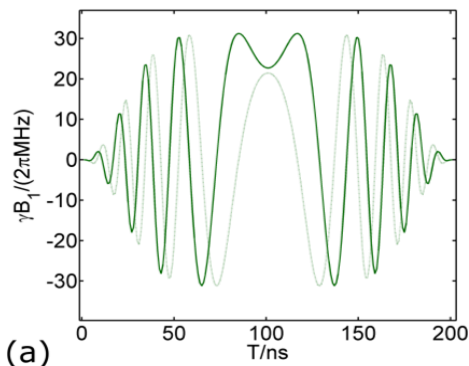
Broadband stimulated echo



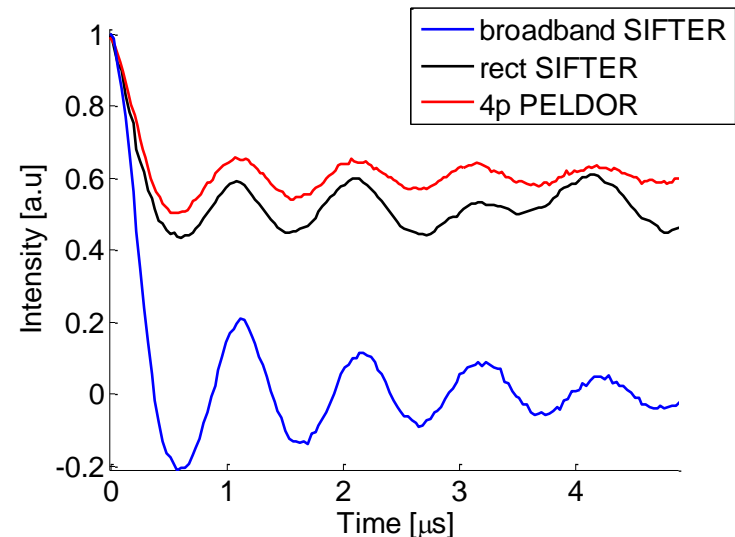
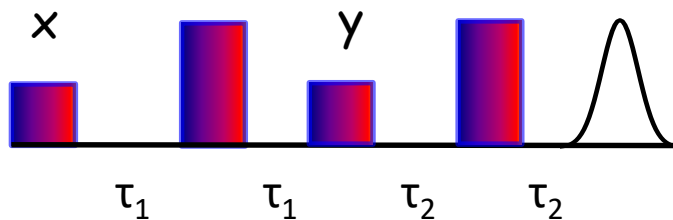
Broadband Hahn echo



Full excitation of nitroxide spectra at X-band possible



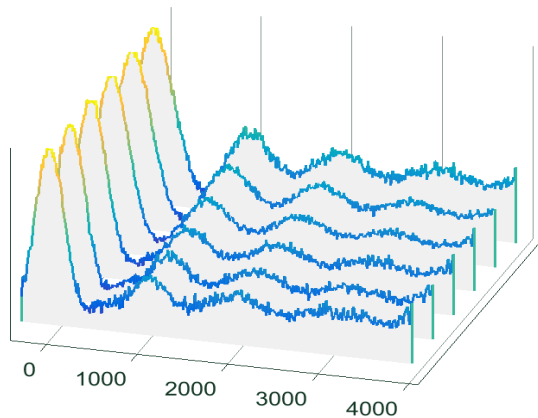
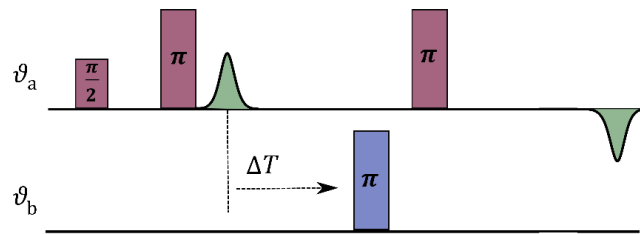
Sech/tanh Pulses
Length of 200 ns
Bandwidth of 200 MHz



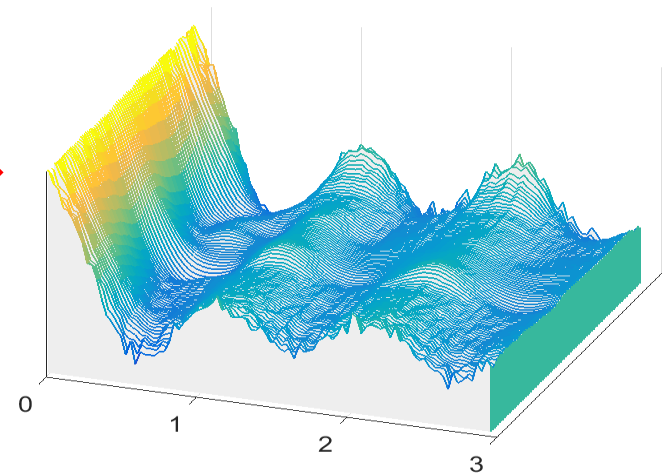
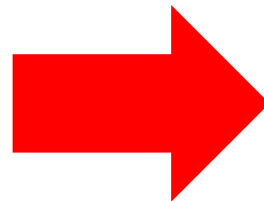
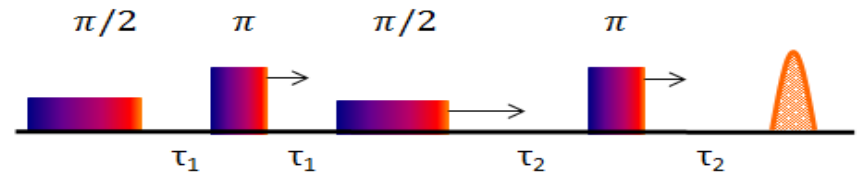
2D SIFTER experiment using broadband pulses

Doll, Jeschke PCCP 18, 23111 (2016)

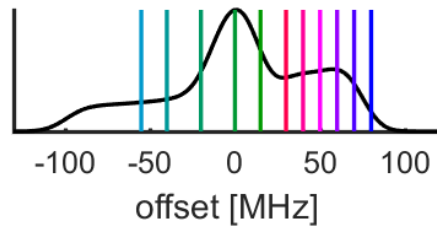
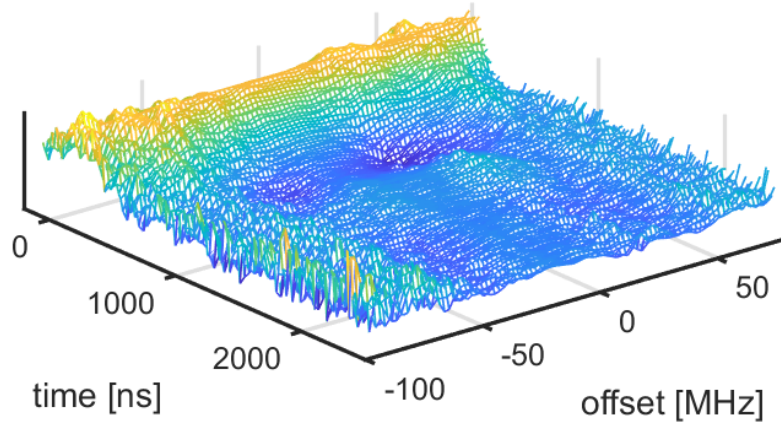
4 pulse PELDOR/DEER
with several probe
frequencies



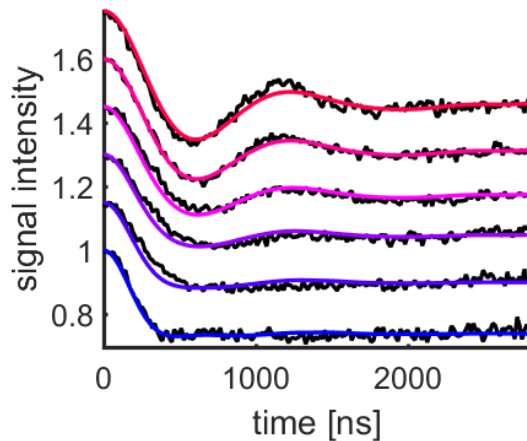
Broadband SIFTER with FT
of Echo signal



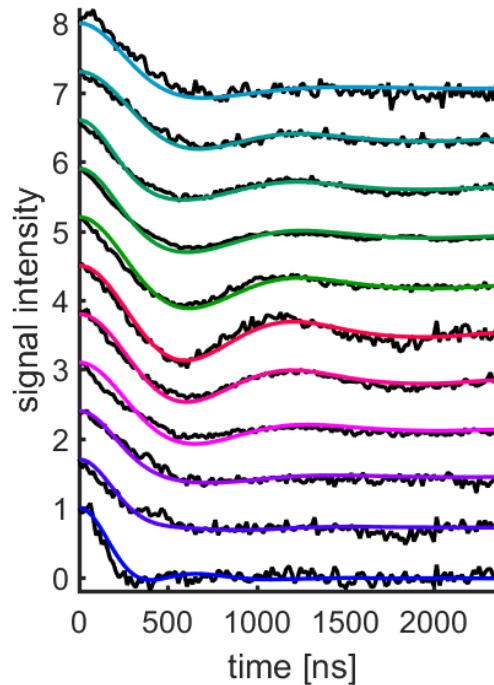
2D broadband SIFTER Experiment on dsDNA



PELDOR

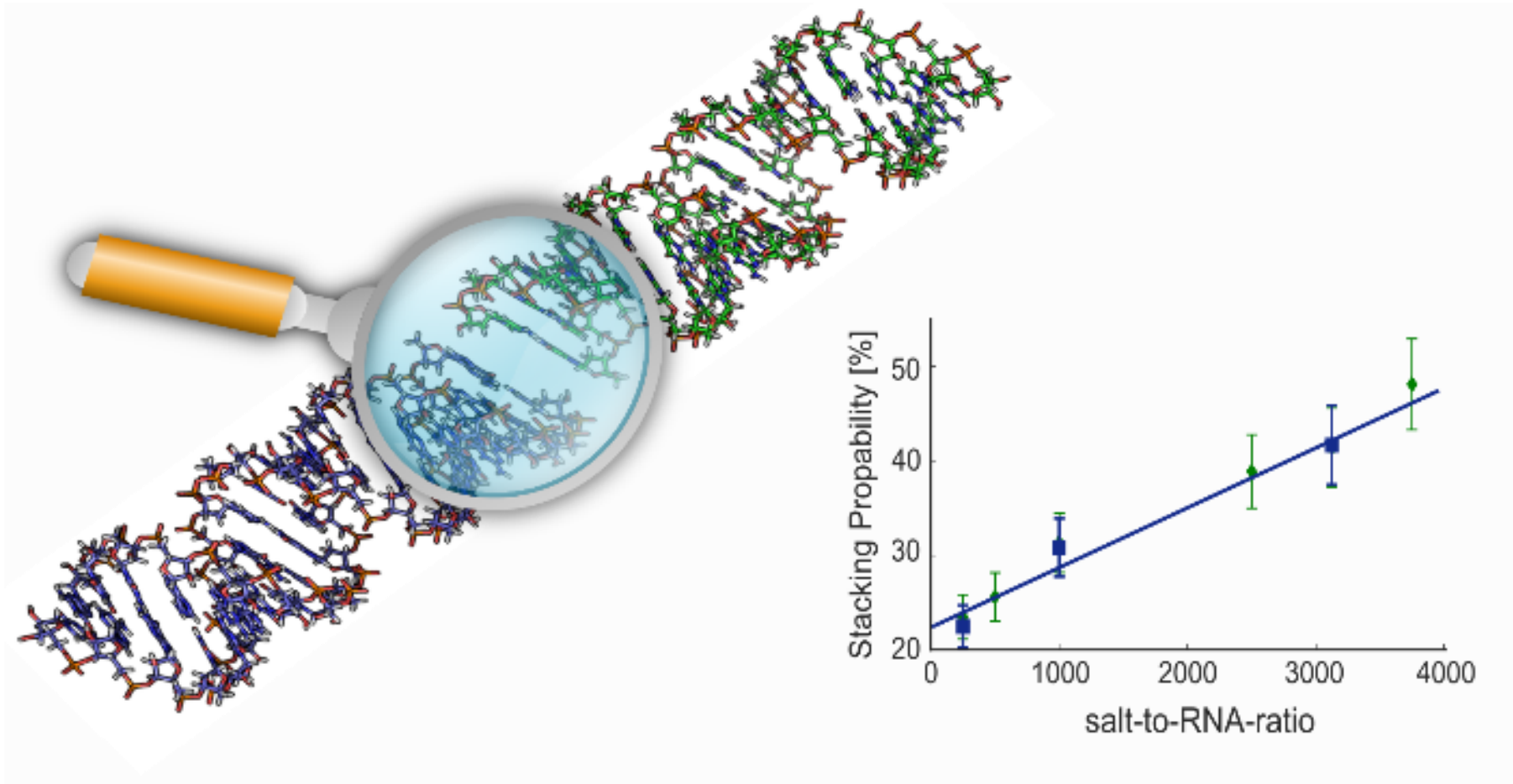


SIFTER



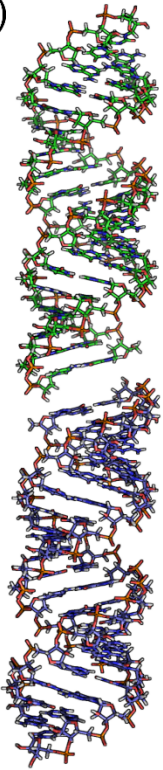
- Faster
- Higher resolution
- Over full spectrum

Intermolecular Interactions of dsRNA

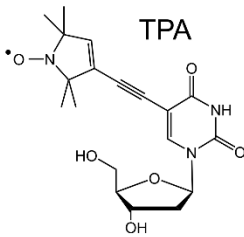


Conformational dynamics of RNA

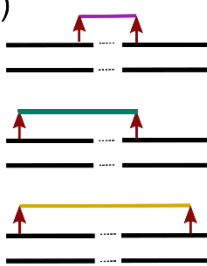
a)



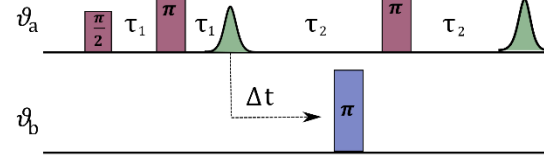
b)



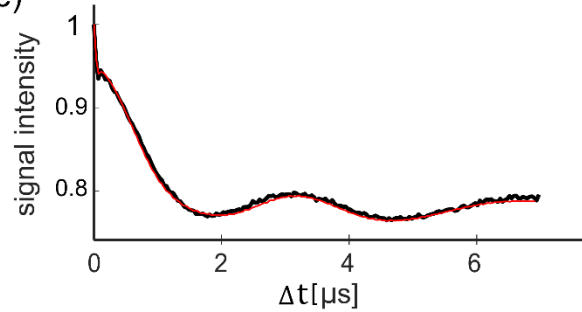
c)



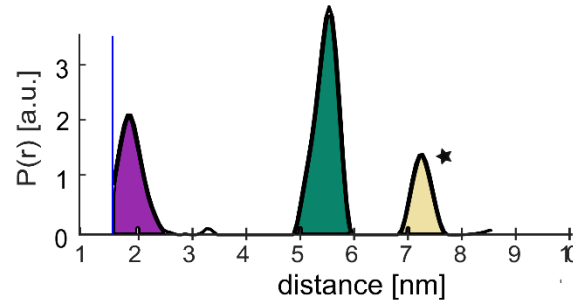
d)



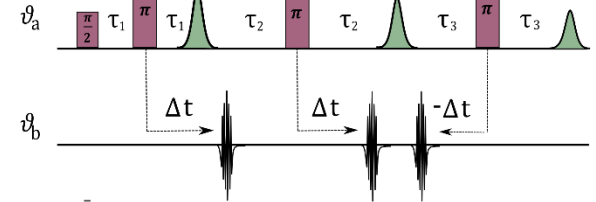
e)



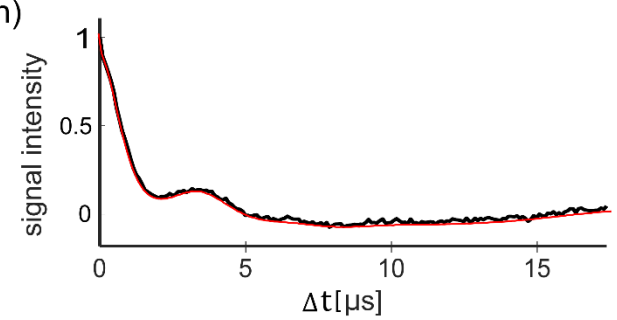
f)



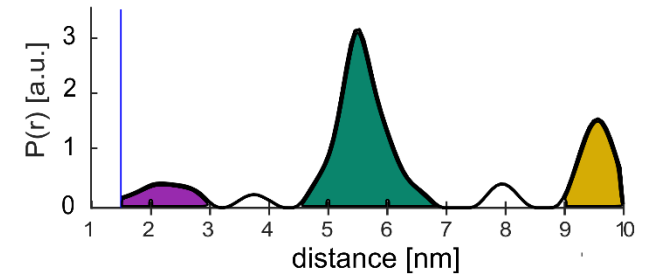
g)



h)

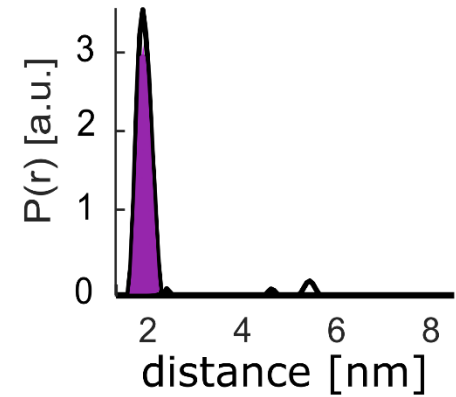
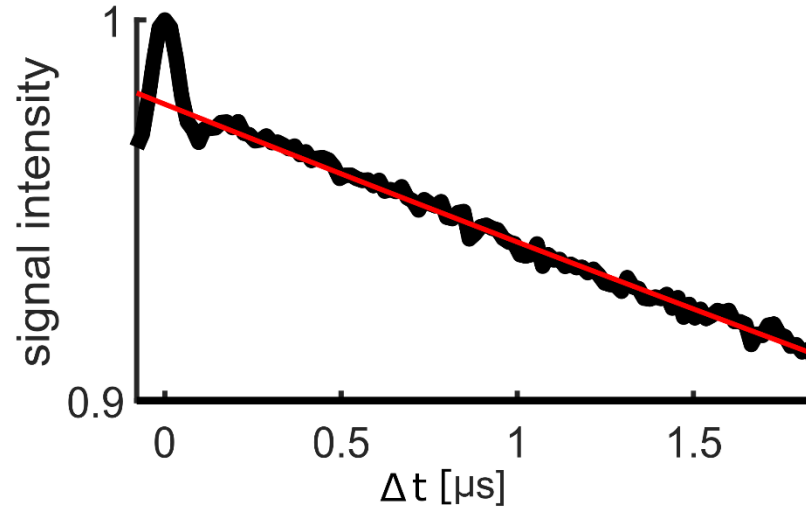
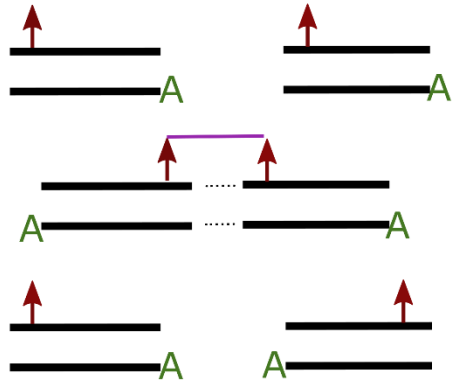


i)

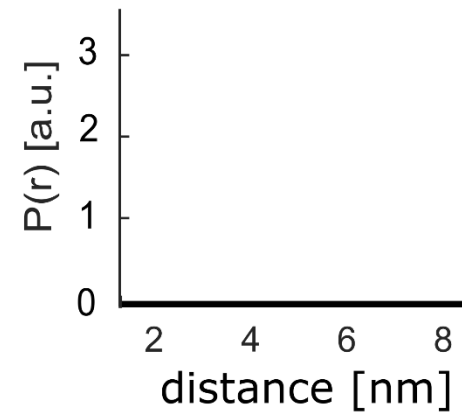
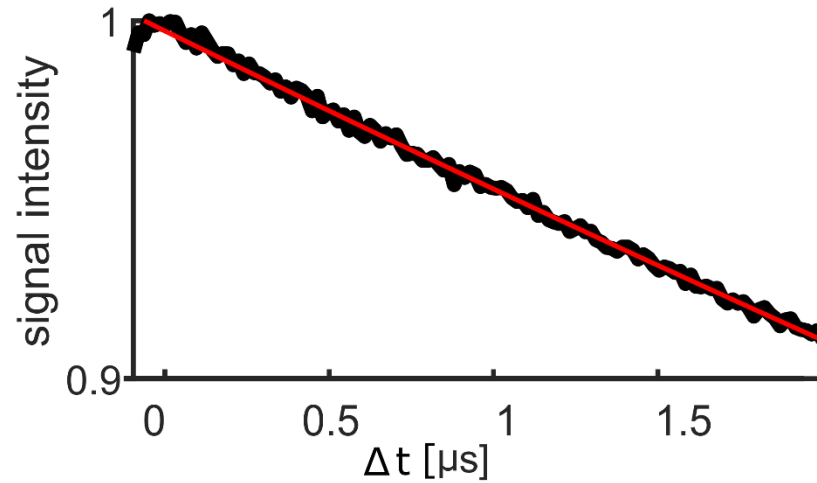
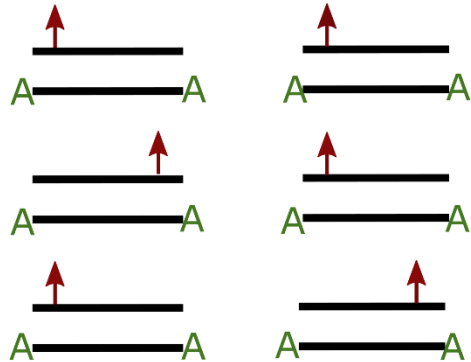


Conformational dynamics of RNA

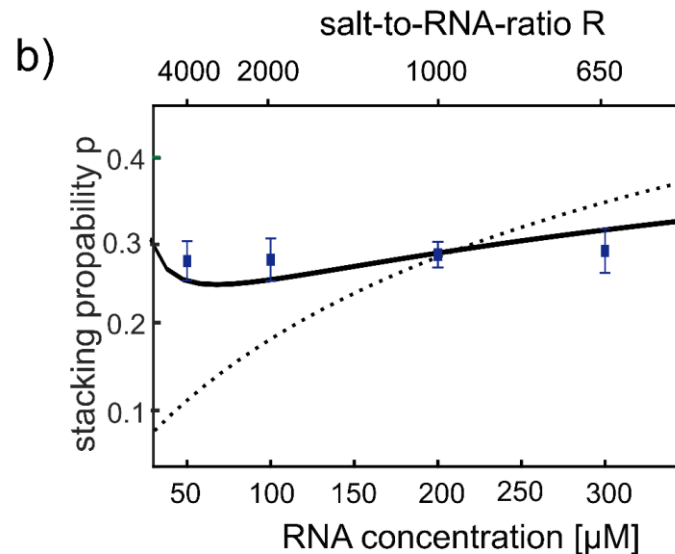
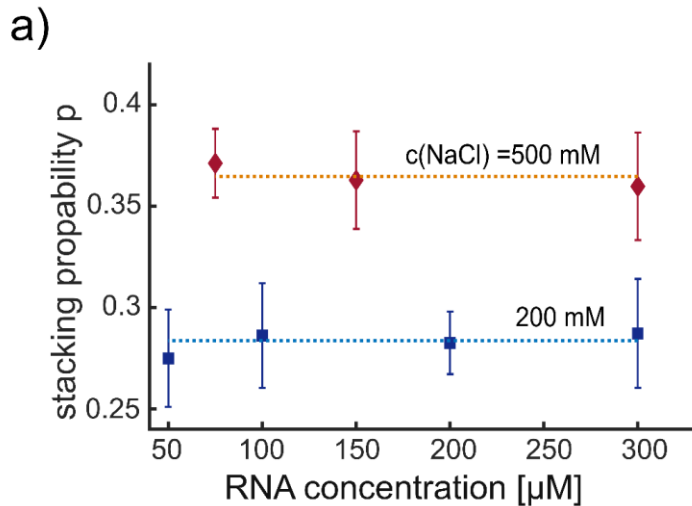
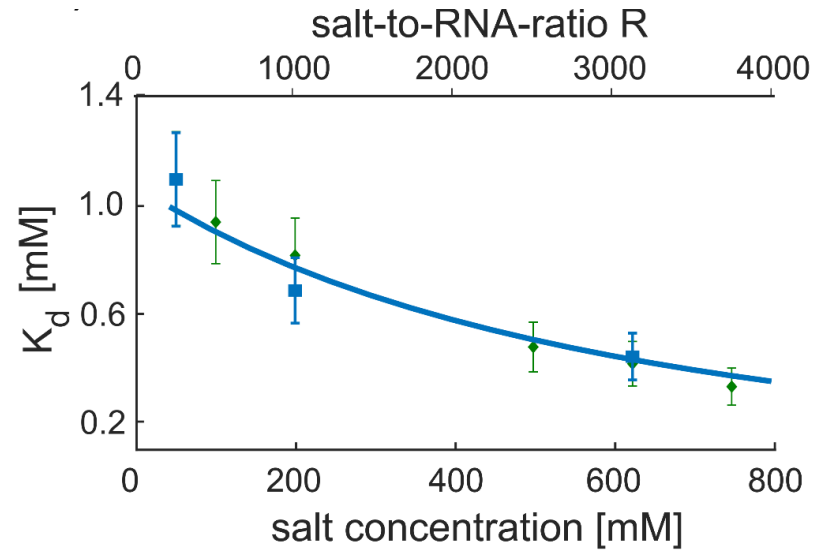
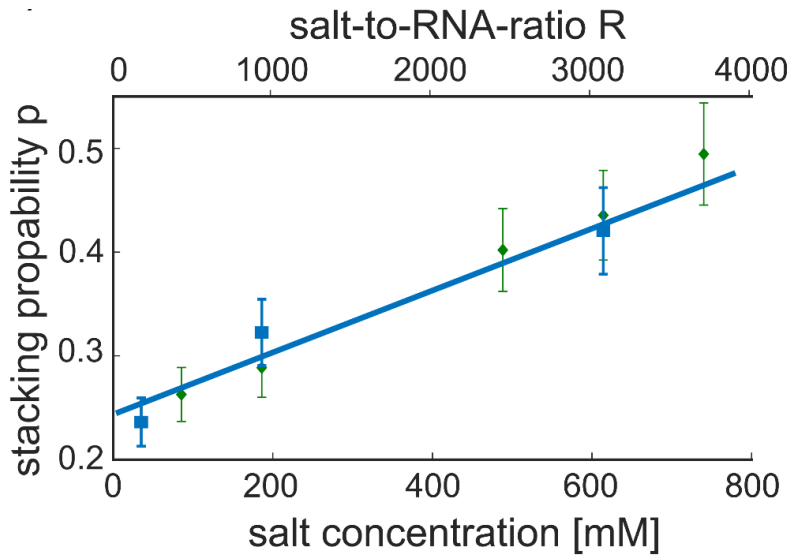
a)



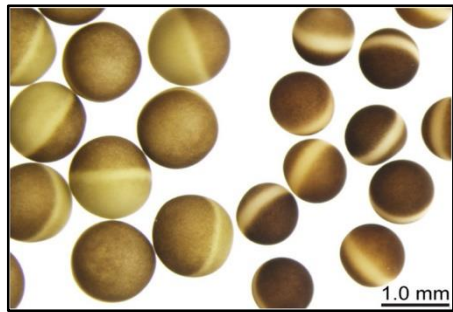
b)



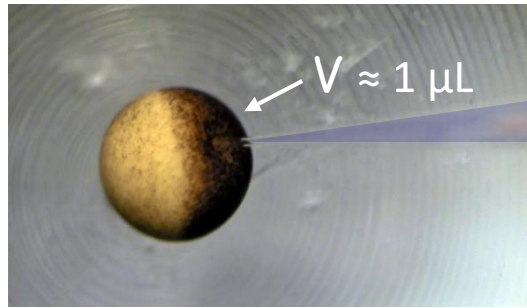
Conformational dynamics of RNA



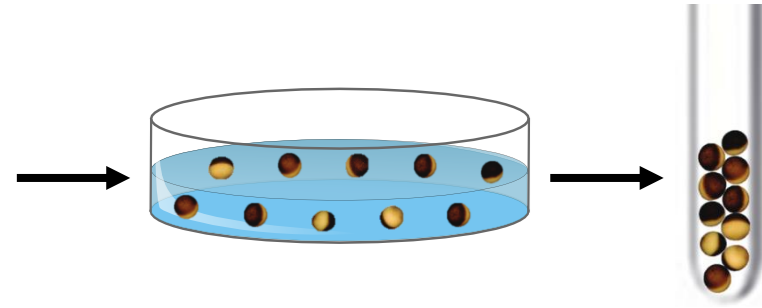
In-cell investigations of NAs



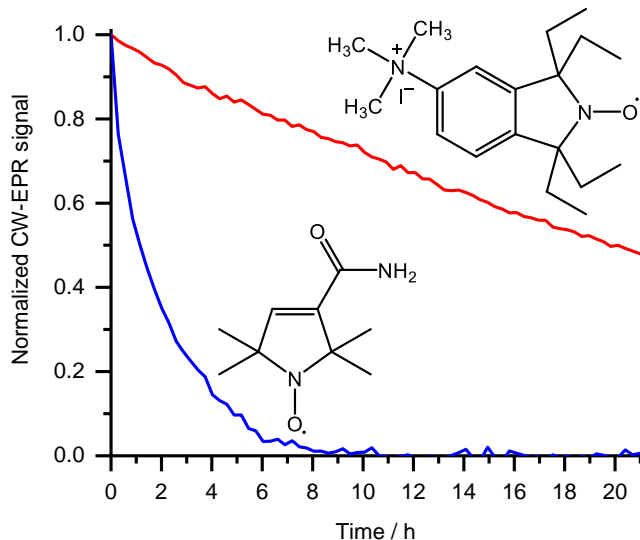
oocyte selection
by morphology



microinjection
(1 mM, 50 nL dsRNA)

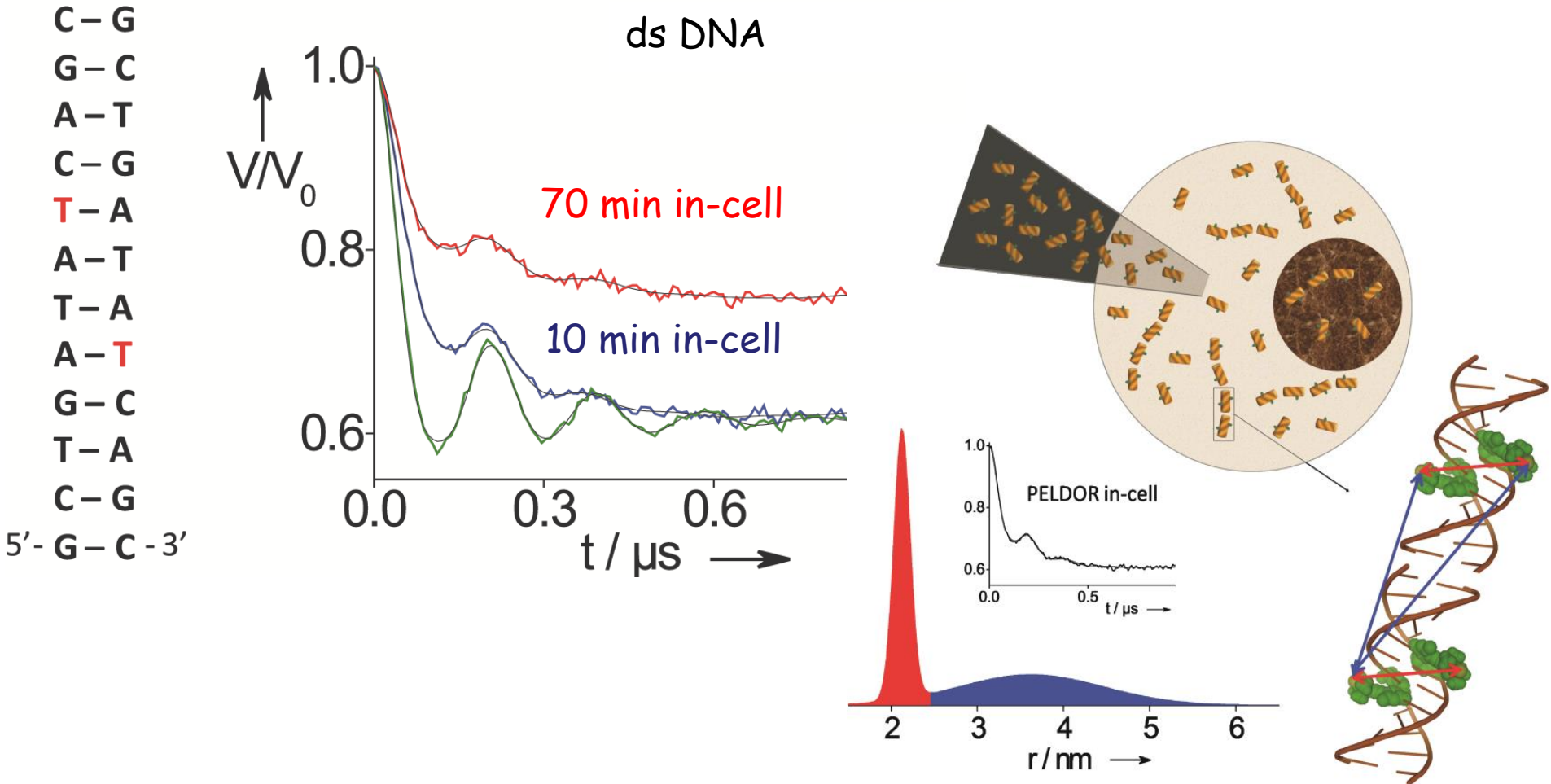


washing and incubation
(60', T = 16°C)



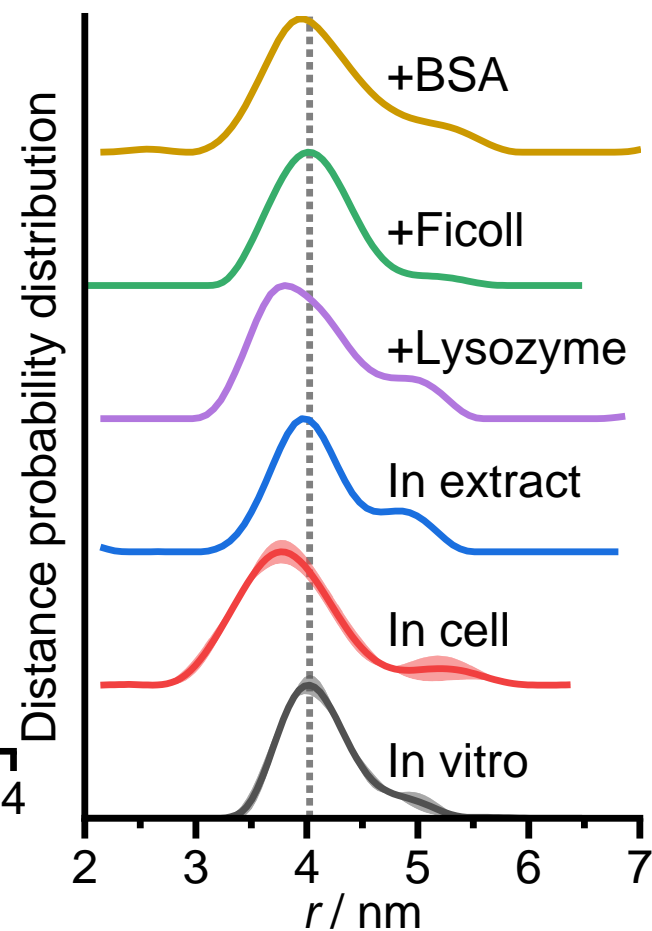
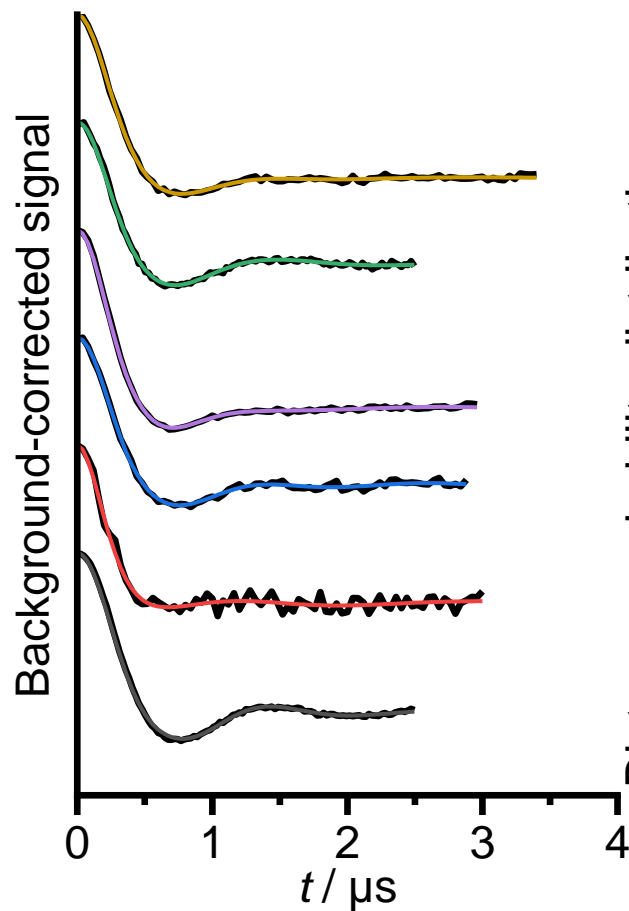
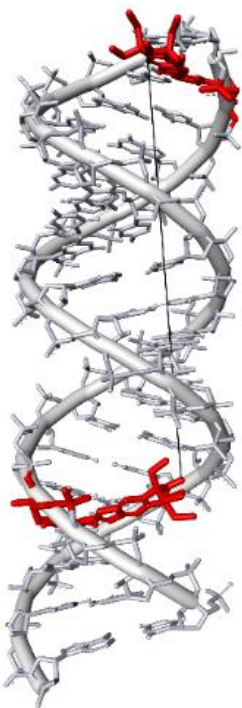
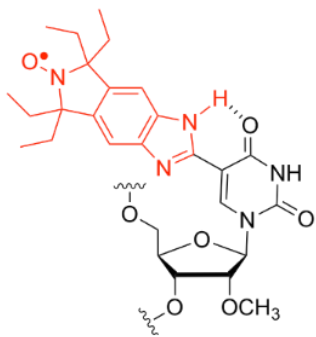
Nitroxide Spin Labels have to be protected against reductions for applications in cells

In-cell applications of PELDOR spectroscopy

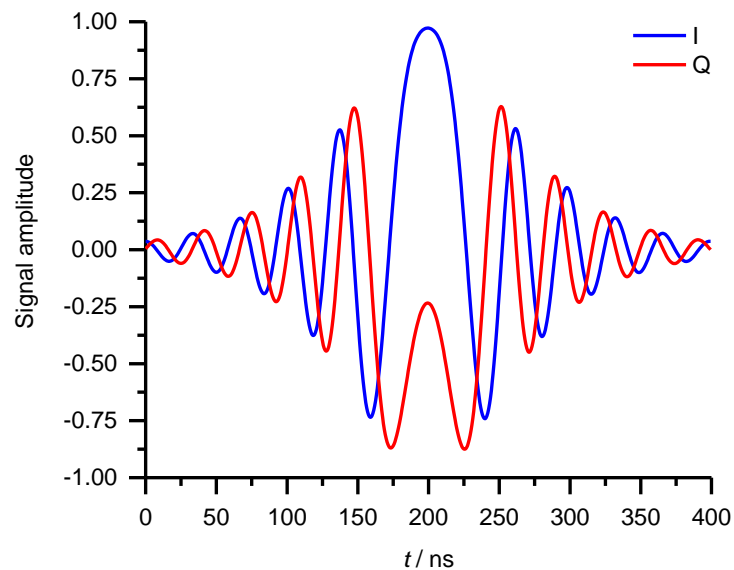
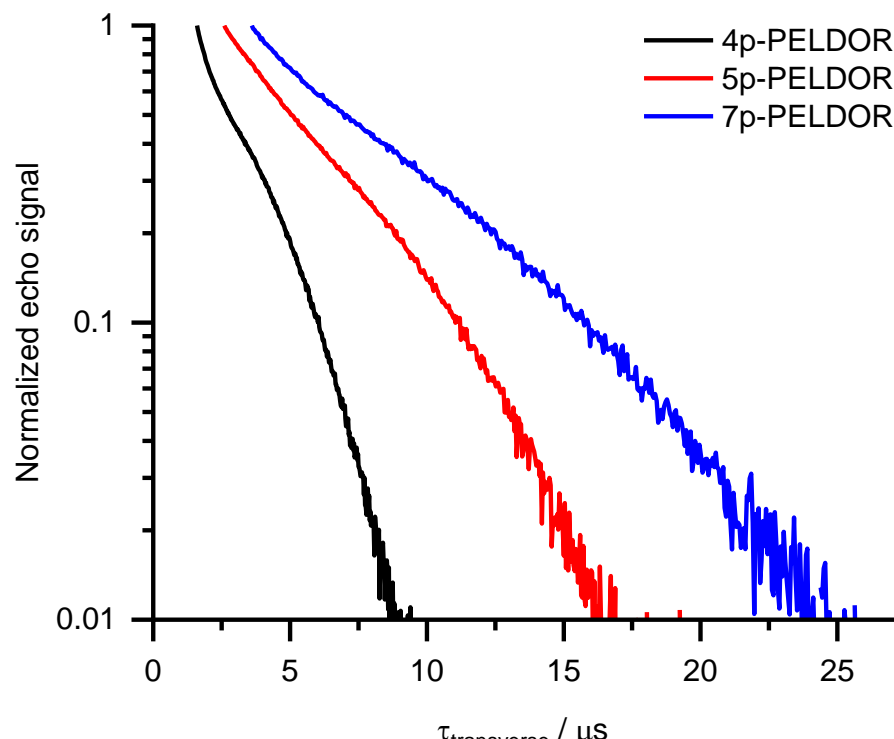
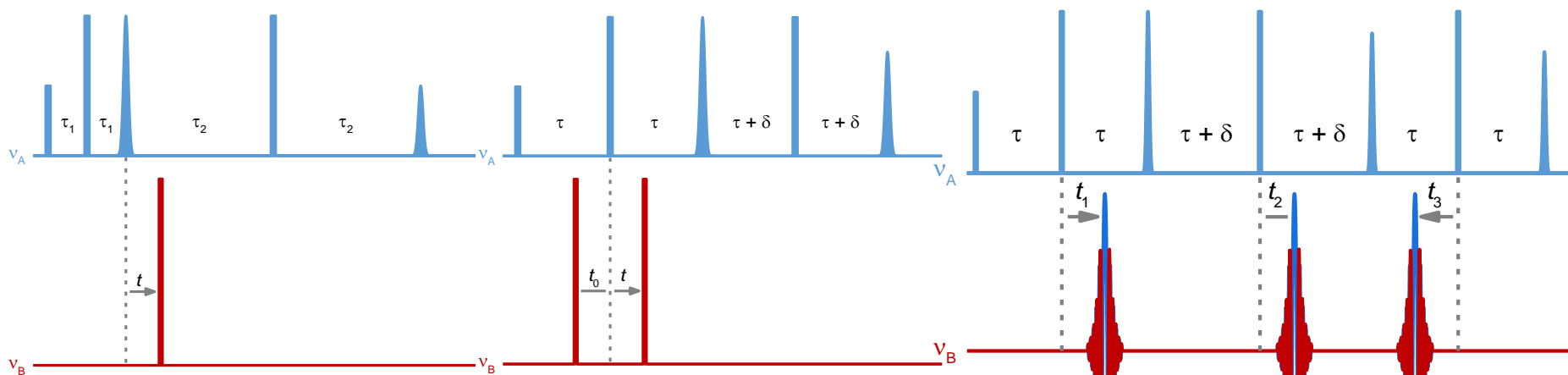


Second distance from ds-DNA
stacking in-cells

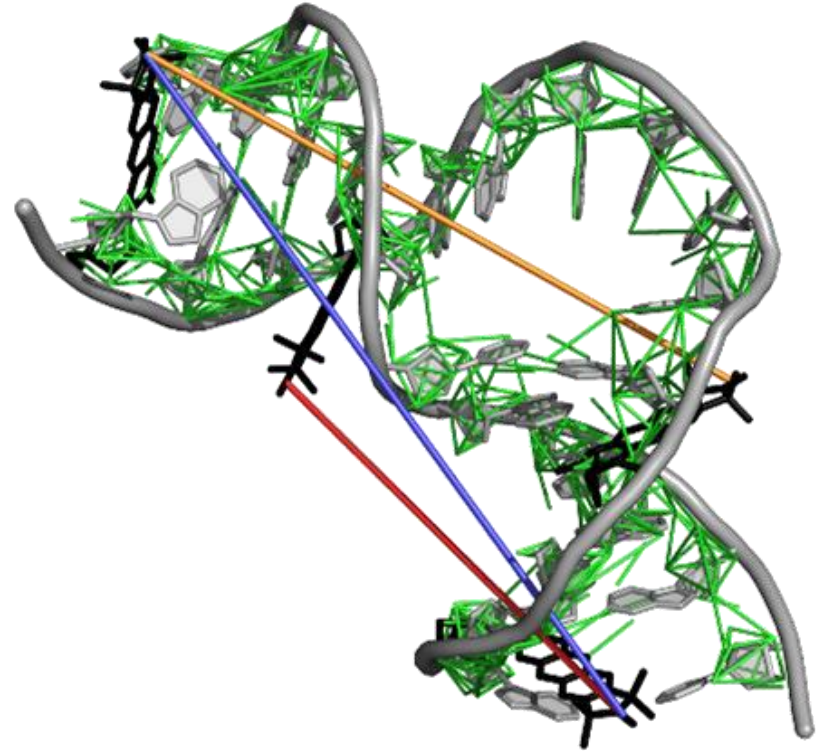
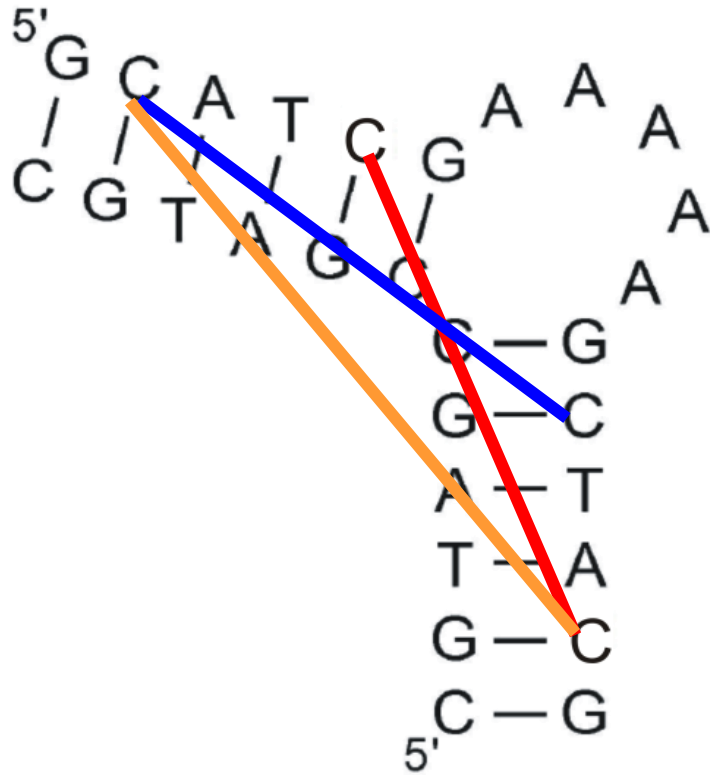
In-cell applications of PELDOR spectroscopy



Prolonged observation time window by CP Sequence



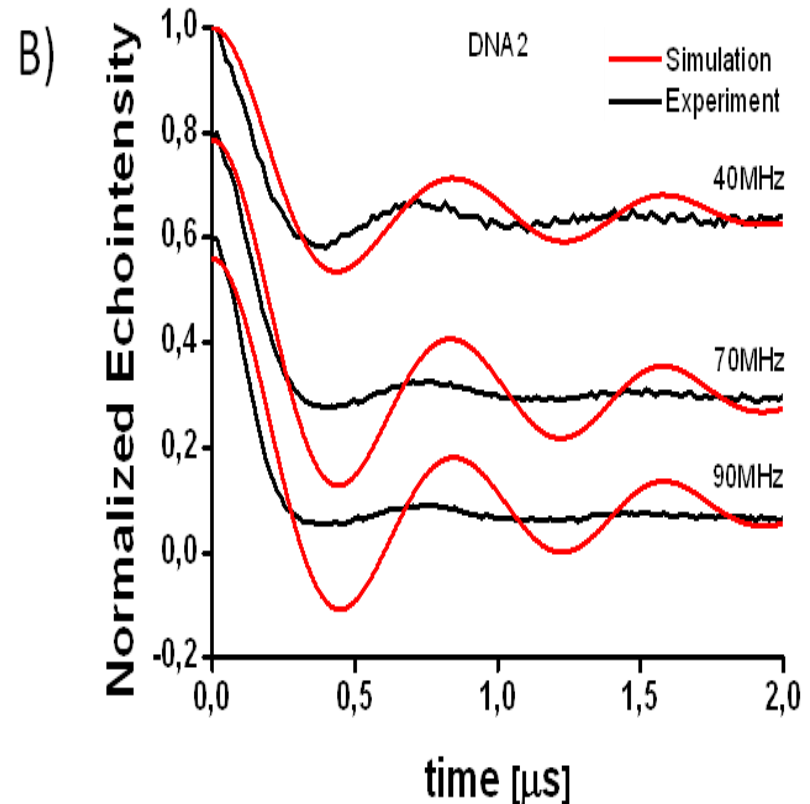
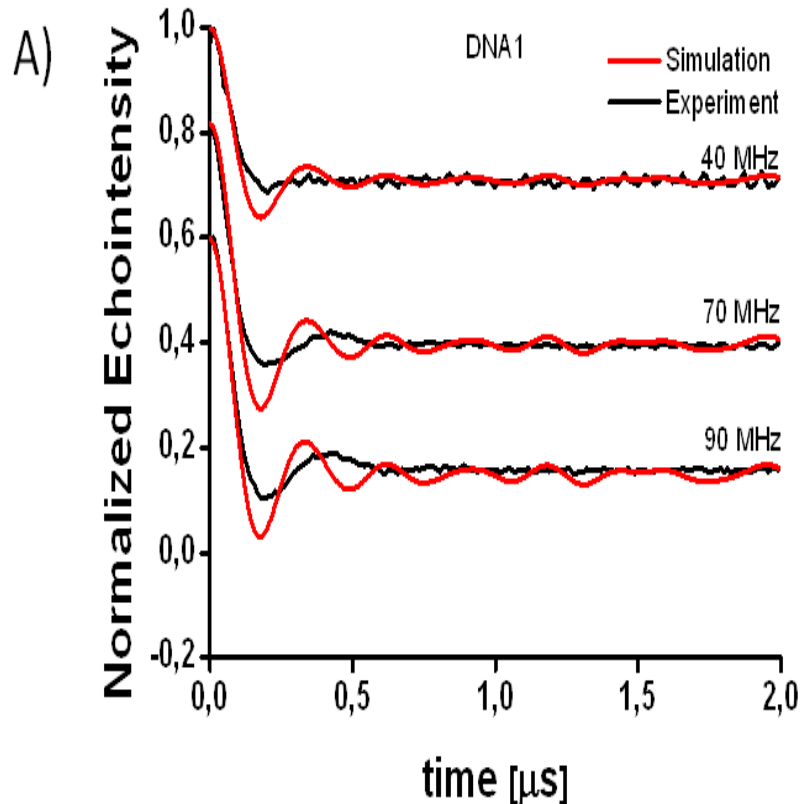
Investigation of DNA with A-bulge



NMR restraints +
3 Pairs of EPR restraints

Comparison with predictions from NMR structure

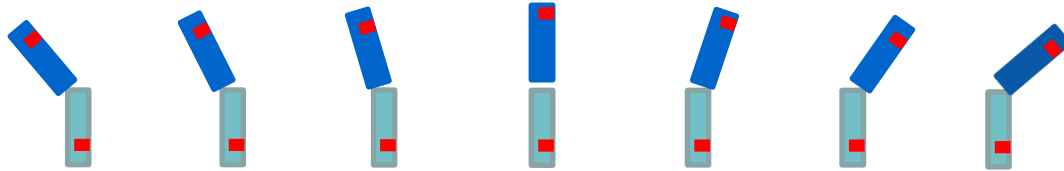
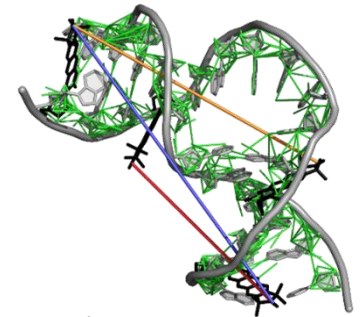
Dornberger et al.
Biochem. 38, 12860 (1999)



PELDOR time traces calculated from NMR structure
(20 best structures) do **not agree** well with PELDOR !

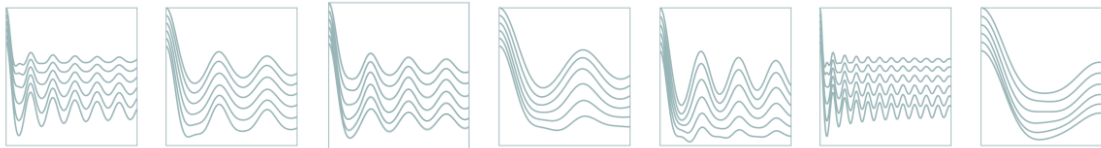
Procedure to determine conformational ensemble

1. Step: Modeling with NMR restraints & PELDOR R's (CYANA)



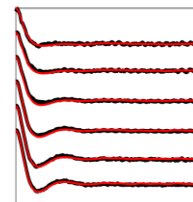
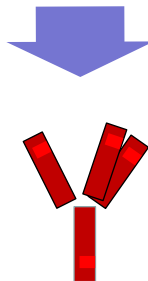
structure library
~15.000 conformers

2. Step: Simulation of PELDOR datasets for each conformer



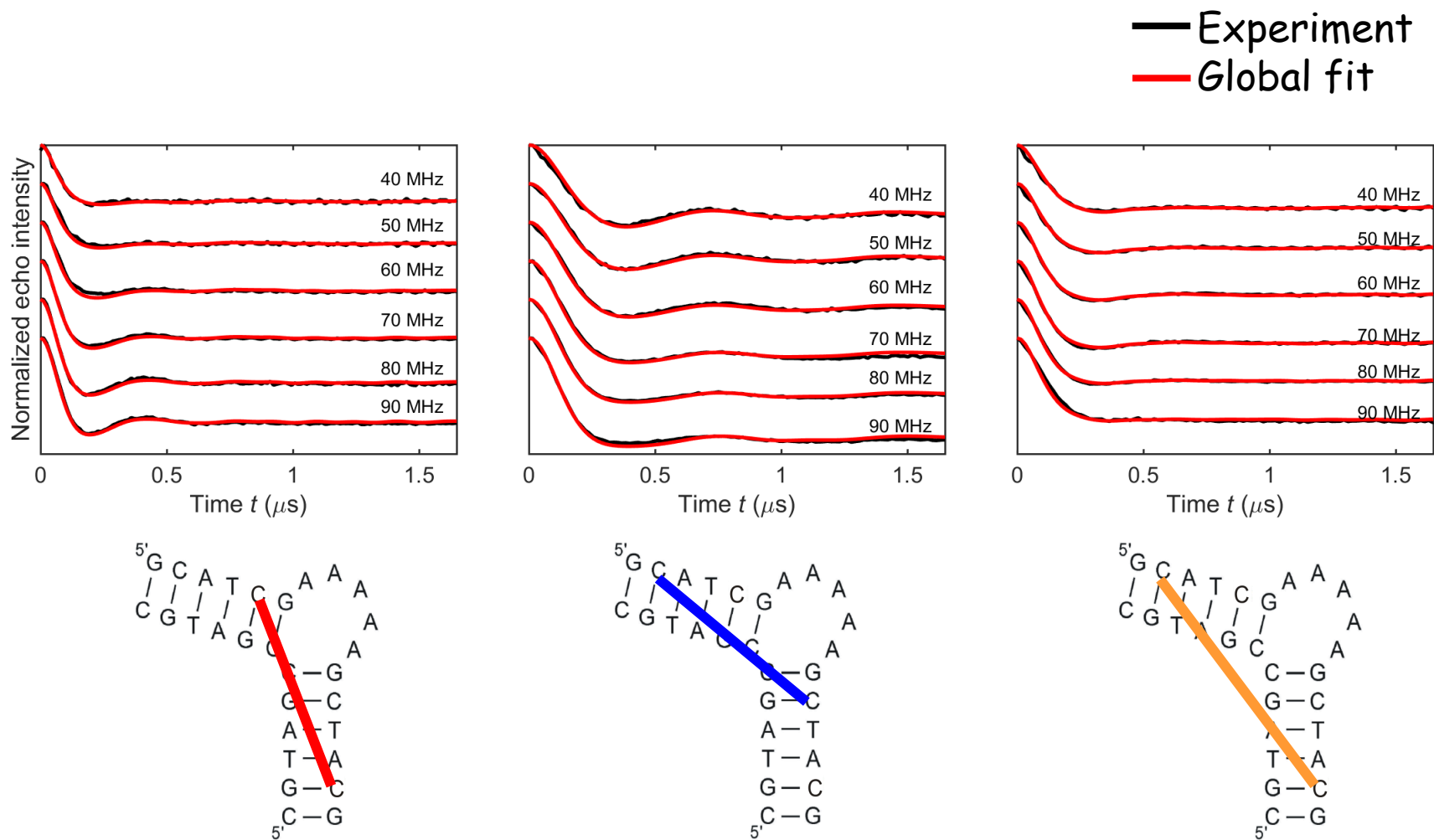
PELDOR
time trace library

3. Step: Iterative fit of experimental PELDOR dataset by PELDOR library



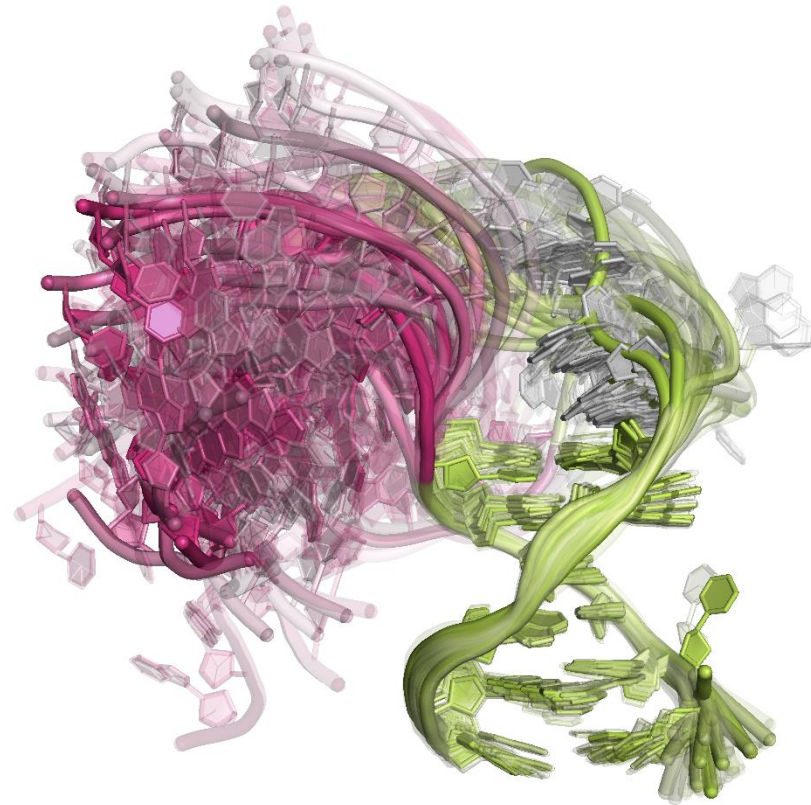
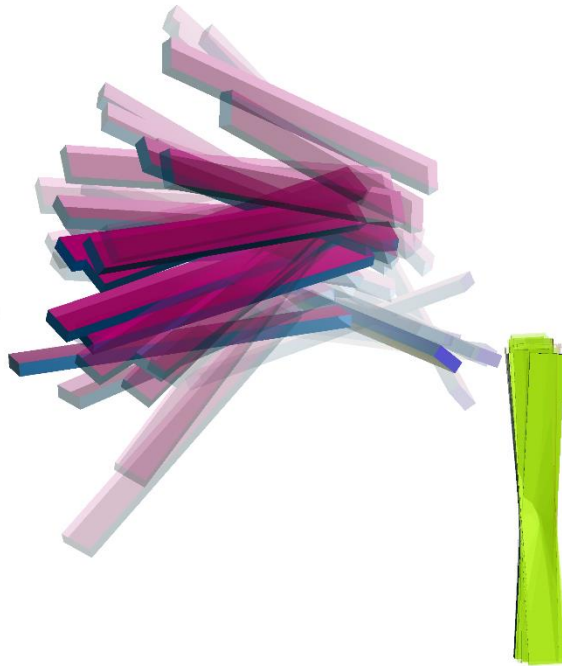
Conformational ensemble
in agreement with all
NMR and PELDOR restraints

Simultaneous Fit of 3 PELDOR datasets

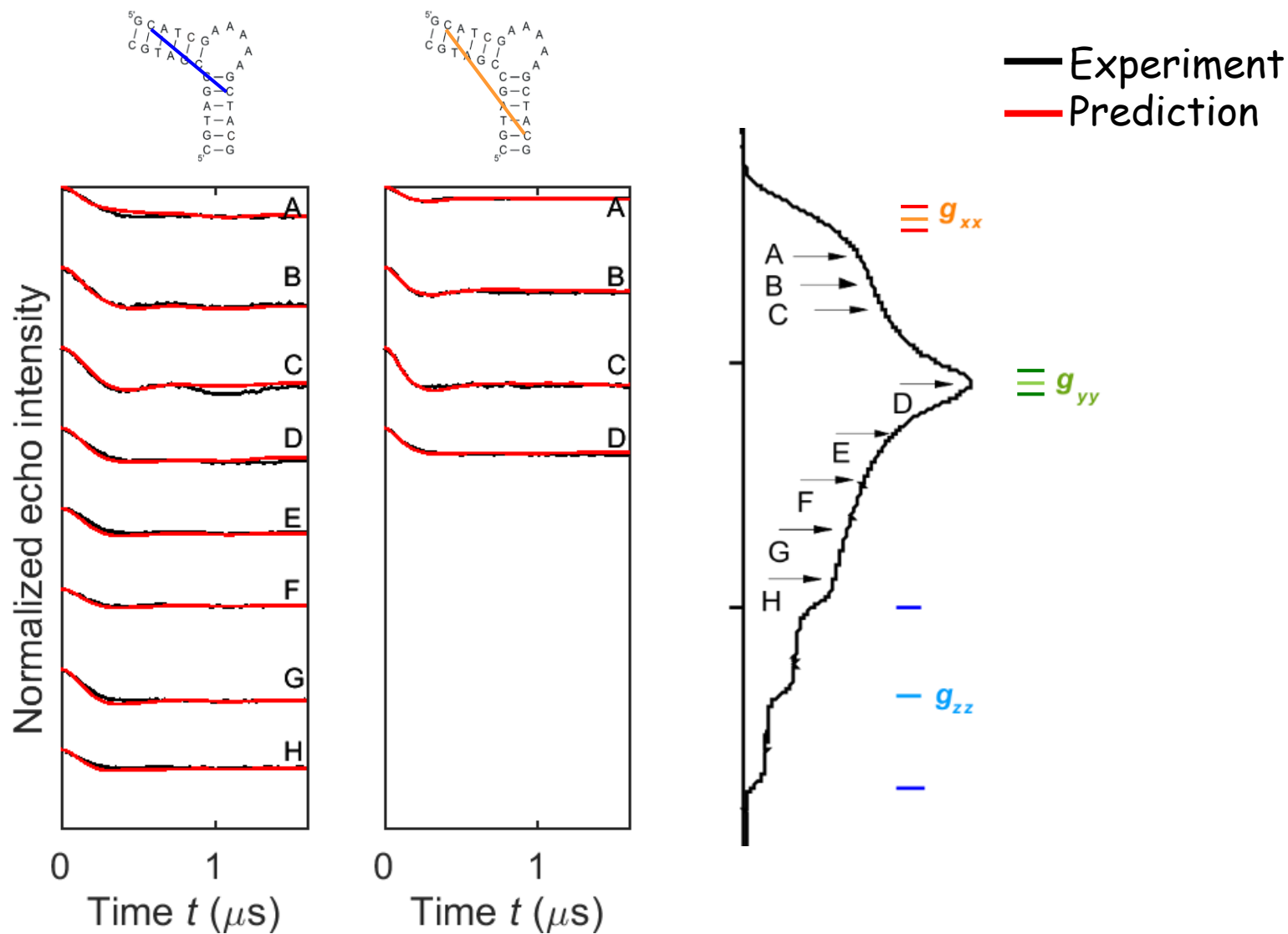


Conformational bundle describing bend DNA

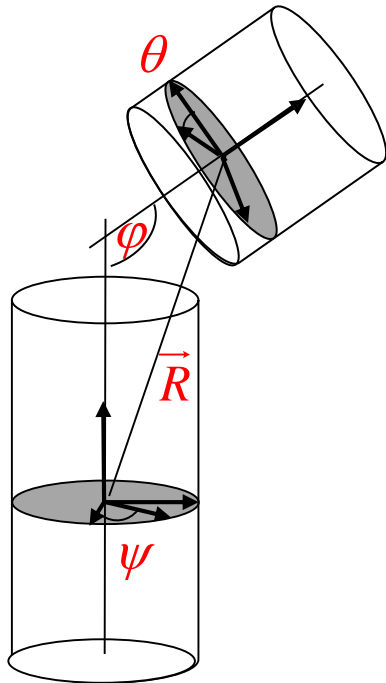
5' G C A T C G A A A
C G T A G C A A A
C G A T G C A
5' C G C T A C G



Evaluation of solution by G-band PELDOR data



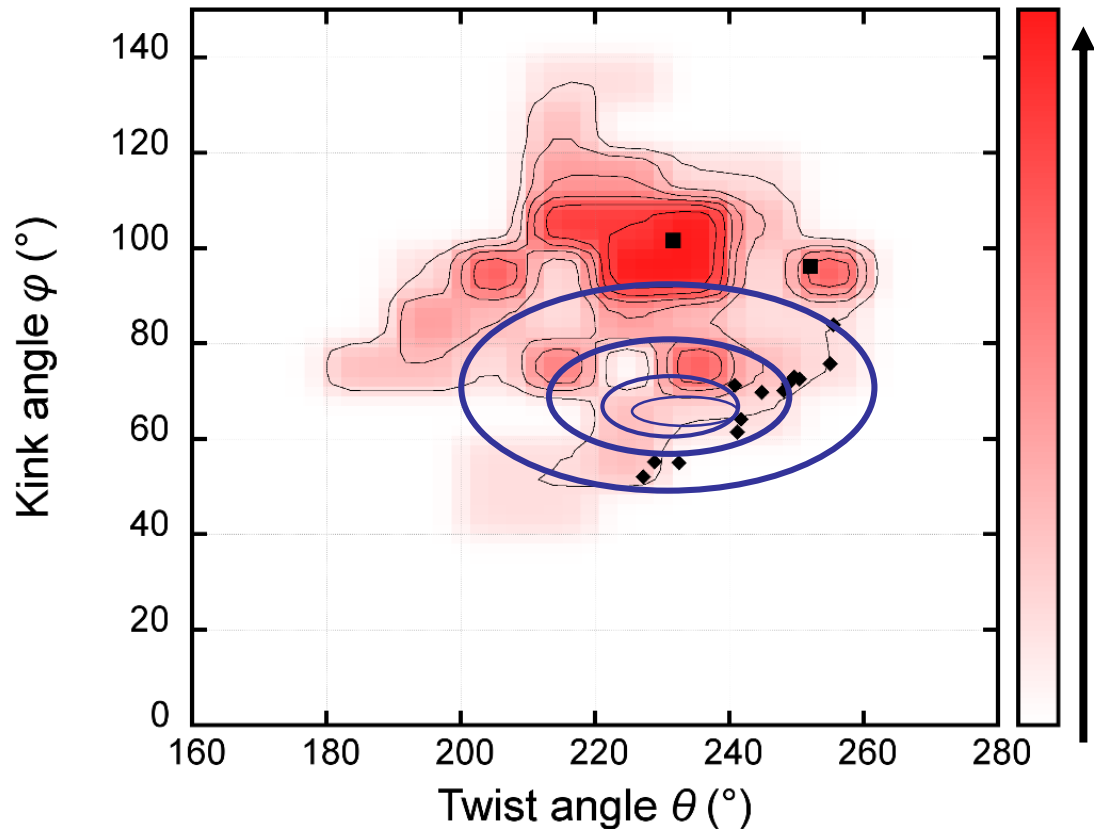
Comparison with NMR alone and FRET



EPR/NMR approach

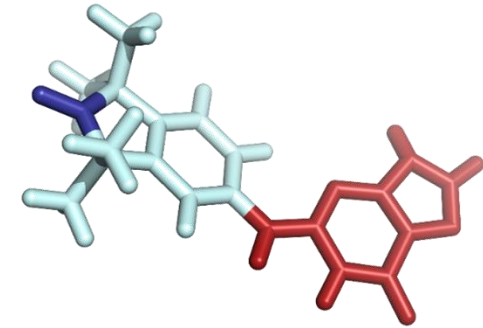
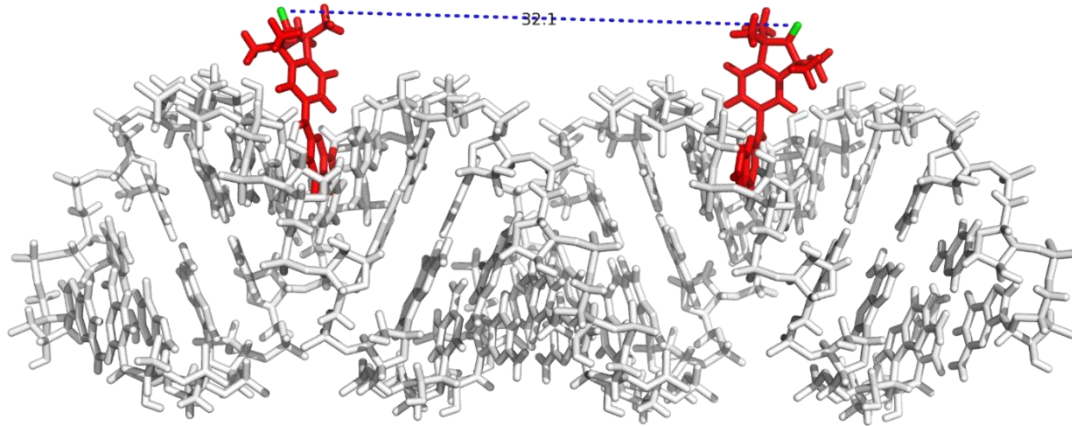
NMR

FRET



Non-covalent labels for nucleic acids

Collaboration with Sigurdsson
(Iceland)



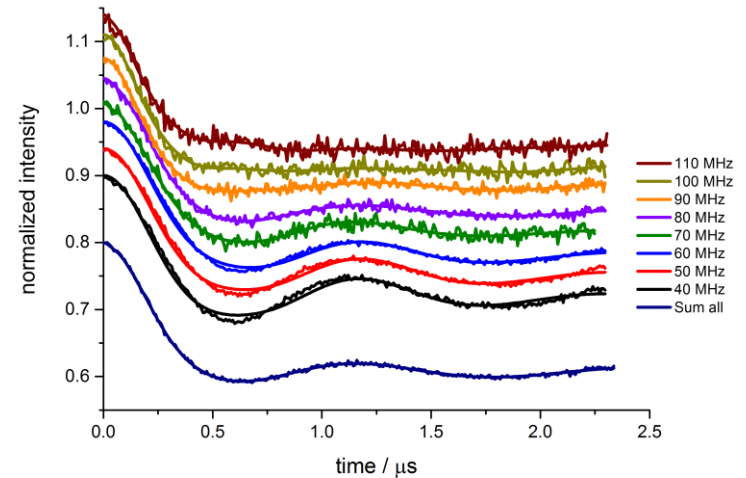
G Spin-label

5'-CGAC-GFA-UCG-CGC-GAU-CCG-UCG-3'

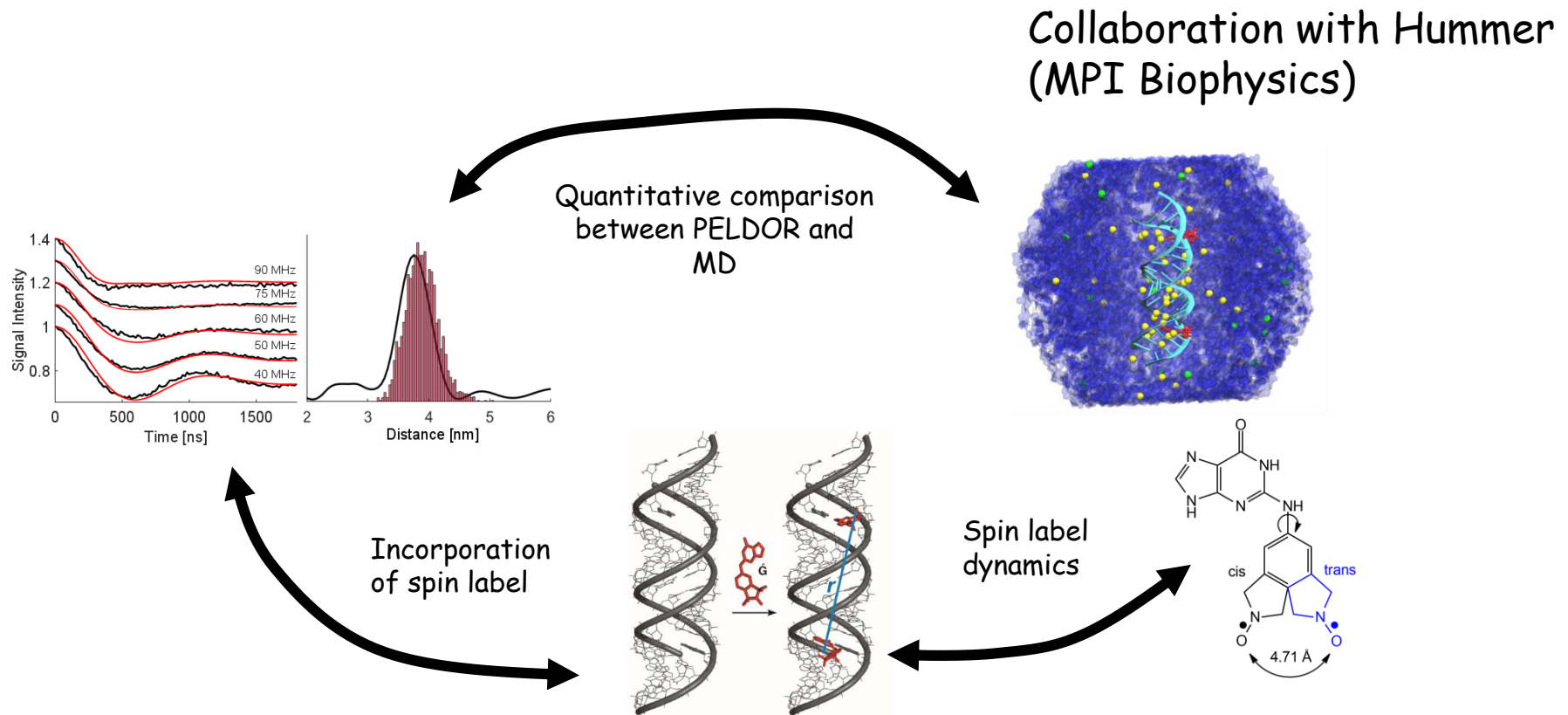
3'-GCUG-CCU-AGC-GCG-CUA-FGC-AGC-5'

High selectivity (abasic sites)
high binding affinity

Rather rigidly incorporated in
double stranded RNA

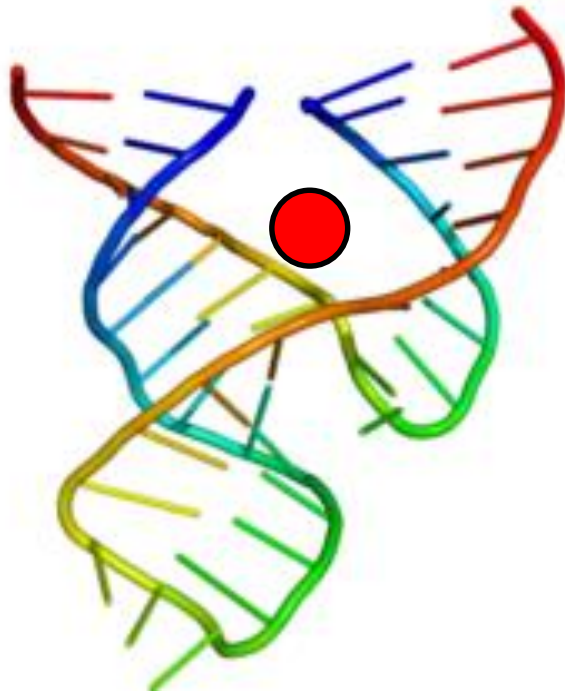


Combining MD simulations with PELDOR



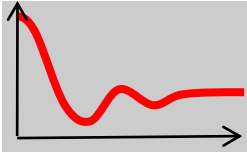
A synergistic approach to conformational dynamics and flexibility of RNA in the 1-10 nm range

Mg ion binding sites in RNA motives

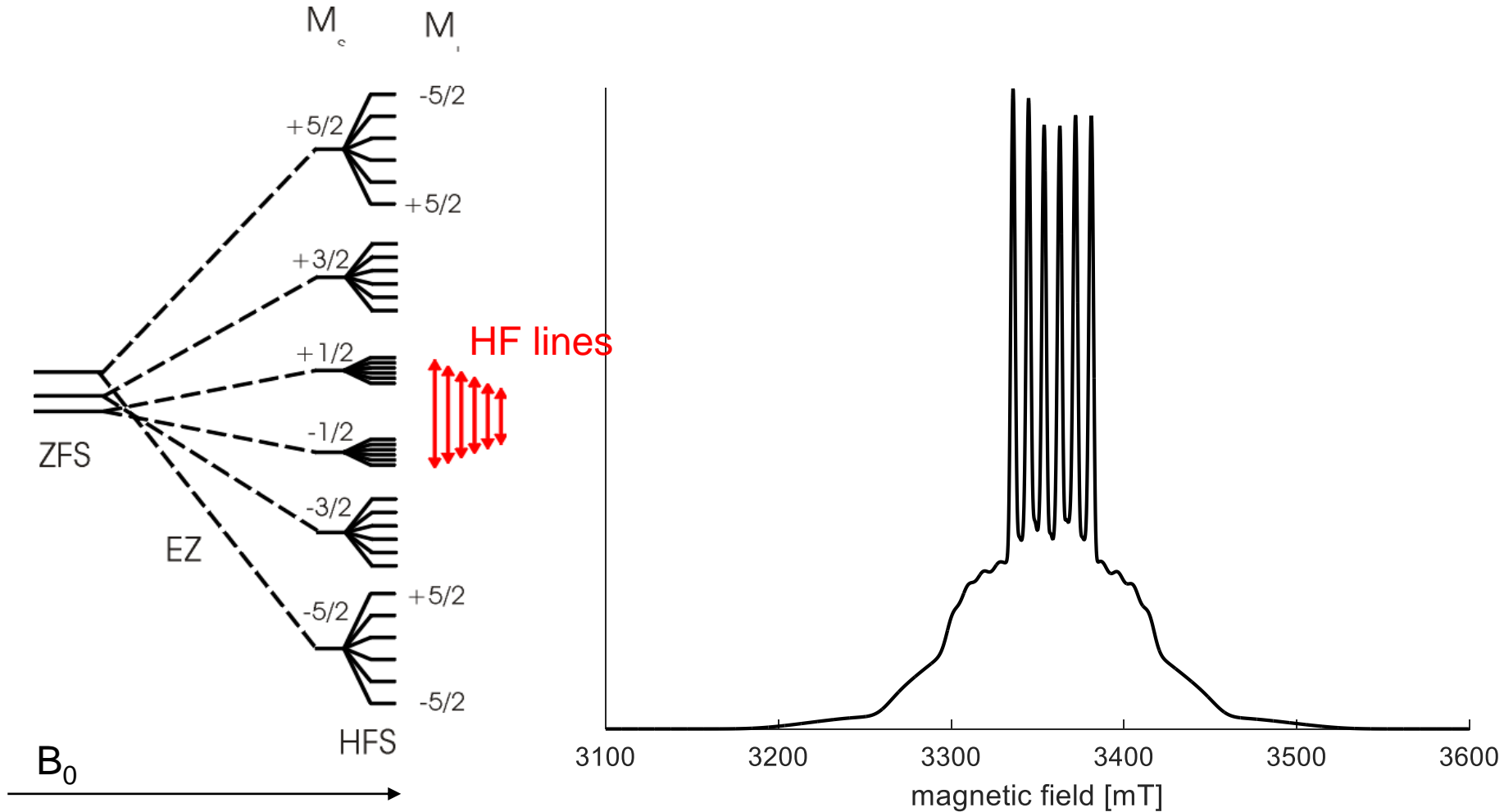


Important for tertiary structure
in
Ribozymes, riboswitches, ...

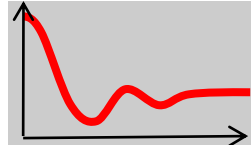
Replacement of
diamagnetic Mg^{2+}
by
paramagnetic Mn^{2+}



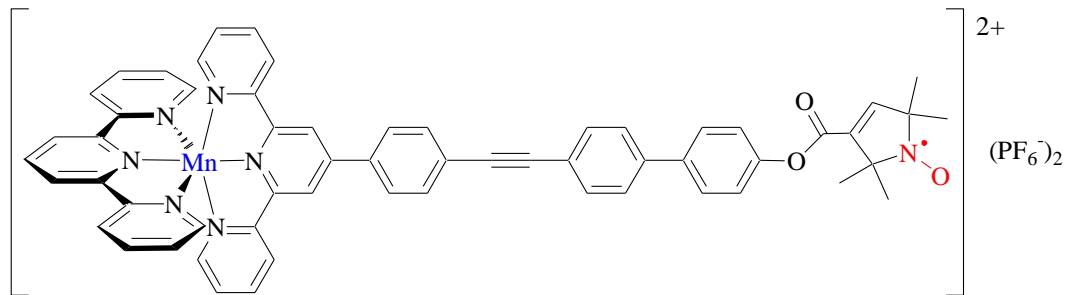
High spin Mn ion ($S=5/2, I=5/2$)



High magnetic fields ($\gg D$) necessary to obtain spin system easy to analyze
 Narrowing of central Mn transition at high fields (D^2/B_0)

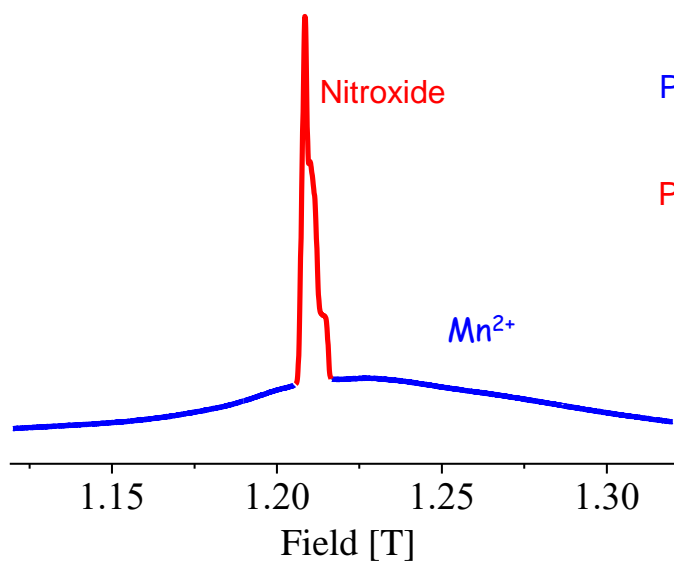


PELDOR on Mn^{2+} -Nitroxide Model-Compound



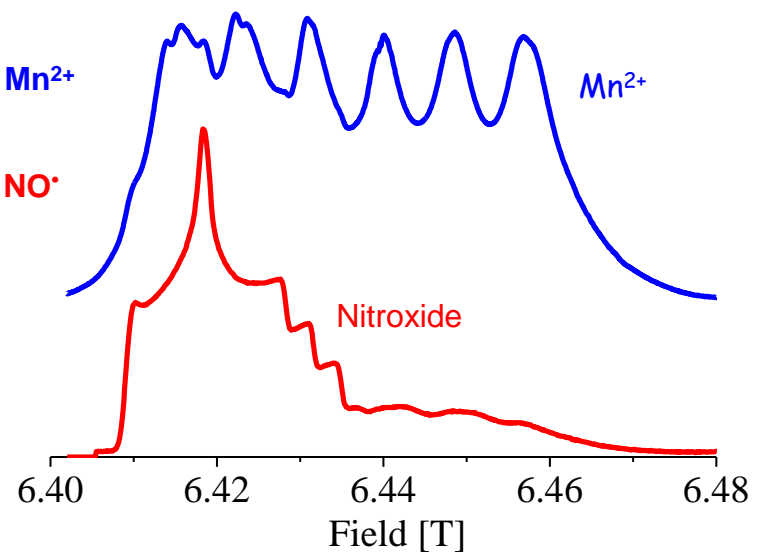
Mn-bis(terpy)-nitroxide with an estimated distance R (Mn^{2+} - $NO\cdot$) ≈ 2.7 nm

Q-band frequencies (34 GHz)

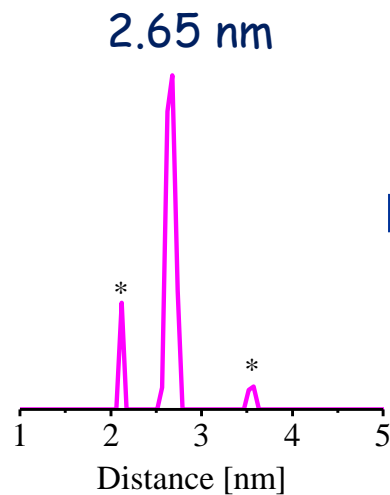
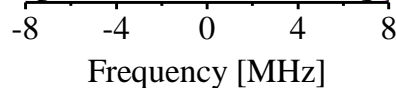
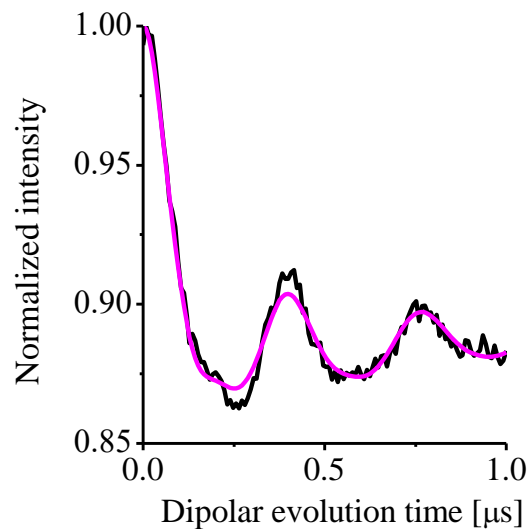
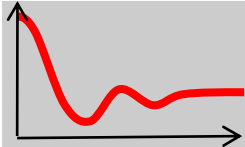


G-band frequencies (180 GHz)

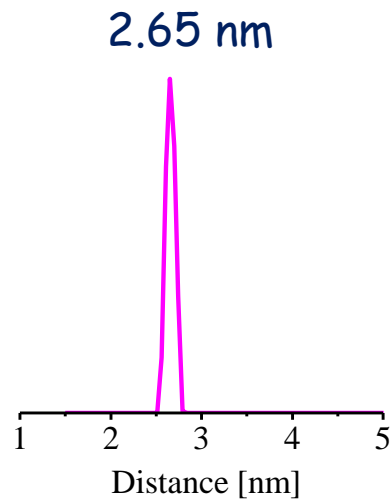
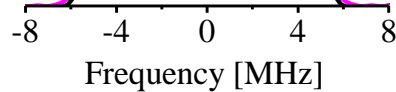
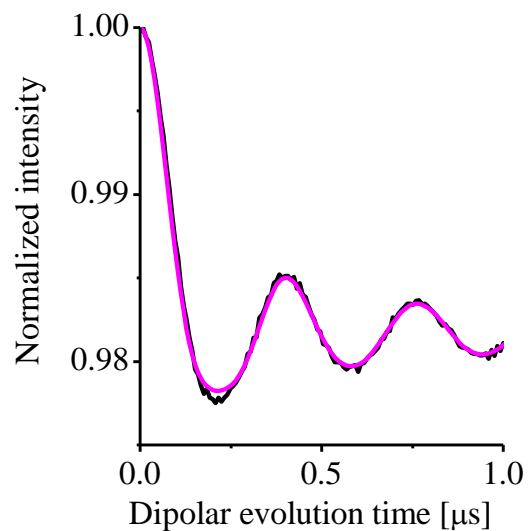
Pulses optimized for Mn^{2+} (15 / 30 ns)
 Pulses optimized for $NO\cdot$ (40/80 ns)



Q-band PELDOR on Model compound

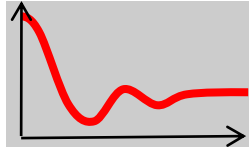


Pump on **NO•**
Detect on **Mn²⁺**



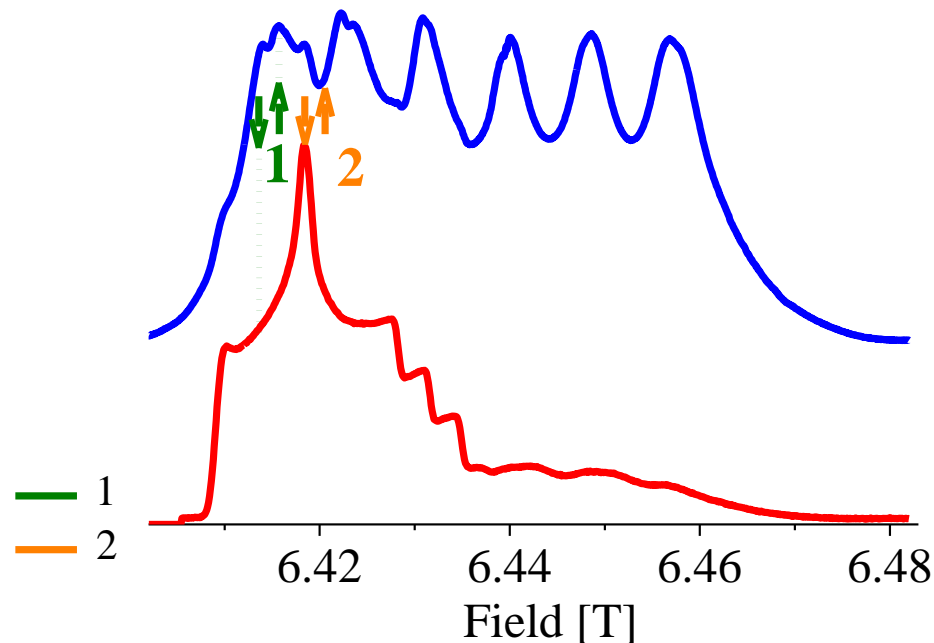
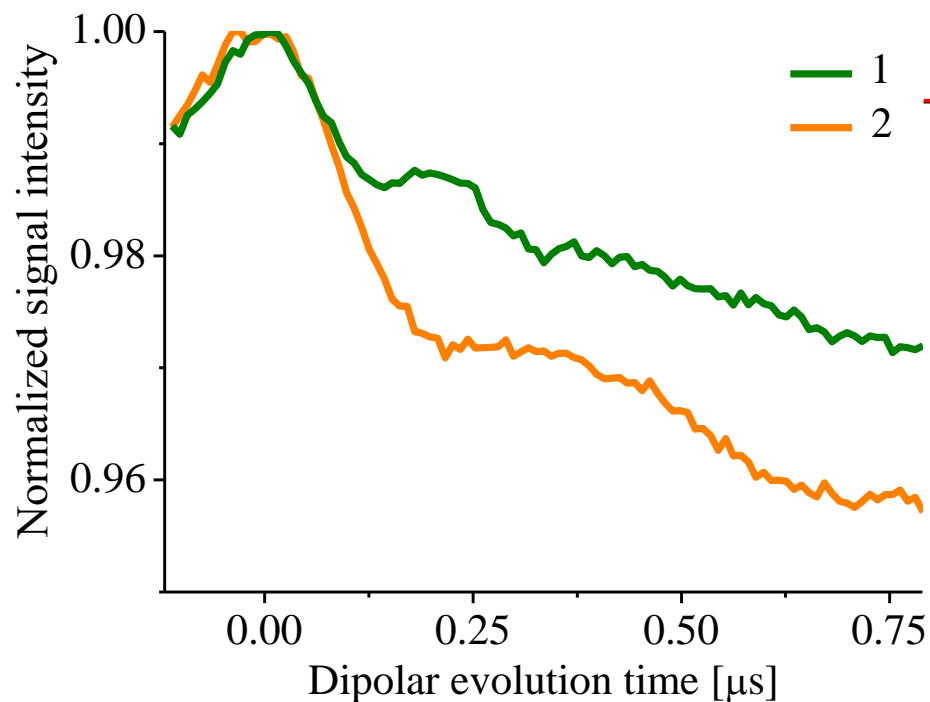
Pump on **Mn²⁺**
Detect on **NO•**

G-band PELDOR on Mn-bis(terpy)-nitroxide



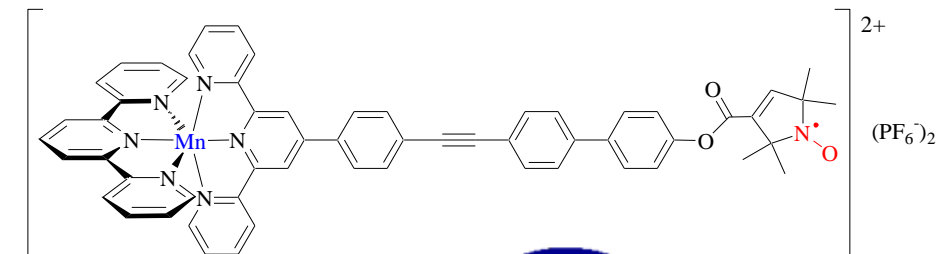
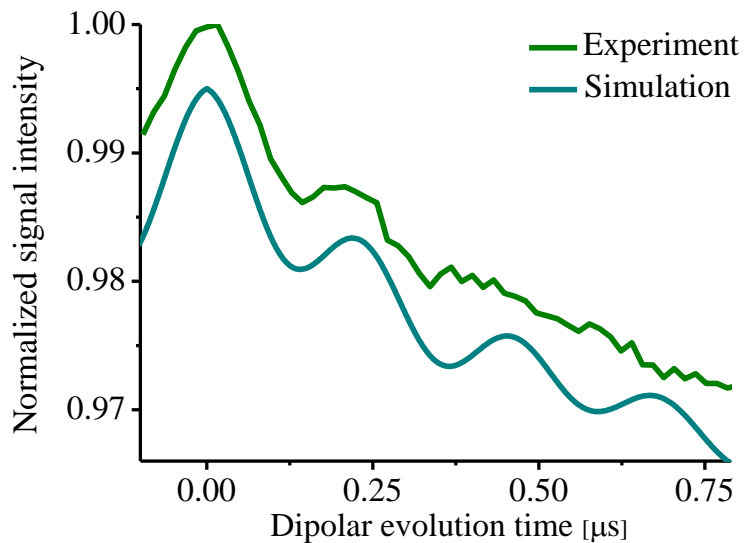
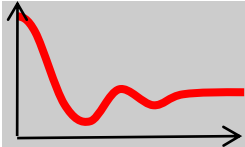
T = 10 K

Different dipolar oscillations observed depending on the pump/probe position (1 / 2)

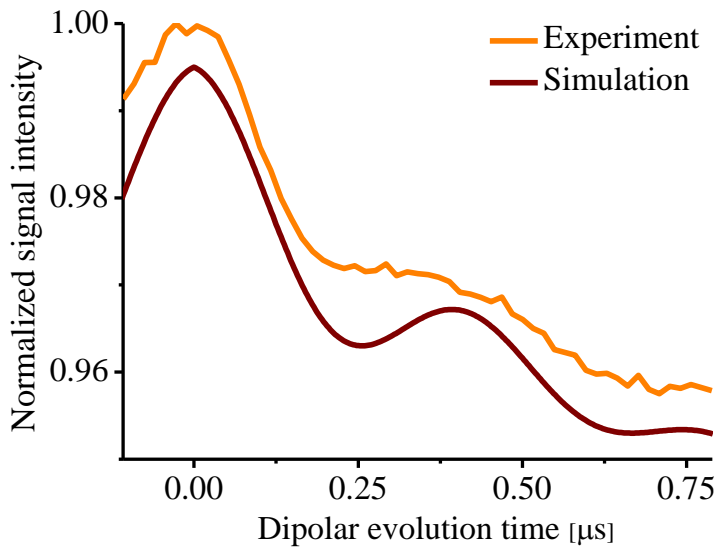
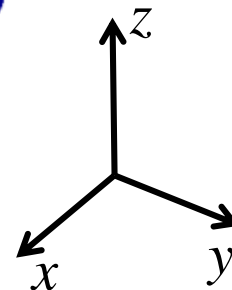
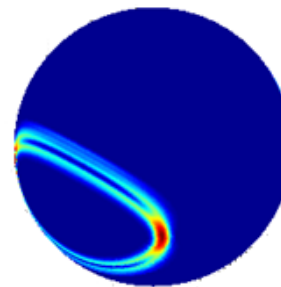


→ Orientation selection effects related to the anisotropic g tensor of the nitroxide radical at high magnetic fields

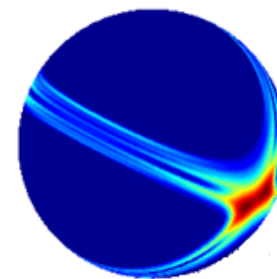
Simulation of orientation effects



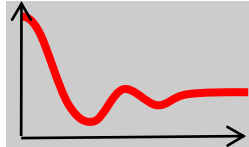
Position 1
(‘close to x’)



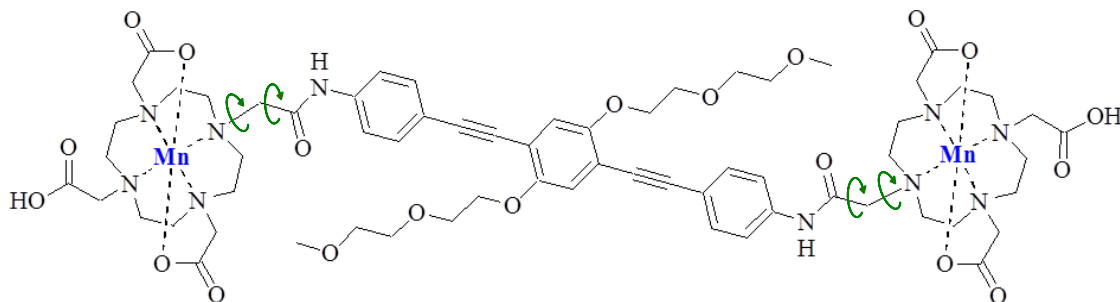
Position 2
(‘y’)



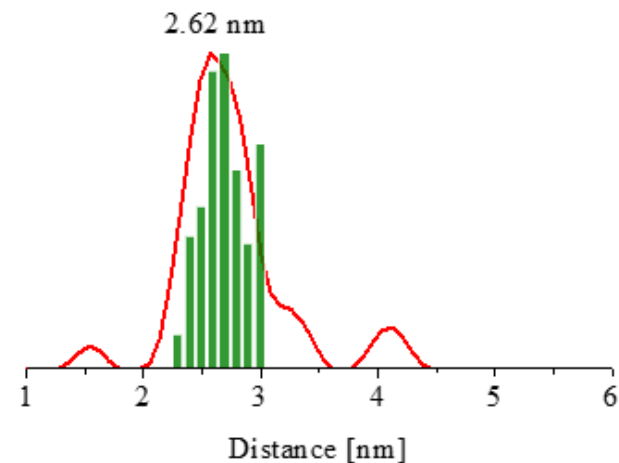
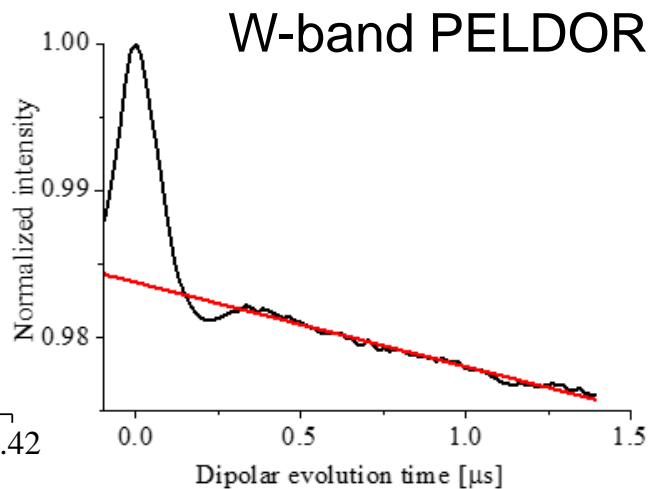
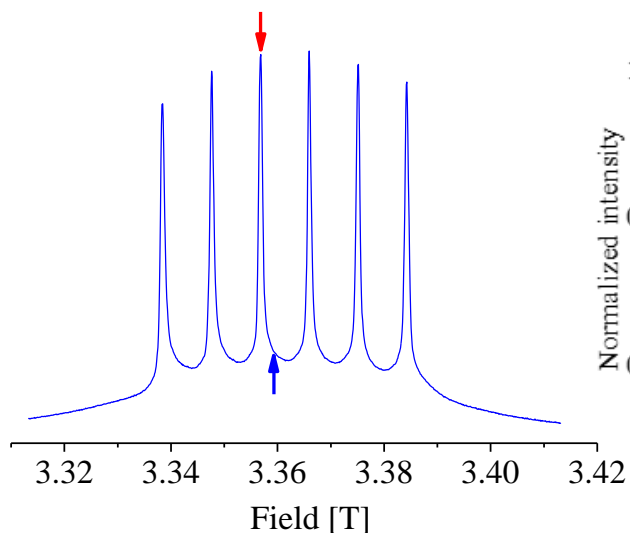
Excited orientations
of the **NO**



Bis-Mn-DOTA complex



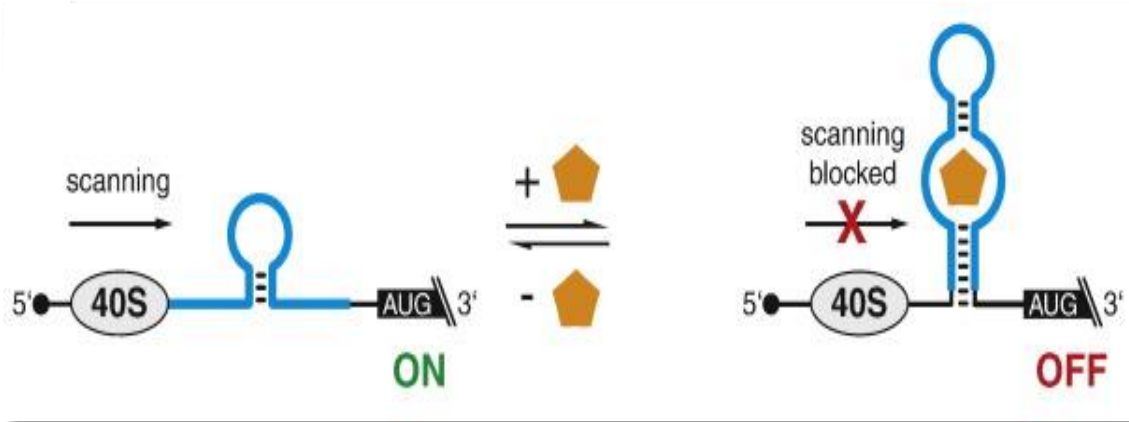
Collaboration with
C. Policar (ENS Paris)
S. Un (CEA Saclay)



Ching et al, *PCCP*, **2015**, 17, 6760.

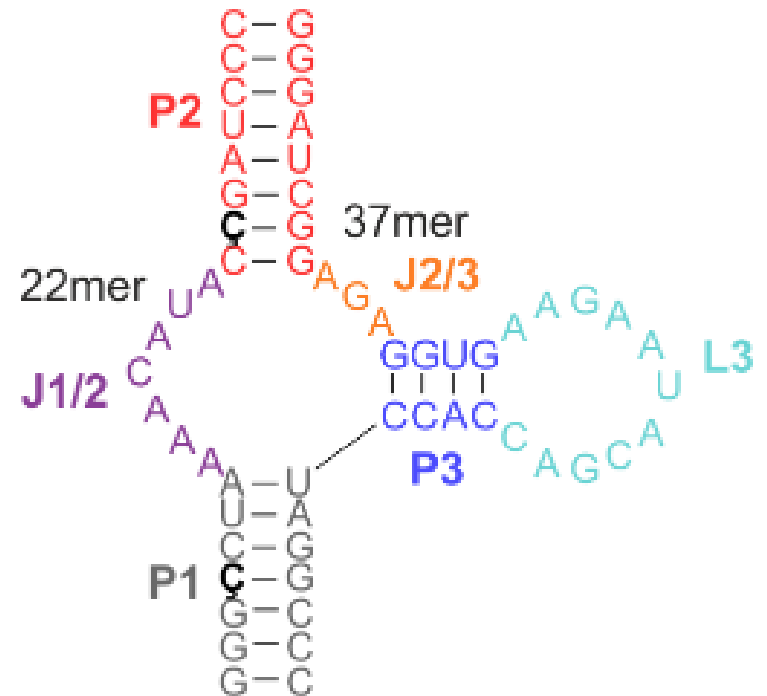
Demay-Drouhard et al, *ChemPhysChem*, **2016**

Tetracyclin-binding Aptamer



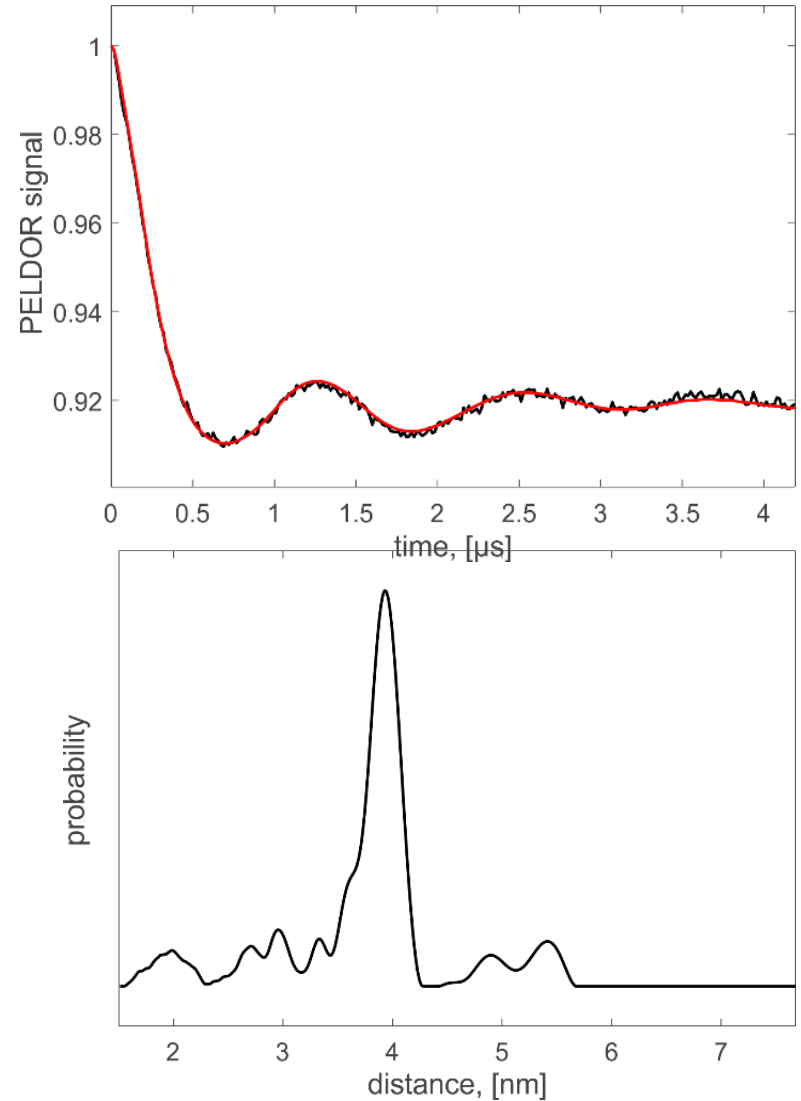
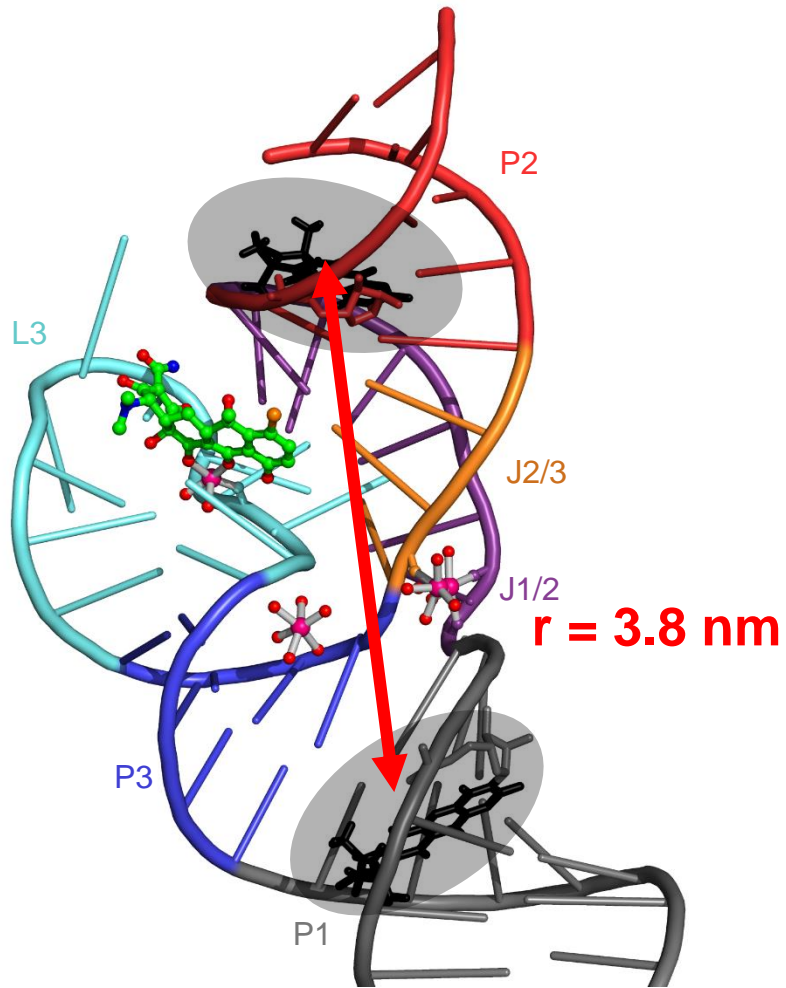
Groher & Suess BBA 2014

Binds TC with $K_d = 5 \text{ nM}$



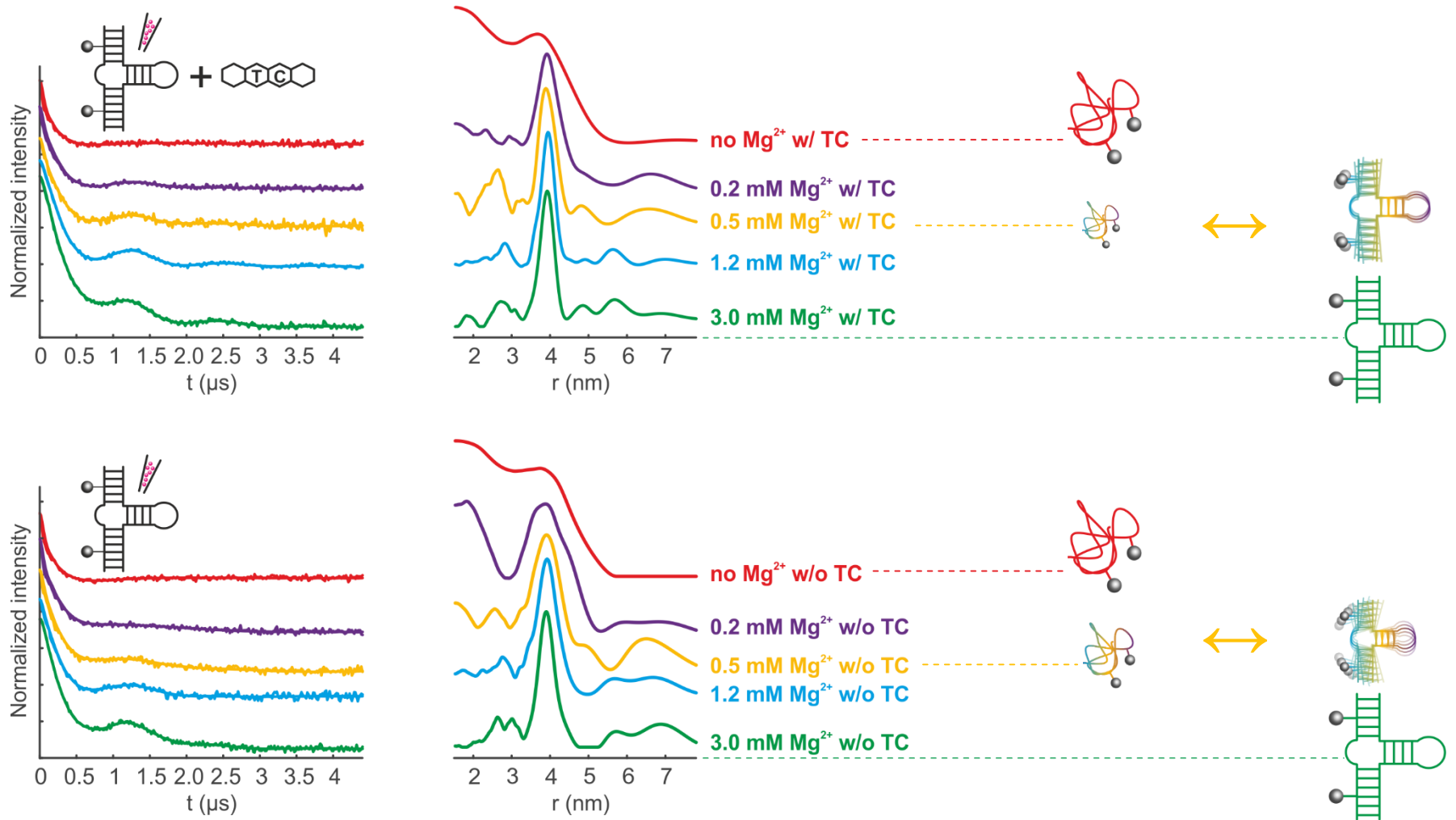
Application to Tetracyclin Aptamer

Xiao et al. Chem. Biol. 2008



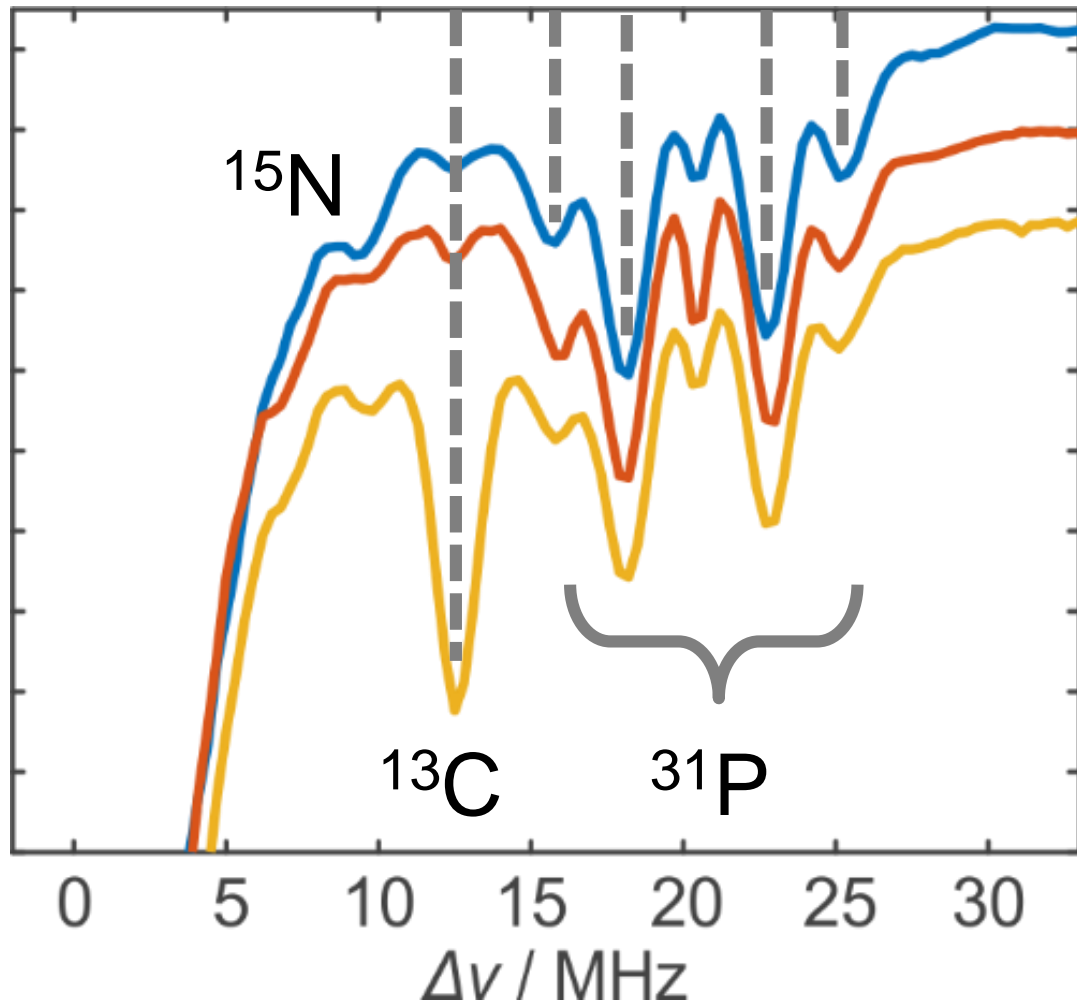
Collaboration with Beatrice Suess (TU Darmstadt)

Stabilization of tertiary structure by Mg^{2+}

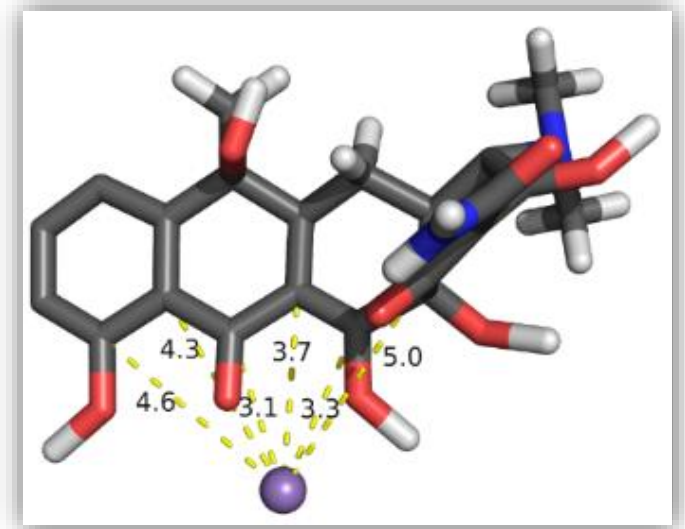


Tetracycline binding structure is already preformed

ELDOR detected NMR on TC aptamer



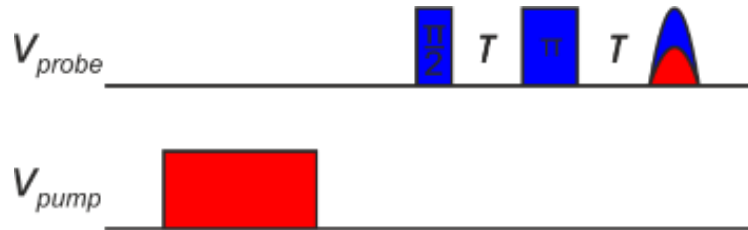
RNA/ Mn^{2+}
RNA/ Mn^{2+} /TC
RNA/ Mn^{2+} / ^{13}C -TC



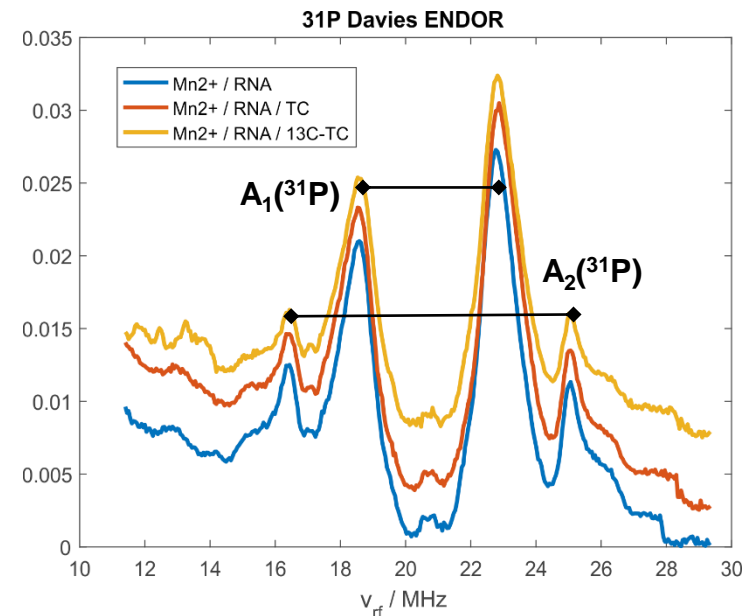
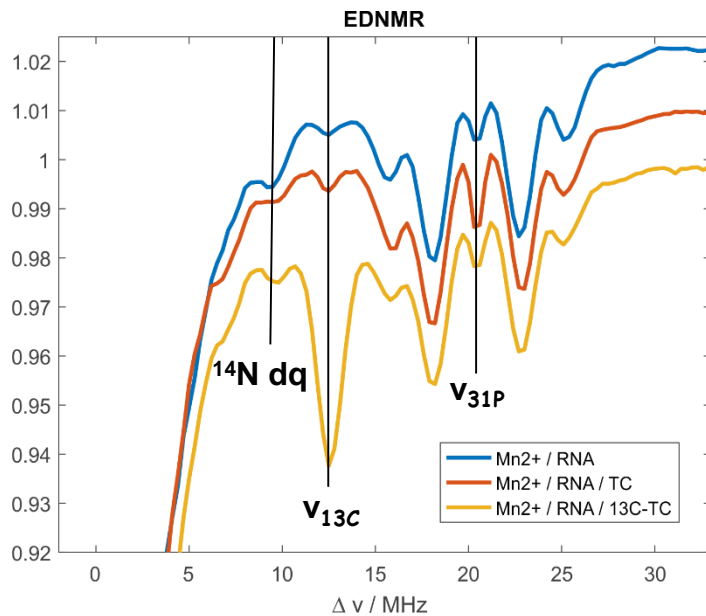
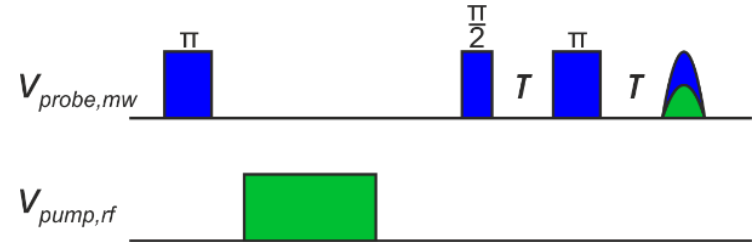
Mn binds to TC and to Aptamer

Comparison EDNMR & ENDOR

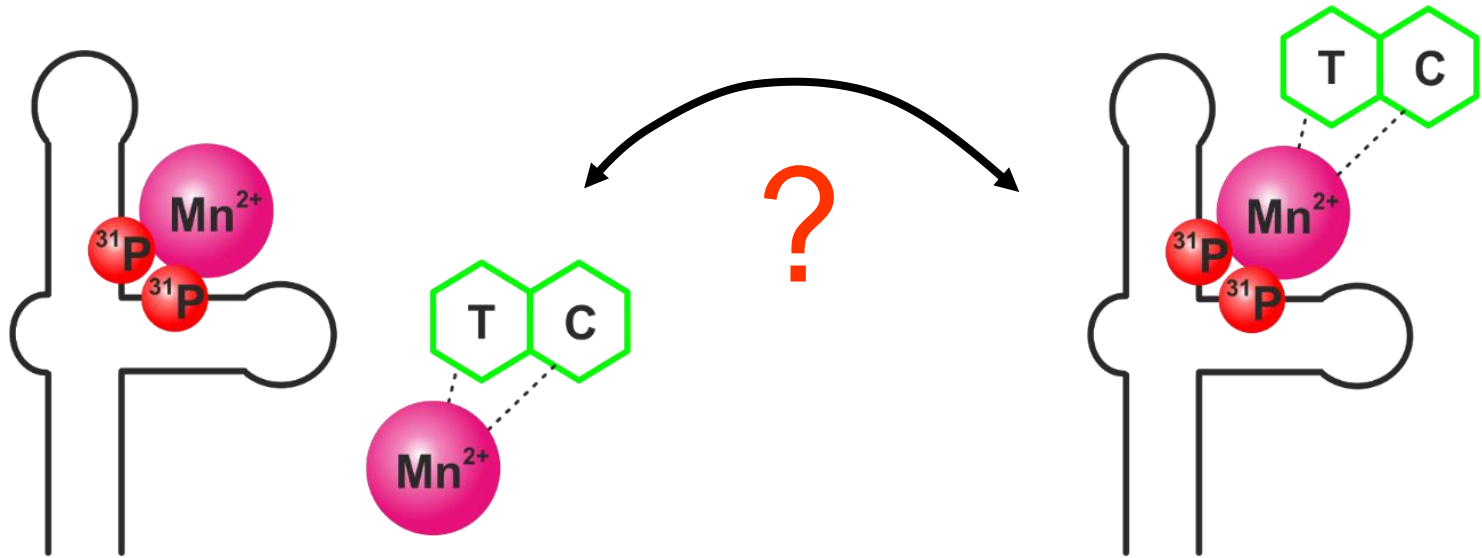
EDNMR



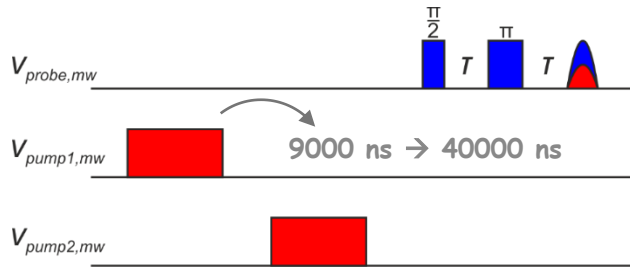
Davies-ENDOR



Comparison EDNMR & ENDOR

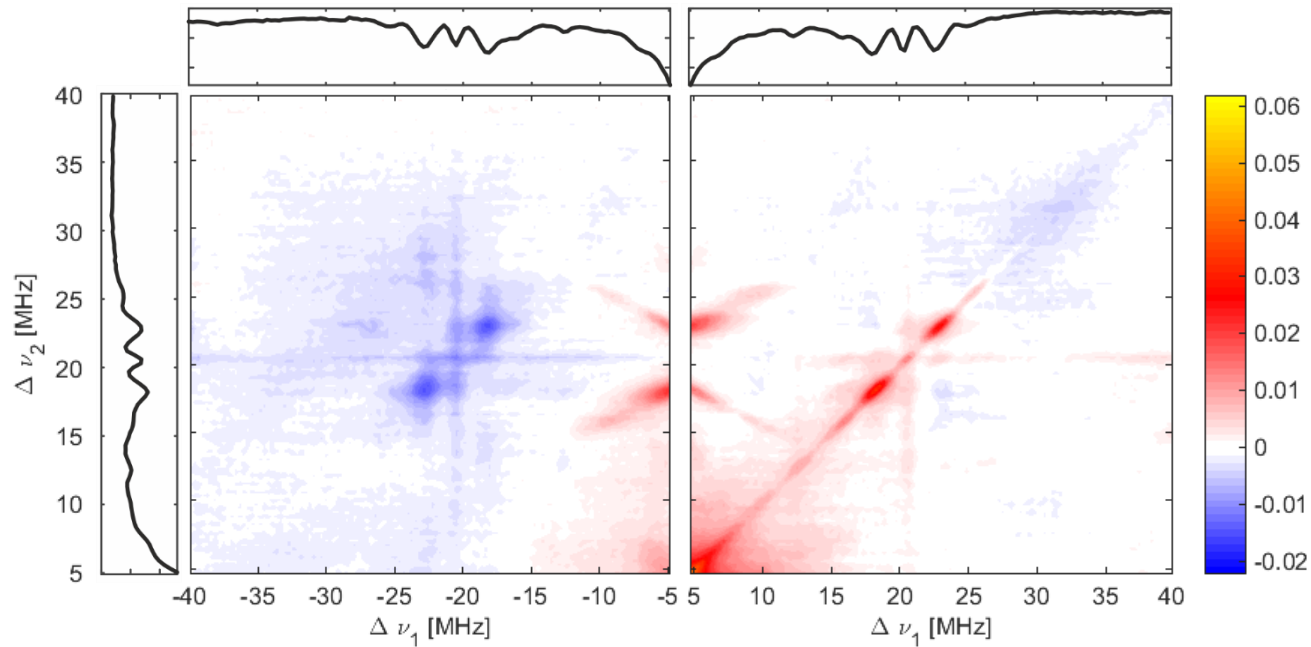


2D-EDNMR applied to TC aptamer

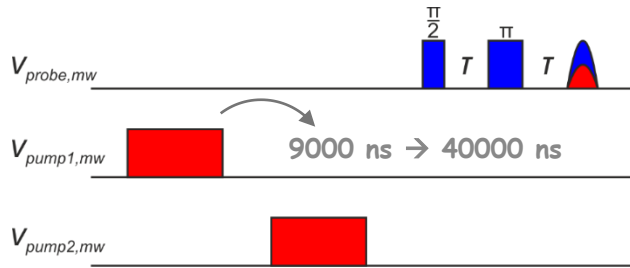


2D-EDNMR

Bg-corrected 2D-EDNMR 1mM RNA/Mn²⁺/TC

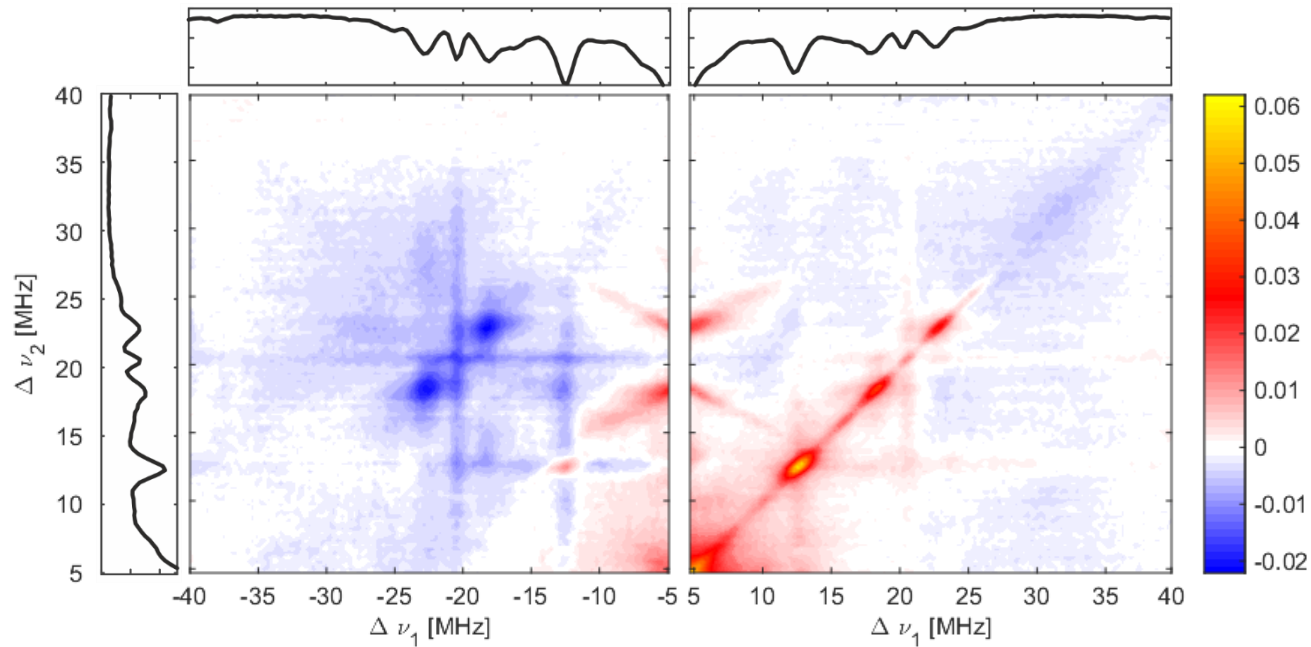


2D-EDNMR applied to TC aptamer

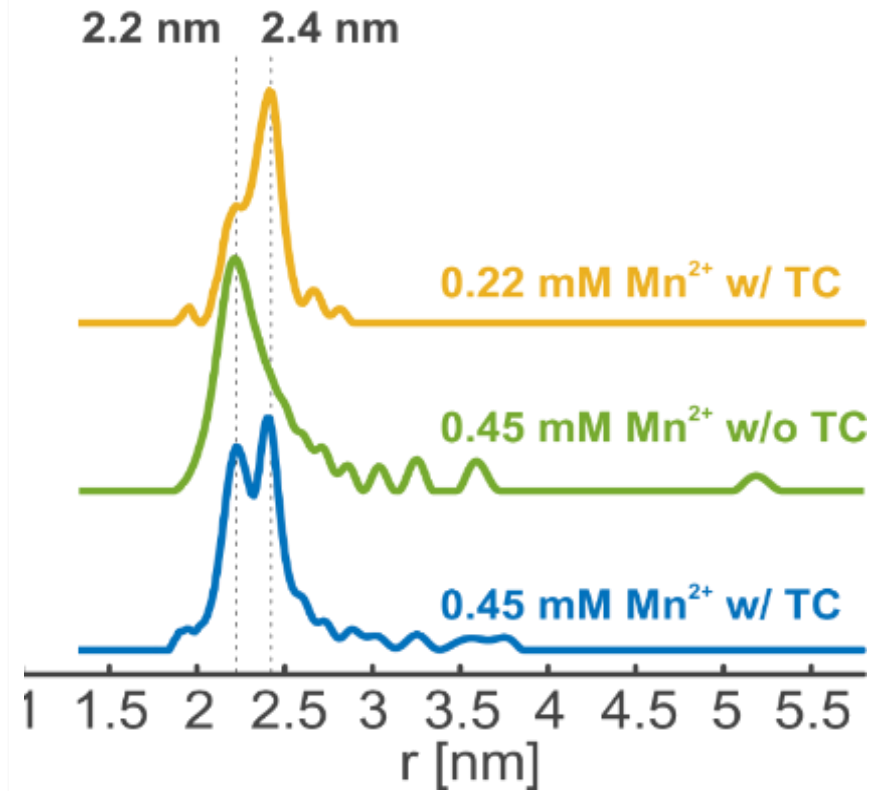
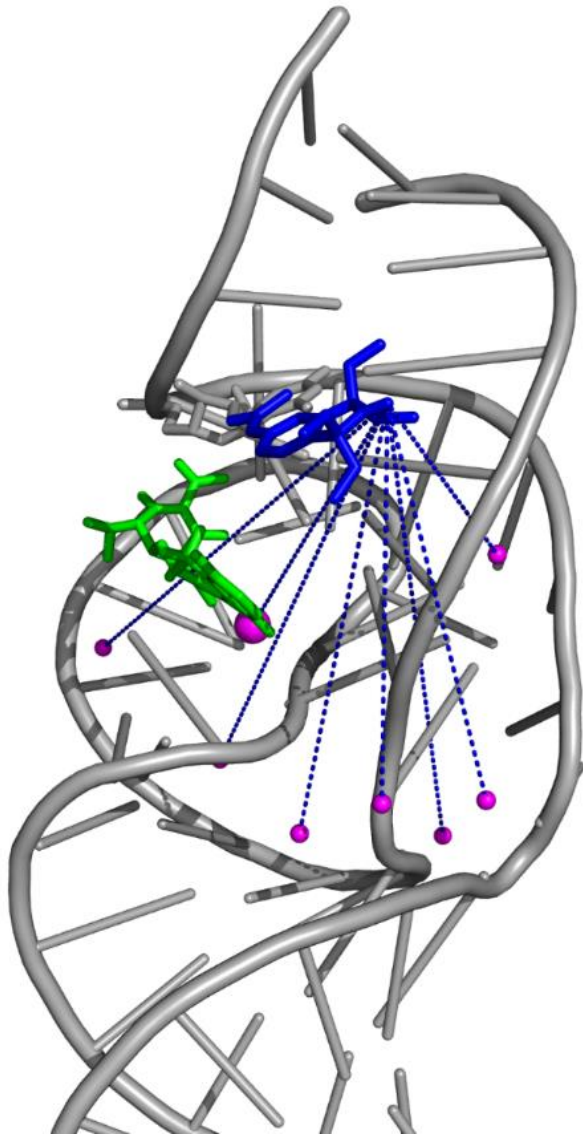


2D-EDNMR

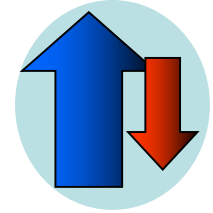
Bg-corrected 2D-EDNMR 1mM RNA/Mn²⁺/¹³C-TC



PELDOR from Cm spin label to Mn^{2+}



2 Mn binding sites can be detected



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